



and a substantial quantity of synthetic ones is from natural precursors [11]. Despite the success made with plant-derived NPs, huge hurdles and challenges remain exist. Hence, investigators and pharmaceutical industries need to invest time and effort to improve both the quality and quantity of nature-derived compounds [12], [13].

It is widely accepted that plant-derived natural compounds are harmless because of their less toxic nature in comparison with pure chemicals [1]. It is evident that bioactive principles compromise/nullify the toxic nature of the remaining additional components of a plant, however, the whole extract exhibits less toxicity with enormous potency [14]. In the last few decades, medicinal chemists have isolated many clinically important bioactive principles/phytocompounds from various traditional medicinal plants [2], [15], [16]. The progress of medicinal drug development from herbal plant sources faces numerous hurdles such as absorption, therapeutic efficiency, and poor compliance. Despite having the above problems, still crude herbs/plants are mostly formulated in various preparations. It is well known that the finding of new novel drug leads from plant sources is time-consuming, to overcome this, the adaptation of appropriate advanced tools for plant material collection, throughput screening assays, bioactive structure, and function elucidation required for drug leads development need the incorporation of multiple advanced techniques [17], [18], [19]. It is well established that the drug leads of plant origin involve simple stepwise preparation and therefore less expensive than synthetic drugs [20], [21], [22]. In this regard, by employing, various approaches such as combinatorial and synthetic chemistry and modeling approaches are highly employed to obtain novel drug and drug lead entities from plant sources [15], [23].

PDT is an emerging alternate mode of regulatory-approved site-specific cancer treatment and it requires the introduction of a harmless photosensitizing (PS) agent that accumulates selectively more in cancer cells [24], [25], [26], [27], [28] with minimal risk to healthy tissue than surrounding cells. In PDT, combining PS absorbs specified wavelengths of light, which in turn produces free radical molecules to eliminate neoplastic cells and tissues. PDT can be employed to alleviate skin diseases, target microbial infections [29], different malignancies [30], [31], and a variety of other complications in cardiovascular, ophthalmic, and immune dysfunctions [32], [33]. There are three well-established PDT-mediated biological mechanisms involved in the destruction of tumorous tissue: cellular, vascular, and immunological [34]. Several cellular organelles such as mitochondria and plasma membrane are known sites of photosensitizer location [34], [35], [36], [37]. However, not all the PS may not localize and bind to specific intracellular organelles or locations but rather bind to the diverse structures, which may lead to the involvement of various

death pathways in PDT [32], [38], [39]. In comparison to standard surgery options, the PDT approach is the non-invasive, precise target, repeated administration of regimen without total-dose limitations associated with radiotherapy, as a result, little or no scarring is found after healing [40]. However, for successful clinical outcomes, photosensitizers should meet certain requirements [41].

Although many studies in the last few decades have shown much interest in PDT for various applications based on its successful outcomes, despite its positive note, yet less interest toward plant-derived PS agents or bioactive principles, especially their phototoxic properties, which are very less tested to date. However, herbal research recently gained momentum to explore plants as sources for new phytotherapeutic agents [16], [19], [42], [43]. For years, a Food and Drug Administration (FDA) approved photoactive sensitizer such as hematoporphyrin derivative, namely Photofrin [44], was extensively employed for the clinical treatment of the vast array of cancers, which include bladder, breast [45], [46], and to kill various microbial organisms [43], [47]. However, it has some restrictions such as (i) its cutaneous tissue retention time for 4–10 weeks after uptake that leads to long-term skin photosensitivity, (ii) patients need to stay for a considerable length of time to avoid light, (iii) its insufficient low wavelength activation compromises tissue penetration, and finally, (iv) its badly defined molecular formula. Hence, the above limitations and drawbacks have encouraged the quest for new and novel sensitizers with ideal characteristic features suited to be adopted in the clinic for better outcomes [46], [48], [49].

In recent years, many established studies have enumerated various merits of ideal photosensitizers [26], [28], [50]. An ideal PS needs to be hydrophilic in nature for easy absorption, non-toxic till exposed to light, and activated by an appropriate wavelength by a tunable laser light source [51]. More importantly, a good PS should generate a good photodynamic outcome based on its cellular localization and selectivity [52], [53]. The following are some of the characteristic features such as (a) it should be a righteous unmixed chemical drug lead with selective absorption by the target, (b) induction of minimum dark effects (i.e., activated only by light irradiation), (c) high photo activity or quantum yield of free radicals species, (d) rapid clearance of PS to avoid irradiation mediated side effects, and finally, (e) stronger absorption in the long wavelength range between ~630 nm and ~800 nm [45], [46], [54], [55]. Based on the above characteristics, most of the currently available synthetic or natural photosensitizers have been identified and employed for various applications [46], [50], [56], [57].

From the available phytochemical literature, the PS metabolites were isolated from 35 families of plants, belonging to 15 various classes of phytochemicals [58]. These secondary metabolites are the products of four biosynthetic pathways such as shikimate, terpenoid,

fatty acid, and polyketide [59]. Previous studies have established that these light-activated isolates or extracts obtained from plants of various taxonomies comprised 44 families which include *Loganiaceae*, *Malpighiaceae*, *Papaveraceae*, *Phytolaccaceae*, *Piperaceae*, *Acanthaceae*, *Campanulaceae*, *Gesneriaceae*, and *Sapotaceae* [58], [60].

A spectrum of herbs-derived extracts have been isolated and tested for their chemotherapeutic properties but their PS ability for PDT is rarely examined [50], [61], [62], [63], [64]. To become a competent novel photosensitizer, it needs to meet and pass certain significant pre-requisite steps for PDT studies. In reality, till date, only a handful of FDA-approved PDT drugs in the clinic such as Photofrin®, Foscan®, and Levulan® to treat mainly skin, gynecological, gastrointestinal, and head and neck (H&N) type of cancers [65], [66]. Besides, in the last few years, quite a sizable quantity of both natural and synthetic compounds have been developed and screened both *in vitro* and *in vivo* as potential PS agents for PDT investigations [34], [56], [67], [68].

Yet, the hunt for new PS from potential natural sources is continuing by adopting crucial advanced scrutinizing steps. In this line, chlorophylls, porphyrins, furocoumarins, chlorins (Chlorin e6 [Ce6]), and a few other emerging PSs are of interest as they have exhibited superior therapeutic efficacy [69]. However, the new prospective PS should meet the eligible criteria to enhance target-specific actions for therapeutic efficiency in PDT and various other wider clinical applications.

It is conceivable that no current PS meets all the clinical requirements [68], [70]. Most of the PS have many disadvantages such as limited cell specificity or selectivity, skin sensitivity to prolonged irradiation, and unpredictable efficacy [41], [71], [72]. Other requirements include the following: (i) PS should be water soluble for intravenous injection, (ii) they should exhibit stronger absorption of light mostly in the near-infrared region, which is required for deep penetration into tissues, (iii) yield high quantum of singlet oxygen ( $O_2$ ) and produce less or no toxicity in the dark, and finally, fast cleared from the body. However, chlorins (Ce6), hypocrellins, hypericin (HY), and curcumin exhibit advantageous characteristics compared to other commonly employed PS. The following cellular organelles are prime target locations for photosensitizers, which include mitochondria, lysosomes, endoplasmic reticulum (ER), plasma membrane, and Golgi, etc., [34], [36], [73]. However, not all the current PS may not localize and bind to specific intracellular organelles or locations but rather bind to diverse structures, which may reflect the involvement of various death pathways in PDT [32], [38], [39], [74].

Many PSs such as chlorophylls, furocoumarins, chlorins including hypocrellins, HY, and curcumin have gained attention in recent years because of their

efficacy [50], [66]. Based on the available literature and efficacy, this review will focus on the chlorin type of natural photosensitizers and their counterparts because of their efficacious pharmacokinetic and photodynamic activities. Most compounds or molecules absorb light and acquire energy, subsequently losing the gained energy through radiationless loss/decay by an internal conversion mechanism. However, the PS molecule's internal conversion is not effective and not sufficient. Rather PS molecules transfer electrons to each other by transferring molecular  $O_2$ . In most cases, energy transfer occurs efficiently in their excited triplet states since this position allows an extended period for electron transfer. In fact, most of the current effective photosensitizers used in the clinics exhibit high quantum yields in their excited triplet state [75], [76].

In general, photosensitizers absorb light maximum at the far-red region, specifically at 668 nm, which falls within the optical window of biological tissues (600–800 nm range). Low-wavelength light fails to penetrate the tissue into deeper regions and produces no results. Similarly, very long wavelengths (800 nm and above) are also not useful as they have insufficient energy to excite tissue  $O_2$  to become singlet and then to generate a substantial yield of reactive oxygen species (ROS) [77]. Avoiding side effects, minimal or no destruction to nearby/surrounding healthy tissue but PS-specific localization to neoplastic lesions is an essential consideration for clinical PDT. It is known that most of the PS-produced ROS do not discriminate between cancer and non-cancerous tissues. Although the selectivity may not be achieved by any natural PS extracts, it can be achieved maximally by combinatorial chemistry or by employing tunable laser light as a source and precise delivery tools technique to target the tumor area/region [76], [77]. Although many clinically approved PS, only a few under clinical trials are currently in use to treat various types of cancer. Some of the synthetic counter partners of PS are listed below:

### **Photolon**

Photolon (1, 3, 5, 8-tetramethyl-4-ethyl-2-vinylchlorin 6 carbonic acetic-7-propionic acid sodium vapor salt), whereas, Ce6 hydrophile PS compound is linked with polyvinylpyrrolidone (PVP) in the ratio of 1:1 [78], [79]. Although, the chemical formula correlates with partially reduced porphyrin moiety, molecularly similar to Ce6, which separates pheophorbide exocyclic dimethyl amine  $\beta$ -ketoester by hydrolysis [80], [81]. The combination product of Photolon® by Ce6 and PVP exhibits better solubility and durability in water leading to superior bioavailability when compared to Ce6 alone [79], [82]. Unlike first-generation porphyrins, second-generation chlorins (Ce6 and derivative of Photolon®) demonstrated a higher ability to assemble in the neoplastic tissue but also cleared fast from the body and strongly absorbed in the red (between 640

and 700 nm) and thus qualify for the treatment deep sheeted and mass tumors [80], [81]. The i.v of Photolon improves the high uptake rate in target tissues which produces not only high tumorotropic but also produces less phototoxic reactions and is removed entirely after a period of PDT from the localized targets.

Previous reports have proven that Photolon has a better therapeutic outcome with increased wavelength to match its absorption peak because of its deeper penetration ability [54], [83] and proven that Photolon® prefers the following intracellular localization order such as: nucleus, mitochondria, lysosomes, and Golgi apparatus [32], [70]. A study by Ali-Seyed *et al.* [32] demonstrated that Photolon-PDT specifically induced apoptosis in CT-26 cells, this apoptotic cell death implies physiological correlates with minimal drug toxicity [84], [85].

Both natural PS Ce6 and its synthetic counterpart Photolon® share many qualities such as rapid accumulation of PS in the targets, faster clearance from the body, and producing a high quantum of singlet O<sub>2</sub> when compared to first-generation PS [86], [87]. Besides they are activated by near-infrared wavelengths (e.g., 664 nm), which enables molecules to enter into deeper layers of tissue [88], compared to 630-nm laser light used for photofrin or porphyrins [89]. Since Ce6 and Photolon® exhibit superior photophysical attribution for PDT such as higher molar absorption in the near-infrared spectrum and prolonged photoexcited triplet states, they are attractive PDT candidates when compared to other current porphyrin-based PDT agents. Previous studies have proven the local application of Photolon® as an ointment/patch form or by oral rinse with infrared irradiation at 665nm exhibits therapeutic potency in xenograft tumors and clinical trials [82], [90]. To support this claim, Ce6-based photosensitizers have recently received more attention due to their high photodynamic activity and therapeutic efficacy [85] against many types of tumors such as nasopharyngeal, melanoma, and bladder [32], [78], [91].

#### **Foscan/M-tetrahydroxyphenyl chlorine (mTHPC)**

Foscan (Biolitec Pharma Ltd., Dublin, Ireland/Germany) is a plant-based chlorine derivative is a PS consisting of temoporfin [92]. When compared to porphyrins, Foscan produces an increased amount of O<sub>2</sub> and exhibits high target specificity due to its hydrophobic nature leading to increased amalgamation of PS in target tissues [93], [94]. However, it is an industrial pure chemical and can produce quick and significant photodynamic reactions (PDR), but its treatment period is shorter in terms of seconds to avoid a longer hospitalization period as well as in the darkroom for more than a day because light exposure including normal room light is sufficient to activate this drug and produce significant severe burn (dark toxicity).

Foscan-PDT is highly effective, and this PS found a special place for primary and recurrent H&N cancer treatments [96]. The biggest disadvantage of Foscan-PDT is so painful to even under anesthesia for most patients who undergo Foscan illumination.

#### **Mono-L-aspartyl chlorin e6 (NPe6)**

This PS was promoted using various generic branded names such as MACE, LS11, and NPe6. This derivative is also called Fotolon (RUE Belmedpreparaty, Minsk, Republic of Belarus). NPe6 is herb-derived chlorine [95], [96] based effective PS bioactive principle to produce the PDR. It is important to note that, unlike Foscan, NPe6 does not cause dark toxicity after infusion for hours, which helps a longer treatment period [96], [97]. Moreover, NPe6 allows shorter periods of single-day infusion and therapy, which is highly acceptable both to patients and practitioners.

#### **Radachlorin**

Radachlorin® (Rada-Farma) and Photoditazine® (Veta-Grand) are hydrosoluble chlorines produced in Russia [98]. Normally Photoditazine® composition consists of Ce6 only whereas, Radachlorin® composition consists of Ce6 (90–95%), p6 (5–7%), and other unpublished components (1–5%). In the Russian Federation, both Radachlorin® and Photoditazine®-PDT are employed in various clinical applications for many types of malignant tumors such as bronchus, esophagus, melanoma, oral and colon, vulva, and various additional types of neoplasia [99], [100]. Radachlorin-PDT does not produce either local or systemic problems and produces good results without skin photosensitization. Radachlorin® also has a few disadvantages such as photo instability similar to other PSs such as porphyrins and phthalocyanines [101]. However, this PS can rapidly be degraded by laser light as it can be demonstrated by their decreasing absorption versus fluorescence intensity [100] when in simple solutions or complex environments.

Chlorins are promising PS agent for PDT when compared to other PS, owing to their absorption and emission spectra falling in the red to far-red wavelength range [102] as the 600–800 nm range of light penetrates deeper into the target tissues, whereas low concentration PS and low wavelength light or doses normally resulted into high phototoxicity [88]. Numerous numbers of FDA-approved chlorin-type PSs such as Talaporfin (LS11, Temoporfin [Foscan, mTHPC, 5,10,15,20-Tetra(m-hydroxyphenyl) chlorin], N-aspartyl Chlorin e6, NPe6), Photolon®, radachlorin (a mixture of three chlorins), MACE, and Photodithazine (glucosamine salt of Ce6) [103], [104] are employed in various clinical applications, which include the treatment of many types of cancer. Most of the chlorines including bacteriochlorins fall in the bandwidth of red

and near-infrared permitting deeper tissue penetration for light, therefore qualifying these PS ideal candidates for PDT of neoplastic tissues [98], [105], [106].

Despite many investigations have outlined the PDT outcome of many synthetic drugs, little or inadequate attention has been paid to herbal plant extracts or plant-derived bioactive principles. Herbal extracts from curative plants are considered safer when compared to synthetic counterparts. Hence, it is appropriate to review some of the socioeconomically important medicinal plants derived from PS phytochemicals and their utilization in PDT to treat various diseases including cancer. In recent years, PDT employed plant-based photosensitive drug leads receiving paramount importance as a green approach in PDT. Besides, this review discusses their possible molecular mechanism to place them in their rightful prospective scientific discipline which will determine the scope of green PDT for the treatment of a wide range of human chronic diseases.

### Chlorins

Photosensitivity and poor absorption of tissue-penetrating low-wavelength PS have led to the search for new ones with many novel characteristics, specifically the sensitizing agents absorbing longer wavelength light for deeper penetration and faster clearance from the normal tissues [106]. Chlorin type of second-generation PS agents has some superior qualities like shorter photosensitization span, red to near-infrared absorption bandwidth and yield higher quantum of O<sub>2</sub>, and target selectivity [107]. The chlorine type of PS exhibits photophysical properties similar to the porphyrin type of macrocycles. For example, bacteriochlorins PS falls in the bandwidth of red and near-infrared permitting deeper tissue penetration for light, therefore qualifying chlorin-type PS is ideal candidate and gaining interest as the preferred choice for photodynamic diagnosis and PDT of neoplastic tissues in the clinic [98], [108].

Chlorins are the main type of nature-derived PS agents originating from chlorophyll a. It has a longer spectrum of absorption wavelength, improved target selectivity, and elicits meager photosensitive reactions. The longer spectra of wavelength between 650 and 700 nm aid deep penetration of targets. These novel characteristics motivate investigators in the field to look for more plant-derived extracts to elucidate the structure and function of many new prospective herb-derived PS [69]. It is important to note that such as porphyrin-type derivatives, chlorin PS can also exhibit two additional hydrogen atoms combined with peripheral pyrrole double bonds. This nature-derived chlorophyll displays strong absorption with appropriate O<sub>2</sub> production with lack of water solubility and instability are the only drawbacks, however, much effort is underway to adopt advanced purification and combinatorial chemistry techniques to attract more attention for various pharmaceutical applications [109].

### Ce6

Ce6 is a nature-derived PS agent and a member of the chlorin family. It is normally obtained from *Spirulina chlorophyll* (*Chlorella ellipsoidea*) and other green plants [77]. Ce6 is lipophilic in nature and exhibits an asymmetric structure with three ionizable carboxylic groups in it but pH dependent [110], [111]. Ce6, (17S,18S)-18-(2-carboxyethyl)-20-(carboxymethyl)-12-ethenyl-7-ethyl-3,8,13,17-tetramethyl-17,18,22,23-tetrahydroporphyrin-2-carboxylic acid, its molecular structure, is C<sub>34</sub>H<sub>36</sub>N<sub>4</sub>O<sub>6</sub> with a molecular weight of 596.67. Ce6 is one of the interesting classes of tetrapyrrole compounds based on their origination and photophysical properties [112]. It is well established that tetrapyrrole backbones are present in numerous biomolecules such as chlorophyll, bacteriochlorophyll, and heme and they are collectively called “pigments of life” [112]. In general, tetrapyrrole type of PS (except bacteriochlorins) tend to generate predominantly Type II (O<sub>2</sub>) and Type I (OH) free radical species which are normally generated by PS with other structures. Although many tetrapyrrole PS agents have been employed for various PDT applications, only a few of them have exhibited their superior actions in the clinic as well as in clinical trials [32], [82]. Ce6 is highly preferred for PDT applications not only for its longer and deeper penetration of red spectra region [113] but also for its low-cost making when compared with other porphyrin-type agents and exhibiting long lifetime in their photoexcited triplet state [114]. It is evident that longer wavelength laser light always penetrates deeper than the lower one (633 nm) commonly used for Photofrin by high sensitizing efficacy and rapid elimination from the body [115] together with the higher values of interconversion coefficient (and, consequently yields high quantum of O<sub>2</sub> when compare to porphyrins (610–620 nm).

### Hypericum

HY is a polycyclic phenanthrenedione biosynthesized by the herb *Hypericum perforatum* L belongs to the genus *Hypericum* [116]. HY is well known as St. John's wort and is the best-described prominent representative from this genus. HY is a new class of novel PS agents exhibiting superior characteristics both *in vitro* and *in vivo* studies when compared to several other photosensitizers currently in usage and investigated [117], [118], [119], [120]. HY binds mainly to the cell membrane of multiple subcellular organelles [121], [122], [123] and metabolizes quickly *in vivo* investigations without exhibiting toxic reactions [124], [125]. It is well established that HY-PDT has shown its potential outcomes against a variety of cancer types [123], [126], [127], microbes including viruses [128], [129]. Besides, HY photosensitizer elicits superior O<sub>2</sub> production and a high quantum yield of fluorescence upon light irradiation [130]. The above

facts strongly support HY as a potential PS for further development and its clinical use for multiple human chronic diseases.

Besides, a secondary metabolite of *Hypericum* HY is also present in other *Hypericum* species [131], [132] and in basidiomycetes (*Dermocybe* spp.) [133], [134] orendophytic fungus grows in *H. perforatum* (*Thielavia subthermophila*) [135], [136], [137]. Since HY is a bioactive PS, it is applied for various clinical applications other than targeting cancer, moreover, its efficacious potentials have been assessed using various cultures of *H. perforatum* and their transgenic clones [138], [139]. Besides, *Hypericum* cultures were tested for various biotechnological applications in terms of their preservation or producing various secondary metabolites or bioactive drug leads [140], [141].

HY exhibits two broad peaks such as white light (500–600 nm) and ultraviolet (UV) light (300–400 nm) compared to many other existing second-generation PS with absorption peaks of 600 nm or above, which is suitable for deeper tissue penetration. These dual absorption peaks weaken its preference in the clinic compared to its counterparts and may be seen as a small setback. However, its current depth of UV-A light penetration is sufficient to apply for dermal and subdermal skin vasculature and may be used in other clinical practices to treat various types of carcinomas such as skin and others including nasopharyngeal, pancreatic, basal cell, bladder, and cancer [142], [143], [144], [145], [146].

Another significant plant-derived PS that was separated from the genus *Hypericum* specifically *Hypocrella bambusae* is natural hypocrellins, particularly hypocrellins A (HA) and B (HB). It is possible to convert HA to HB under certain circumstances. In fact, HY is a precursor or a parent compound of HA and HB and a soluble peryloquinone derivative. Like HY, these HA, HB compounds also exhibit strong red spectra absorption peaks and have shown strong photodynamic effects on tumors [126], [127], anti-viral potency, especially against human immunodeficiency virus type I and vesicular stomatitis [128], [129].

Recent technological advancements in the isolation and synthesis of HA and HB and their mechanism of action against various cancer types offer hope for its application in PDT [6], [25], [26], [128]. More importantly, many compelling evidence suggests that apoptosis type of cell death is involved which is mostly preferred in the clinic for better PDT outcomes without having any side effects. In traditional medicine, HA and HB were both used to treat vitiligo, psoriasis, and other skin conditions. Strong photodynamic tasks, minimal dark toxic reactions, the high quantum yield of O<sub>2</sub>, and other impressive physicochemical attributions make hypocrellins an attractive drug-lead compound in the pursuit of new and novel herb-derived PS agents [7], [8], [9], [128], [147], [148]. The above qualities led to numerous investigations for various experimental

hypocrellins on the priority list as possible novel photosensitizers employed for use in PDT of various tumors and various other applications in the clinic.

*Curcuma longa* (*Zingiberaceae*) is mostly employed in Indian conventional medication as a therapeutic for a wide range of illnesses, which include blood, hepatic, and stomach disorders, infections, and inflammation. One of *C. Longa*'s main bioactive substances is curcumin [149]. Numerous pharmacological activities, including antioxidant, anti-inflammatory, antimicrobial, antitumor, and hepatoprotective properties, have been linked to curcumin [150], [151]. In addition, curcumin is widely used in medications, food, and cosmetics. Scientists have been very interested in curcumin and curcuminoids' photoactive potential for many years [152].

According to previous investigations, curcumin fulfills all PS requirements and exhibits phototoxic effects [153], [154]. It has been determined that a daily dosage of 12 g/kg body weight of curcumin is reported as recommended and safe. The use of curcumin in anticancer research is made possible by its selectivity for the target cells. Curcumin was absorbed more preferentially or selectively by a number of tumor cell lines than by normal, healthy cells. Because of its high extinction coefficient and wide absorption spectrum between 300 nm and 500 nm, curcumin has the potential to cause significant phototoxicity even at micromolar doses [153], [154].

## Mechanism of Action of PS

PDT may be a promising treatment for patients with tumors. Despite recent advancements, the mechanism of its action of many PS is poorly understood and is different from the cytotoxic effects induced by known antitumor drugs because the exact molecular mechanism of action of PDT is still subject to many research efforts. However, it is known that multiple factors play a role in governing the outcome of PDT as well as the effective mechanism of action of irradiated PS including chlorins [155], [156]. C6-PDT targets not only neoplastic cells, microvasculature, and inflammatory and immune systems of hosts. It appears clear that the combination of all these components is required to achieve long-term control of the tumor. Many well-demonstrated results have indicated that the better outcome of PDT treatment mainly depends on the type, concentration, and intracellular localization of the photosensitizer [32], [157]. In addition, light wavelength, light fluence, and fluence rate are important to ensure sufficient O<sub>2</sub> availability and supply [106], [155], [158]. Besides the above, it is important to ensure sufficient light reaches the target and how the PS interacts with cells subcellular localization in the target tissue/tumor

also PS should be available for a certain time interval between its application and irradiation of the target cells [159]. Besides, the conditions in which the PDR occurs as they are crucial to determining the outcome of PDT to a certain extent [34], [160].

For many PSs, mainly chlorins found mitochondria as an important sub-cellular target used in PDT because these PSs are able to induce mitochondria-mediated apoptosis and cellular damage after illumination [161]. In most cases, it is accepted that the accumulation of a PS including Ce6 in mitochondria but less with ER [162], [163] leads to the activation of the apoptotic pathway in the cell. Kessel and Poretz [164] reported that Ce6 was located in the plasma membrane and/or mitochondria and later by Ali-Seyed *et al.* [32] confirmed their findings and also reported some additional sub-cellular targets including nucleus, lysosomes, and Golgi apparatus. However, this study failed to demonstrate both Ce6 and Photolon in ER. The mechanism of action of Ce6-induced PDT (Ce6-PDT) was not clear previously because the majority of investigations concentrate on tumor cells, but its detailed mechanism is mostly started clear now. The generation of highly reactive O<sub>2</sub> and the formation of ROS lead to the rise of free radical stress leads to the dysfunction of mitochondria and ER function and, subsequently, to the execution of all types of death programs such as apoptosis, necrosis, and autophagy [32], [34], [164], [165]. Besides the above, chlorin-PDT induces hypoxia by altering tumor vasculature, which leads to microvascular shutdown [166], [167] and induces inflammatory and immune responses [168].

Apoptotic type of programmed cell death by anti-cancer agents is initiated by the loss of mitochondrial potential [169]. Many investigations have observed notable disorders of mitochondrial potential 3 h PDT in nearly 50% of irradiated cells. It is known that the disruption of mitochondrial potential by free radicals leads to cytochrome C release into the cytoplasm, which in turn stimulates caspase cascades, which include executioner caspase-3 at the final stage. Moon *et al.* [170] have shown Ce6-PDT both *in vitro* and *in vivo* using a rat tumor model. In this study, 3-week-old male Sprague–Dawley rats were inoculated with RK3E-ras cells, followed by the administration of Ce6 for 24 h. PDT was employed using an advanced laser diode at a light dose of 100 J/cm<sup>2</sup>. Ce6-PDT induces apoptosis through the activation of caspase-3 and its downstream target such as PARP cleavage and the reduction of anti-apoptotic bcl-2. The *in vivo* experiments confirmed the above and demonstrated that Ce6-PDT led to a significant reduction in tumor size. These results suggest that Ce6-PDT can effectively arrest tumor growth by inhibiting cell proliferation and inducing apoptosis.

## Conclusion

In summary, at present, PDT has emerged as preferred therapeutic options for multiple malignant cancers. Photosensitizer Ce6 and its synthetic counterparts have proven to be useful in designing PDT as the most promising agents for clinical use against various types of cancer. Besides, various established investigations have demonstrated that PDT under specific therapeutic windows, which are ideal and successful cancer treatment by inducing apoptotic cell death. However, one can not rule out the possibility of eliciting potential complications in Ce6-based PDT *in vivo*, which targets multiple cell types including normal cells if an inappropriate wavelength of light sources is used. Although a discussion of every perspective of chlorin PSs on their anti-cancer tested in the last few years is beyond the scope of any single review, the present review discussed mostly the anti-cancer prospects not the other important potentials like anti-microbial efficacy of this photosensitizer Ce6 and its synthetic counterparts.

## References

1. Beutler JA. Natural products as a foundation for drug discovery. *Curr Protoc Pharmacol.* 2009;46:9.11.1-9.21. <https://doi.org/10.1002/0471141755.ph0911s46>  
PMid:20161632
2. Nasim N, Sandeep IS, Mohanty S. Plant-derived natural products for drug discovery: Current approaches and prospects. *Nucleus (Calcutta).* 2022;65(3):399-411. <https://doi.org/10.1007/s13237-022-00405-3>  
PMid:36276225
3. Bohr A, Memarzadeh K. The rise of artificial intelligence in healthcare applications. In: *Artificial Intelligence in Healthcare.* Cambridge: Academic Press; 2020. p. 25-60.
4. Mathur S, Hoskins C. Drug development: Lessons from nature. *Biomed Rep.* 2017;6(6):612-4. <https://doi.org/10.3892/br.2017.909>  
PMid:28584631
5. Mello MM. Barriers to ensuring access to affordable prescription drugs. *Annu Rev Pharmacol Toxicol.* 2020;60:275-89. <https://doi.org/10.1146/annurev-pharmtox-010919-023518>  
PMid:31136248
6. Ali SM, Chee SK, Yuen GY, Olivo M. Photodynamic therapy induced Fas-mediated apoptosis in human carcinoma cells. *Int J Mol Med* 2002;9(3):257-70.
7. Diwu Z. Novel therapeutic and diagnostic applications of hypocrellins and hypericins. *Photochem Photobiol.* 1995;61(6):529-39. <https://doi.org/10.1111/j.1751-1097.1995.tb09903.x>  
PMid:7568399
8. Diwu ZJ, Haugland RP, Liu J, Lown JW, Miller GG, Moore RB, *et al.* Photosensitization by anticancer agents 21: New perylene- and

- aminonaphthoquinones. *Free Radic Biol Med.* 1996;20(4):589-93. [https://doi.org/10.1016/0891-5849\(95\)02061-6](https://doi.org/10.1016/0891-5849(95)02061-6)  
PMid:8904300
9. Xu S, Chen S, Zhang M, Shen T, Zhao Y, Liu Z, *et al.* Butylamino-demethoxy-hypocrellins and photodynamic therapy decreases human cancer *in vitro* and *in vivo*. *Biochim Biophys Acta.* 2001;1537(3):222-32. [https://doi.org/10.1016/s0925-4439\(01\)00074-6](https://doi.org/10.1016/s0925-4439(01)00074-6)  
PMid:11731224
  10. Wainwright CL, Teixeira MM, Adelson DL, Braga FC, Buenz EJ, David B, *et al.* Future directions for the discovery of natural product-derived immunomodulating drugs: An IUPHAR positional review. *Pharmacol Res.* 2022;177:106076. <https://doi.org/10.1016/j.phrs.2022.106076>  
PMid:35074524
  11. Veeresham C. Natural products derived from plants as a source of drugs. *J Adv Pharm Technol Res.* 2012;3(4):200-1. <https://doi.org/10.4103/2231-4040.104709>  
PMid:23378939
  12. Talib WH, Alsalahat I, Daoud S, Abutayeh RF, Mahmud AI. Plant-derived natural products in cancer research: Extraction, mechanism of action, and drug formulation. *Molecules.* 2020;25(22):10.3390/molecules25225319. <https://doi.org/10.3390/molecules25225319>  
PMid:33202681
  13. Atanasov AG, Zotchev SB, Dirsch VM, International Natural Product Sciences Taskforce, Supuran CT. Natural products in drug discovery: Advances and opportunities. *Nat Rev Drug Discov.* 2021;20(3):200-16. <https://doi.org/10.1038/s41573-020-00114-z>  
PMid:33510482
  14. Wagner H, Ulrich-MG. Synergy research: Approaching a new generation of phytopharmaceuticals. *Phytomedicine.* 2009;16(2-3):97-110. <https://doi.org/10.1016/j.phymed.2008.12.018>  
PMid:19211237
  15. Geysen HM, Schoenen F, Wagner D, Wagner R. Combinatorial compound libraries for drug discovery: An ongoing challenge. *Nat Rev Drug Discov.* 2003;2(3):222-30. <https://doi.org/10.1038/nrd1035>  
PMid:12612648
  16. Vijayaraghavan K, Rajkumar J, Bukhari SN, Al-Sayed B, Seyed MA. *Chromolaena odorata*: A neglected weed with a wide spectrum of pharmacological activities (Review). *Mol Med Rep.* 2017;15(3):1007-16. <https://doi.org/10.3892/mmr.2017.6133>  
PMid:28112383
  17. Jantan I, Syed Nasir AB, Mohamed Ali SM, Wai LK, Mosaik MA. The evolving role of natural products from the tropical rainforests as a replenishable source of new drug leads. In: *Drug Discovery and Development-From Molecules to Medicine.* London: IntechOpen; 2015. p. 3-38.
  18. Amit KA, Chandrashekar DR, Shripal MC. *Natural Products in Drug Discovery. Pharmacognosy - Medicinal Plants.* London: IntechOpen; 2019.
  19. Seyed MA, Vijayaraghavan K. Dengue virus infections and anti-dengue virus activities of *Andrographis paniculata*. *Asian Pac J Trop Med.* 2020;13(2):49.
  20. Katiyar C, Gupta A, Kanjilal S, Katiyar S. Drug discovery from plant sources: An integrated approach. *Ayu.* 2012;33(1):10-9. <https://doi.org/10.4103/0974-8520.100295>  
PMid:23049178
  21. Seyed MA. A comprehensive review on *Phyllanthus* derived natural products as potential chemotherapeutic and immunomodulators for a wide range of human diseases. *Biocat Agric Biotechnol.* 2019;17:529-37.
  22. Li CQ, Lei HM, Hu QY, Li GH, Zhao PJ. Recent advances in the synthetic biology of natural drugs. *Front Bioeng Biotechnol.* 2021;9:691152. <https://doi.org/10.3389/fbioe.2021.691152>  
PMid:34395399
  23. Mouhssen L. The success of natural products in drug discovery. *Pharmacol Pharm.* 2013;4:17-31.
  24. Dougherty TJ. Photodynamic therapy. *Photochem Photobiol.* 1993;58:895-900.
  25. Ali SM, Olivo M, Yuen GY, Chee SK. Photodynamic-induced apoptosis of human nasopharyngeal carcinoma cells using Hypocrellins. *Int J Oncol.* 2001;19(3):633-43. <https://doi.org/10.3892/ijo.19.3.633>  
PMid:11494047
  26. Ali SM, Olivo M. Bio-distribution and subcellular localization of Hypericin and its role in PDT induced apoptosis in cancer cells. *Int J Oncol.* 2002;21(3):531-40.
  27. Robertson CA, Evans DH, Abrahamse H. Photodynamic therapy (PDT): A short review on cellular mechanisms and cancer research applications for PDT. *J Photochem Photobiol B.* 2009;96(1):1-8. <https://doi.org/10.1016/j.jphotobiol.2009.04.001>  
PMid:19406659
  28. Aziz B, Aziz I, Khurshid A, Raoufi E, Esfahani FN, Jalilian Z, *et al.* An overview of potential natural photosensitizers in cancer photodynamic therapy. *Biomedicines.* 2023;11(1):224. <https://doi.org/10.3390/biomedicines11010224>  
PMid:36672732
  29. Hamblin MR, Hasan T. Photodynamic therapy: A new antimicrobial approach to infectious disease? *Photochem Photobiol Sci.* 2004;3(5):436-50. <https://doi.org/10.1039/b311900a>  
PMid:15122361
  30. Gitika BK, Sharma SK, Huang YY, Dai T, Hamblin MR. Photodynamic therapy for infections. *Lasers Surg Med.* 2011;43(7):755-67. <https://doi.org/10.1002/lsm.21080>  
PMid:22057503
  31. Sharma SK, Mroz P, Dai T, Huang YY, St Denis TG, Hamblin MR. Photodynamic therapy for cancer and for infections: What is the difference? *Isr J Chem.* 2012;52(8-9):691-705. <https://doi.org/10.1002/ijch.201100062>  
PMid:23248387
  32. Ali-Sayed M, Bhuvaneshwari R, Soo KC, Olivo M. Photolon™ -photosensitization induces apoptosis via ROS-mediated cross-talk between mitochondria and lysosomes. *Int J Oncol.* 2011;39(4):821-31. <https://doi.org/10.3892/ijo.2011.1109>  
PMid:21725591
  33. Geltzer A, Turalba A, Vedula SS. Surgical implantation of steroids with antiangiogenic characteristics for treating neovascular age-related macular degeneration. *Cochrane Database Syst Rev.* 2013;1(1):CD005022. <https://doi.org/10.1002/14651858.CD005022.pub3>  
PMid:23440797
  34. Buytaert E, Dewaele M, Agostinis P. Molecular effectors of multiple cell death pathways initiated by photodynamic therapy. *Biochim Biophys Acta.* 2007;1776(1):86-107. <https://doi.org/10.1016/j.bbcan.2007.07.001>  
PMid:17693025
  35. Luo Y, Kessel D. Initiation of apoptosis versus necrosis by photodynamic therapy with chloroaluminum phthalocyanine (Review). *Photochem Photobiol.* 1997;66(4):479-83. <https://doi.org/10.1111/j.1751-1097.1997.tb03176.x>  
PMid:9337618
  36. Piette J, Volanti C, Vantieghem A, Matroule JY, Habraken Y, Agostinis P. Cell death and growth arrest in response to photodynamic therapy with membrane-bound

- photosensitizers. *Biochem Pharmacol.* 2003;66(8):1651-9. [https://doi.org/10.1016/s0006-2952\(03\)00539-2](https://doi.org/10.1016/s0006-2952(03)00539-2)  
PMid:14555246
37. Wang KN, Liu LY, Qi G, Chao XJ, Ma W, Yu Z, *et al.* Light-driven cascade mitochondria-to-nucleus photosensitization in cancer cell ablation. *Adv Sci (Weinh).* 2021;8(8):2004379. <https://doi.org/10.1002/advs.202004379>  
PMid:33898198
38. Yoo JO, Ha KS. New insights into the mechanisms for photodynamic therapy-induced cancer cell death. *Int Rev Cell Mol Biol.* 2012;295:139-74. <https://doi.org/10.1016/B978-0-12-394306-4.00010-1>  
PMid:22449489
39. Yang J, Griffin A, Qiang Z, Ren J. Organelle-targeted therapies: A comprehensive review on system design for enabling precision oncology. *Signal Transduct Target Ther.* 2022;7(1):379. <https://doi.org/10.1038/s41392-022-01243-0>  
PMid:36402753
40. Nowak-Stępniewska A, Wiktorska K, Małecki M, Romiszewska A, Padzik-Graczyk A. Cytotoxicity of PP(Arg)(2)- and Hp(Arg)(2)-mediated photodynamic therapy and early stage of apoptosis induction in prostate carcinoma *in vitro*. *Acta Biochim Pol.* 2011;58(4):497-505.
41. Lima E, Reis LV. Photodynamic therapy: From the basics to the current progress of N-heterocyclic-bearing dyes as effective photosensitizers. *Molecules.* 2023;28(13):5092. <https://doi.org/10.3390/molecules28135092>  
PMid:37446758
42. Marrelli M, Menichini G, Provenzano E, Conforti F. Applications of natural compounds in the photodynamic therapy of skin cancer. *Curr Med Chem.* 2014;21(12):1371-90. <https://doi.org/10.2174/092986732112140319094324>  
PMid:23531223
43. Vaou N, Stavropoulou E, Voidarou C, Tsigalou C, Bezirtzoglou E. Towards advances in medicinal plant antimicrobial activity: A review study on challenges and future perspectives. *Microorganisms.* 2021;9(10):2041. <https://doi.org/10.3390/microorganisms9102041>  
PMid:34683362
44. Newman DJ, Cragg GM. Natural products as sources of new drugs from 1981 to 2014. *J Nat Prod.* 2016;79(3):629-61. <https://doi.org/10.1021/acs.jnatprod.5b01055>  
PMid:26852623
45. Juarranz A, Jaén P, Sanz-Rodríguez F, Cuevas J, González S. Photodynamic therapy of cancer. Basic principles and applications. *Clin Transl Oncol.* 2008;10(3):148-54. <https://doi.org/10.1007/s12094-008-0172-2>  
PMid:18321817
46. O'Connor AE, Gallagher WM, Byrne AT. Porphyrin and nonporphyrin photosensitizers in oncology: Preclinical and clinical advances in photodynamic therapy. *Photochem Photobiol.* 2009;85(5):1053-74. <https://doi.org/10.1111/j.1751-1097.2009.00585.x>  
PMid:19682322
47. Hamblin MR, Chiang LY, Lakshmanan S, Huang YY, Garcia-Diaz M, Karimi M, *et al.* Nanotechnology for photodynamic therapy: A perspective from the Laboratory of Dr. Michael R. Hamblin in the Wellman Center for Photomedicine at Massachusetts General Hospital and Harvard Medical School. *Nanotechnol Rev.* 2015;4(4):359-72. <https://doi.org/10.1515/ntrev-2015-0027>  
PMid: 26640747
48. Pervaiz S, Olivo M. Art and science of photodynamic therapy. *Clin Exp Pharmacol Physiol.* 2006;33(5-6):551-6. <https://doi.org/10.1111/j.1440-1681.2006.04406.x>  
PMid:16700893
49. Algorri JF, López-Higuera JM, Rodríguez-Cobo L, Cobo A. Advanced light source technologies for photodynamic therapy of skin cancer lesions. *Pharmaceutics.* 2023;15(8):2075. <https://doi.org/10.3390/pharmaceutics15082075>  
PMid:37631289
50. Kubrak TP, Kołodziej P, Sawicki J, Mazur A, Kozirowska K, Aebisher D. Some natural photosensitizers and their medicinal properties for use in photodynamic therapy. *Molecules.* 2022;27(4):1192. <https://doi.org/10.3390/molecules27041192>  
PMid:35208984
51. Brancalion L, Moseley H. Laser and non-laser light sources for photodynamic therapy. *Lasers Med Sci.* 2002;17(3):173-86. <https://doi.org/10.1007/s101030200027>  
PMid:12181632
52. Boyle RW, Dolphin D. Structure and biodistribution relationships of photodynamic sensitizers. *Photochem Photobiol.* 1996;64(3):469-85. <https://doi.org/10.1111/j.1751-1097.1996.tb03093.x>  
PMid:8806226
53. Benov L. Photodynamic therapy: Current status and future directions. *Med Princ Pract.* 2015;24 Suppl 1(Suppl 1):14-28. <https://doi.org/10.1159/000362416>  
PMid:24820409
54. Detty MR, Gibson SL, Wagner SJ. Current clinical and preclinical photosensitizers for use in photodynamic therapy. *J Med Chem.* 2004;47(16):3897-915. <https://doi.org/10.1021/jm040074b>  
PMid:15267226
55. Udrea AM, Smarandache A, Dinache A, Mares C, Nistorescu S, Avram S, *et al.* Photosensitizers-loaded nanocarriers for enhancement of photodynamic therapy in melanoma treatment. *Pharmaceutics.* 2023;15(8):2124. <https://doi.org/10.3390/pharmaceutics15082124>  
PMid:37631339
56. Palumbo G. Photodynamic therapy and cancer: A brief sightseeing tour. *Expert Opin Drug Deliv.* 2007;4(2):131-48. <https://doi.org/10.1517/17425247.4.2.131>  
PMid:17335411
57. Sarbadhikary P, George BP, Abrahamse H. Potential application of photosensitizers with high-Z elements for synergic cancer therapy. *Front Pharmacol.* 2022;13:921729. <https://doi.org/10.3389/fphar.2022.921729>  
PMid:35837287
58. Downum KR, Wen J. The Occurrence of Photosensitizers among higher plants. In: *Light-Activated Pest Control*. Ch. 11. Washington, DC: The American Chemical Society; 1995. p. 135-43.
59. Thirumurugan D, Cholarajan A, Raja SS, Vijayakumar R. An introductory chapter. In: *Secondary Metabolites-Sources Applications*. London: IntechOpen; 2018. p. 1-21.
60. Kennedy DO, Wightman EL. Herbal extracts and phytochemicals: Plant secondary metabolites and the enhancement of human brain function. *Adv Nutr.* 2011;2(1):32-50. <https://doi.org/10.3945/an.110.000117>  
PMid:22211188
61. Marrelli M, Statti G, Conforti F. A Review of Biologically Active Natural Products from Mediterranean Wild Edible Plants: Benefits in the Treatment of Obesity and Its Related Disorders. *Molecules.* 2020;25(3):649. <https://doi.org/10.3390/molecules25030649>  
PMid: 32028716
62. Jong WW, Tan PJ, Kamarulzaman FA, Mejin M, Lim D, Ang I, *et al.* Photodynamic activity of plant extracts from Sarawak, Borneo. *Chem Biodivers.* 2013;10(8):1475-86. <https://doi.org/10.1002/cbdv.201200303>

- PMid:23939795
63. Mansoori B, Mohammadi A, Amin Doustvandi M, Mohammadnejad F, Kamari F, Gjerstorff MF, *et al.* Photodynamic therapy for cancer: Role of natural products. *Photodiagnosis Photodyn Ther.* 2019;26:395-404. <https://doi.org/10.1016/j.pdpdt.2019.04.033>  
PMid:31063860
  64. Foresto E, Gilardi P, Ibarra LE, Cogno IS. Light-activated green drugs: How we can use them in photodynamic therapy and mass-produce them with biotechnological tools. *Phytomed Plus.* 2021;1(3):100044. <https://doi.org/10.1016/j.phyplu.2021.100044>
  65. haneshwar S, Patil K, Bulbule M, Kinjawadekar V, Joshi D, Joshi V. Photodynamic therapy for cancer. *Int J Pharm Sci Rev Res.* 2014;27(2):125-41.
  66. Baskaran R, Lee J, Yang SG. Clinical development of photodynamic agents and therapeutic applications. *Biomater Res.* 2018;22:25. <https://doi.org/10.1186/s40824-018-0140-z>  
PMid:30275968
  67. Berlanda J, Kiesslich T, Engelhardt V, Krammer B, Plaetzer K. Comparative *in vitro* study on the characteristics of different photosensitizers employed in PDT. *J Photochem Photobiol B.* 2010;100(3):173-80. <https://doi.org/10.1016/j.jphotobiol.2010.06.004>  
PMid:20599390
  68. Almadi KH, Alkahtany MF, Almutairi B. Influence of synthetic and natural photosensitizers activated by photodynamic therapy on extrusion bond strength of fiber post to radicular dentin. *Pak J Med Sci.* 2021;37(7):1912-7. <https://doi.org/10.12669/pjms.37.7.4331>  
PMid:34912417
  69. Shrestha R, Mallik SK, Lim J, Gurung P, Magar TBT, Kim YW. Efficient synthesis of chlorin e6 and its potential photodynamic immunotherapy in mouse melanoma by the abscopal effect. *Int J Mol Sci.* 2023;24(4):10.3390/ijms24043901. <https://doi.org/10.3390/ijms24043901>  
PMid:36835310
  70. Sobaniec S, Bernaczyk P, Pietruski J, Cholewa M, Skurska A, Dolińska E, *et al.* Clinical assessment of the efficacy of photodynamic therapy in the treatment of oral lichen planus. *Lasers Med Sci.* 2013;28(1):311-6. <https://doi.org/10.1007/s10103-012-1153-9>  
PMid:22814895
  71. Allison RR, Sibata C, Mang TS, Bagnato VS, Downie GH, Hu XH, *et al.* Photodynamic therapy for chest wall recurrence from breast cancer. *Photodiagnosis Photodyn Ther.* 2004;1(2):157-71. [https://doi.org/10.1016/S1572-1000\(04\)00039-0](https://doi.org/10.1016/S1572-1000(04)00039-0)  
PMid:25048186
  72. Overchuk M, Weersink RA, Wilson BC, Zheng G. Photodynamic and photothermal therapies: Synergy opportunities for nanomedicine. *ACS Nano.* 2023;17(9):7979-8003. <https://doi.org/10.1021/acsnano.3c00891>  
PMid:37129253
  73. Kessel D, Luo Y. Mitochondrial photodamage and PDT-induced apoptosis. *J Photochem Photobiol B.* 1998;42(2):89-95. [https://doi.org/10.1016/s1011-1344\(97\)00127-9](https://doi.org/10.1016/s1011-1344(97)00127-9)  
PMid:9540214
  74. Galanou MC, Theodossiou TA, Tsiourvas D, Sideratou Z, Paleos CM. Interactive transport, subcellular relocation and enhanced phototoxicity of hypericin encapsulated in guanidylated liposomes via molecular recognition. *Photochem Photobiol.* 2008;84(5):1073-83. <https://doi.org/10.1111/j.1751-1097.2008.00392.x>  
PMid:18627515
  75. Zhao J, Wu W, Sun J, Guo S. Triplet photosensitizers: From molecular design to applications. *Chem Soc Rev.* 2013;42(12):5323-51. <https://doi.org/10.1039/c3cs35531d>  
PMid:23450221
  76. Blum NT, Zhang Y, Qu J, Lin J, Huang P. Recent advances in self-exciting photodynamic therapy. *Front Bioeng Biotechnol.* 2020;8:594491. <https://doi.org/10.3389/fbioe.2020.594491>  
PMid:33195164
  77. Yoon I, Li JZ, Shim YK. Advance in photosensitizers and light delivery for photodynamic therapy. *Clin Endosc.* 2013;46(1):7-23. <https://doi.org/10.5946/ce.2013.46.1.7>  
PMid:23423543
  78. Chin WW, Heng PW, Bhuvanewari R, Lau WK, Olivo M. The potential application of chlorin e6-polyvinylpyrrolidone formulation in photodynamic therapy. *Photochem Photobiol Sci.* 2006;5(11):1031-7. <https://doi.org/10.1039/b605772a>  
PMid:17077899
  79. Trukhachova T. Safety and Efficacy of Photosensitizer Photolon (Fotolon) in Photodynamic Therapy. In: *Proceeding SPIE 11070, 17<sup>th</sup> International Photodynamic Association World Congress, 1107037; 2019.* <https://doi.org/10.1117/12.2528083>
  80. Waidelich R. Laser-induced lithotripsy and photodynamic therapy in urology: A short introduction to current laser applications. *Med Laser Appl.* 2010;25(1):14-9.
  81. Li JH, Chen ZQ, Huang Z, Zhan Q, Ren FB, Liu JY, *et al.* *In vitro* study of low intensity ultrasound combined with different doses of PDT: Effects on C6 glioma cells. *Oncol Lett.* 2013;5(2):702-6. <https://doi.org/10.3892/ol.2012.1060>  
PMid:23420417
  82. Copley L, Pauline WV, Wirtz KW, Iqbal Parker M, Leaner VD. Photolon, a chlorin e6 derivative, triggers ROS production and light-dependent cell death via necrosis. *Int J Biochem Cell Biol.* 2008;40(2):227-35. <https://doi.org/10.1016/j.biocel.2007.07.014>  
PMid:17822943
  83. Isakau HA, Parkhats MV, Knyukshto VN, Dzhagarov BM, Petrov EP, Petrov PT. Toward understanding the high PDT efficacy of chlorin e6-polyvinylpyrrolidone formulations: Photophysical and molecular aspects of photosensitizer-polymer interaction *in vitro.* *J Photochem Photobiol B.* 2008;92(3):165-74. <https://doi.org/10.1016/j.jphotobiol.2008.06.004>  
PMid:18656379
  84. Juzeniene A, Thu Tam TT, Iani V, Moan J. 5-Methyltetrahydrofolate can be photodegraded by endogenous photosensitizers. *Free Radic Biol Med.* 2009;47(8):1199-204. <https://doi.org/10.1016/j.freeradbiomed.2009.07.030>  
PMid:19647791
  85. Trukhachova TV, Shliakhtsin SV, Cerkovsky DA, Istomin YP. A novel finished formulation of the photosensitizer Photolon® for topical application. Evaluation of the efficacy in patients with basal-cell carcinoma of the skin. *Photodiagn Photodyn Ther.* 2011;8:200-1.
  86. Sharma S, Bakal J, Oliver-Fernandez A, Blair J. Photodynamic therapy with verteporfin for subfoveal choroidal neovascularization in age-related macular degeneration: Results of an effectiveness study. *Arch Ophthalmol.* 2004;122(6):853-6. <https://doi.org/10.1001/archophth.122.6.853>  
PMid:15197060
  87. Horibe S, Nagai J, Yumoto R, Tawa R, Takano M. Accumulation and photodynamic activity of chlorin e6 in cisplatin-resistant human lung cancer cells. *J Pharm Sci.* 2011;100(7):3010-7. <https://doi.org/10.1002/jps.22501>  
PMid:21274848
  88. Spikes JD. Chlorins as photosensitizers in biology and medicine. *J Photochem Photobiol B.* 1990;6(3):259-74. [https://doi.org/10.1016/1011-1344\(90\)85096-f](https://doi.org/10.1016/1011-1344(90)85096-f)  
PMid:2120404

89. Ding HL, Wang XL, Wang HW, Huang Z. Successful treatment of refractory facial acne using repeat short-cycle ALA-PDT: Case study. *Photodiagnosis Photodyn Ther.* 2011;8(4):343-6. <https://doi.org/10.1016/j.pdpdt.2011.07.003>  
PMid:22122923
90. Cabrera H, Castro J, Grassi HC, Andrades ED, López-Rivera SA. The effect of photodynamic therapy on contiguous untreated tumor. *Dermatol Surg.* 2012;38(7 Pt 1):1097-9. <https://doi.org/10.1111/j.1524-4725.2012.02400.x>  
PMid:22471374
91. Thong PS, Olivo M, Kho KW, Bhuvanewari R, Chin WW, Ong KW, *et al.* Immune response against angiosarcoma following lower fluence rate clinical photodynamic therapy. *J Environ Pathol Toxicol Oncol.* 2008;27(1):35-42. <https://doi.org/10.1615/jenvironpatholtoxicoloncol.v27.i1.40>  
PMid:18551894
92. Marchal S, François A, Dumas D, Guillemin F, Bezdetsnaya L. Relationship between subcellular localisation of Foscan® and caspase activation in photosensitized MCF-7 cells. *Br J Cancer.* 2007;96(6):944-51. <https://doi.org/10.1038/sj.bjc.6603631>  
PMid:17325708
93. Dobson J, de Queiroz GF, Golding JP. Photodynamic therapy and diagnosis: Principles and comparative aspects. *Vet J.* 2018;233:8-18. <https://doi.org/10.1016/j.tvjl.2017.11.012>  
PMid:29486883
94. Meier D, Botter SM, Campanile C, Robl B, Gräfe S, Pellegrini G, *et al.* Foscan and foslip based photodynamic therapy in osteosarcoma in vitro and in intratibial mouse models. *Int J Cancer.* 2017;140(7):1680-92. <https://doi.org/10.1002/ijc.30572>  
PMid:27943293
95. Spikes JD, Bommer JC. Photosensitizing properties of mono-L-aspartyl chlorin e6 (NPe6): A candidate sensitizer for the photodynamic therapy of tumors. *J Photochem Photobiol B.* 1993;17(2):135-43. [https://doi.org/10.1016/1011-1344\(93\)80006-u](https://doi.org/10.1016/1011-1344(93)80006-u)  
PMid:8459317
96. Yumita N, Iwase Y, Nishi K, Ikeda T, Komatsu H, Fukai T, *et al.* Sonodynamically-induced antitumor effect of mono-L-aspartyl chlorin e6 (NPe6). *Anticancer Res* 2011;31(2):501-6.
97. Aizawa K, Okunaka T, Ohtani T, Kawabe H, Yasunaka Y, O'Hata S, *et al.* Localization of mono-L-aspartyl chlorin e6 (NPe6) in mouse tissues. *Photochem Photobiol.* 1987;46(5):789-93. <https://doi.org/10.1111/j.1751-1097.1987.tb04849.x>  
PMid:3441501
98. Ferreira S, Juliana Menezes PF, Kurachi C, Sibata C, Allison RR, Bagnato V. Photostability of different chlorine photosensitizers. *Laser Phys Lett.* 2008;5:156-61.
99. Mirzaei H, Djavid GE, Hadizadeh M, Jahanshiri-Moghadam M, Hajian P. The efficacy of Radachlorin-mediated photodynamic therapy in human hepatocellular carcinoma cells. *J Photochem Photobiol B.* 2015;142:86-91. <https://doi.org/10.1016/j.jphotobiol.2014.11.007>  
PMid:25528192
100. Ghoadarzi R, Changizi V, Montazerabadi AR, Eyvazzadaeh N. Assessing of integration of ionizing radiation with Radachlorin-PDT on MCF-7 breast cancer cell treatment. *Lasers Med Sci.* 2016;31(2):213-9. <https://doi.org/10.1007/s10103-015-1844-0>  
PMid:26690358
101. Kochneva EV, Filonenko EV, Vakulovskaya EG, Scherbakova EG, Seliverstov OV, Markichev NA, *et al.* Photosensitizer radachlorin®: Skin cancer PDT phase II clinical trials. *Photodiagnosis Photodyn Ther.* 2010;7(4):258-67. <https://doi.org/10.1016/j.pdpdt.2010.07.006>  
PMid:21112549
102. Anand S, Rollakanti KR, Brankov N, Brash DE, Hasan T, Maytin EV. Fluorouracil enhances photodynamic therapy of squamous cell carcinoma via a p53-independent mechanism that increases protoporphyrin IX levels and tumor cell death. *Mol Cancer Ther.* 2017;16(6):1092-101. <https://doi.org/10.1158/1535-7163.MCT-16-0608>  
PMid:28336806
103. Gijssens A, De Witte P. Photocytotoxic action of EGF-PVA-Sn(IV) chlorin e6 and EGF-dextran-Sn(IV)chlorin e6 internalizable conjugates on A431 cells. *Int J Oncol.* 1998;13(6):1171-7. <https://doi.org/10.3892/ijo.13.6.1171>  
PMid:9824627
104. Brasseur N, Ouellet R, La Madeleine C, van Lier JE. Water-soluble aluminium phthalocyanine-polymer conjugates for PDT: Photodynamic activities and pharmacokinetics in tumour-bearing mice. *Br J Cancer.* 1999;80(10):1533-41. <https://doi.org/10.1038/sj.bjc.6690557>  
PMid:10408394
105. Bonnett R, Djelal BD, Nguyen A. Physical and chemical studies related to the development of m-THPC (FOSCAN®) for the photodynamic therapy (PDT) of tumours. *J. Porphyrins Phthalocyanines.* 2001;5:652-61.
106. Sibata CH, Colussi VC, Oleinick NL, Kinsella TJ. Photodynamic therapy: A new concept in medical treatment. *Braz J Med Biol Res.* 2000;33(8):869-80. <https://doi.org/10.1590/s0100-879x2000000800002>  
PMid:11023333
107. Zhang J, Jiang C, Figueiró Longo JP, Azevedo RB, Zhang H, Muehlmann LA. An updated overview on the development of new photosensitizers for anticancer photodynamic therapy. *Acta Pharm Sin B.* 2018;8(2):137-46. <https://doi.org/10.1016/j.apsb.2017.09.003>  
PMid:29719775
108. Allison RR, Bagnato VS, Cuenca R, Downie GH, Sibata CH. The future of photodynamic therapy in oncology. *Future Oncol.* 2006;2(1):53-71. <https://doi.org/10.2217/14796694.2.1.53>  
PMid:16556073
109. Busch T, Cengel KA, Finlay J. Pheophorbide a as a photosensitizer in photodynamic therapy: *In vivo* considerations. *Cancer Biol Ther.* 2009;8(6):540-2. <https://doi.org/10.4161/cbt.8.6.8067>  
PMid:19252412
110. Mojzisoava H, Bonneau S, Vever-Bizet C, Braut D. Cellular uptake and subcellular distribution of chlorin e6 as functions of pH and interactions with membranes and lipoproteins. *Biochim Biophys Acta.* 2007;1768(11):2748-56. <https://doi.org/10.1016/j.bbamem.2007.07.002>  
PMid:17692283
111. Shim G, Lee S, Kim YB, Kim CW, Oh YK. Enhanced tumor localization and retention of chlorin e6 in cationic nanolipoplexes potentiate the tumor ablation effects of photodynamic therapy. *Nanotechnology.* 2011;22(36):365101. <https://doi.org/10.1088/0957-4484/22/36/365101>  
PMid:21841215
112. Battersby AR. Tetrapyrroles: The pigments of life. *Nat Prod Rep.* 2000;17(6):507-26. <https://doi.org/10.1039/b002635m>  
PMid:11152419
113. Li Z, Wang C, Cheng L, Gong H, Yin S, Gong Q, *et al.* PEG-functionalized iron oxide nanoclusters loaded with chlorin e6 for targeted, NIR light induced, photodynamic therapy. *Biomaterials.* 2013;34(36):9160-70. <https://doi.org/10.1016/j.biomaterials.2013.08.041>  
PMid:24008045
114. Sun L, Li Q, Hou M, Gao Y, Yang R, Zhang L, *et al.* Light-activatable Chlorin e6 (Ce6)-imbedded erythrocyte membrane vesicles camouflaged Prussian blue nanoparticles for synergistic

- photothermal and photodynamic therapies of cancer. *Biomater Sci.* 2013;6(11):2881-95. <https://doi.org/10.1039/c8bm00812d> PMID:30192355
115. Kostenich GA, Zhuravkin IN, Zhavrid EA. Experimental grounds for using chlorin e6 in the photodynamic therapy of malignant tumors. *J Photochem Photobiol B.* 1994;22(3):211-7. [https://doi.org/10.1016/1011-1344\(93\)06974-8](https://doi.org/10.1016/1011-1344(93)06974-8) PMID:8014753
116. Brockmann H, Haschad MN, Maier K, et al. About hypericin, the photodynamically active dye from *Hypericum perforatum*. *Nat Sci.* 1939;27:550.
117. Abels C, Szeimies RM, Steinbach P, Richert C, Goetz AE. Targeting of the tumor microcirculation by photodynamic therapy with a synthetic porphycene. *J Photochem Photobiol B.* 1997;40(3):305-12. [https://doi.org/10.1016/s1011-1344\(97\)00074-2](https://doi.org/10.1016/s1011-1344(97)00074-2) PMID:9372621
118. Blant SA, Woodtli A, Wagnières G, Fontollet C, van den Bergh H, Monnier P. *In vivo* fluence rate effect in photodynamic therapy of early cancers with tetra(m-hydroxyphenyl) chlorin. *Photochem Photobiol.* 1996;64(6):963-8. <https://doi.org/10.1111/j.1751-1097.1996.tb01862.x> PMID:8972639
119. Ahmad N, Gupta S, Feyes DK, Mukhtar H. Involvement of Fas (APO-1/CD-95) during photodynamic-therapy-mediated apoptosis in human epidermoid carcinoma A431 cells. *J Invest Dermatol.* 2006;115(6):1041-6. <https://doi.org/10.1046/j.1523-1747.2000.00147.x> PMID:11121139
120. Lam M, Oleinick NL, Nieminen AL. Photodynamic therapy-induced apoptosis in epidermoid carcinoma cells. Reactive oxygen species and mitochondrial inner membrane permeabilization. *J Biol Chem.* 2001;276(50):47379-86. <https://doi.org/10.1074/jbc.M107678200> PMID:11579101
121. Xu NF, Li JF, Cao EH, Wang JZ. Direct observation of dynamic process of cellular uptake of hypocrellin A in HeLa cells. *Acta Bio Physica Sinica.* 1995;11:261-6.
122. Miller GG, Brown K and Ballengrud AM. Preclinical assessment of Hypocrellins and hypocrellin B derivatives as sensitizers for photodynamic therapy of cancer: Progress update. *Photochem Photobiol.* 1995;65:714-22.
123. Lavie G, Mazur Y, Lavie D, Meruelo D. The chemical and biological properties of hypericin—a compound with a broad spectrum of biological activities. *Med Res Rev.* 1995;15(2):111-9. <https://doi.org/10.1002/med.2610150203> PMID:7739292
124. Miller GG, Brown K, Moore RB, Diwu ZJ, Liu J, Huang L, et al. Uptake kinetics and intracellular localization of hypocrellin photosensitizer for photodynamic therapy: Preclinical assessment of Hypocrellin A and Hypocrellin B as sensitizers for PDT of cancers. *Photochem Photobiol.* 1995;61(6):632-638. <https://doi.org/10.1111/j.1751-1097.1995.tb09880.x> PMID: 7568409
125. Miller GG, Brown K, Moore RB, Diwu ZJ, Liu J, Huang L, et al. Uptake kinetics and intracellular localization of hypocrellin photosensitizer for photodynamic therapy: A confocal microscopy study. *Photochem Photobiol.* 1995;61(6):632-8. <https://doi.org/10.1111/j.1751-1097.1995.tb09880.x> PMID:7568409
126. Dong CY, Jia HT, Ma CM. The inhibitory effect of the new photosensitizer hypocrellin A on experimental tumors. *Chin J Biochem.* 1987;20:468-72.
127. Kamuhabwa AR, Agostinis P, D'Hallewin MA, Kasran A, de Witte PA. Photodynamic activity of hypericin in human urinary bladder carcinoma cells. *Anticancer Res.* 2000;20(4):2579-84.
128. Hudson JB, Zhou J, Chen J, Harris L, Yip L, Towers GH. Hypocrellin, from *Hypocrella bambusa*, is phototoxic to human immunodeficiency virus. *Photochem Photobiol.* 1994;60(3):253-5. <https://doi.org/10.1111/j.1751-1097.1994.tb05100.x> PMID:7972377
129. Hirayama J, Ikebuchi K, Abe H, Kwon KW, Ohnishi Y, Horiuchi M, et al. Photoactivation of virus infectivity by hypocrellin A. *Photochem Photobiol.* 1997;66:697-700.
130. Diwu Z, Lown JW. Photosensitization by anticancer agents 12. Perylene quinonoid pigments, a novel type of singlet oxygen sensitizer. *J Photochem Photobiol Chem.* 1992;64(3):273-87. [https://doi.org/10.1016/1010-6030\(92\)85002-C](https://doi.org/10.1016/1010-6030(92)85002-C)
131. Kitanov GM. Hypericin and pseudohypericin in some *Hypericum* species. *Biochem Syst Ecol.* 2001;29(2):171-8. [https://doi.org/10.1016/s0305-1978\(00\)00032-6](https://doi.org/10.1016/s0305-1978(00)00032-6) PMID:11106845
132. Ayan AK, Cirak C, Kevseroglu K, Ozen T. Hypericin in some *Hypericum* species from Turkey. *Asian J Plant Sci.* 2004;3:200-2.
133. Dewick PM. *Medicinal Natural Products: A Biosynthetic Approach.* 2<sup>nd</sup> ed. Chichester: John Wiley & Sons Ltd.; 2002.
134. Garnica S, Weiss M, Oberwinkler F. Morphological and molecular phylogenetic studies in South American *Cortinarius* species. *Mycol Res.* 2003;107(Pt 10):1143-56. <https://doi.org/10.1017/s0953756203008414> PMID:14635763
135. Kusari S, Lamshöft M, Zühlke S, Spiteller M. An endophytic fungus from *Hypericum perforatum* that produces hypericin. *J Nat Prod.* 2008;71(2):159-62. <https://doi.org/10.1021/np070669k> PMID:18220354
136. Kusari S, Zühlke S, Kosuth J, Cellárová E, Spiteller M. Light-independent metabolomics of endophytic *Thielavia subthermophila* provides insight into microbial hypericin biosynthesis. *J Nat Prod.* 2009;72(10):1825-35. <https://doi.org/10.1021/np9002977> PMID:19746917
137. Kucharíková A, Kimáková K, Janfelt C, Čellárová E. Interspecific variation in localization of hypericins and phloroglucinols in the genus *Hypericum* as revealed by desorption electrospray ionization mass spectrometry imaging. *Physiol Plant.* 2016;157(1):2-12. <http://doi.org/10.1111/ppl.12422> PMID:26822391
138. Cellárová E. Effect of exogenous morphogenetic signals on differentiation *in vitro* and secondary metabolite formation in the genus *Hypericum*. In: Odabas MS, Çirak C, editors. *Medicinal and Aromatic Plant Science and Biotechnology 5 (Special Issue 1)*. Ikenobe: Global Science Books; 2011. p. 62-9.
139. Košuth J, Koperdákóvá J, Tolonen A, Hohtola A, Cellárová E. The content of hypericins and phloroglucinols in *Hypericum perforatum* L. seedlings at early stage of development. *Plant Sci.* 2003;165:515-21.
140. Urbanová M, Kosuth J, Cellárová E. Genetic and biochemical analysis of *Hypericum perforatum* L. plants regenerated after cryopreservation. *Plant Cell Rep.* 2006;25(2):140-7. <https://doi.org/10.1007/s00299-005-0050-0> PMID:16456647
141. Brunáková K, Petijová L, Zámecník J, Turecková V, Cellárová E. The role of ABA in the freezing injury avoidance in two *Hypericum* species differing in frost tolerance and potential to synthesize hypericins. *Plant Cell Tissue Organ Cult.* 2015;122:45-56.
142. Bhuvaneswari R, Gan YY, Yee KK, Soo KC, Olivo M. Effect of hypericin-mediated photodynamic therapy on the expression of vascular endothelial growth factor in human nasopharyngeal carcinoma. *Int J Mol Med.* 2007;20(4):421-8.
143. Olivo M, Du HY, Bay BH. Hypericin lights up the way for the

- potential treatment of nasopharyngeal cancer by photodynamic therapy. *Curr Clin Pharmacol*. 2006;1(3):217-22. <https://doi.org/10.2174/157488406778249370>  
PMid:18666746
144. Kaihong Z, Lijin J. Conversion of Hypocrellin A in alkaline and neutral media. *Chin J Org Chem*. 1989;9:252.
145. Estey EP, Brown K, Diwu Z, Liu J, Lown JW, Miller GG, *et al*. Hypocrellins as photosensitizers for photodynamic therapy: A screening evaluation and pharmacokinetic study. *Cancer Chemother Pharmacol*. 1996;37(4):343-50. <https://doi.org/10.1007/s002800050395>  
PMid:8548880
146. Zhenjun D, Lown JW. Hypocrellins and their use in photosensitization. *Photochem Photobiol*. 1990;52(3):609-16. <https://doi.org/10.1111/j.1751-1097.1990.tb01807.x>  
PMid:2284353
147. Wakdikar S. Global health care challenge: Indian experiences and new prescriptions. *Electron J Biotechnol*. 2004;7:214-20.
148. Bhutani KK, Gohil VM. Natural products drug discovery research in India: Status and appraisal. *Indian J Exp Biol*. 2010;48(3):199-207.
149. Zhang HA, Kitts DD. Turmeric and its bioactive constituents trigger cell signaling mechanisms that protect against diabetes and cardiovascular diseases. *Mol Cell Biochem*. 2021;476(10):3785-814. <https://doi.org/10.1007/s11010-021-04201-6>  
PMid:34106380
150. Rathaur P, Raja W, Ramteke PW, Suchit AJ. Turmeric: The golden spice of life. *Int J Pharm Sci Res*. 2012;3:1987-94.
151. Tung BT, Nham DT, Hai NT, Thu DK. *Curcuma longa*, the Polyphenolic curcumin compound and pharmacological effects on liver. In: Watson RR, Preedy VR, editors. *Dietary Interventions in Liver Disease*. Ch. 10. Cambridge, MA, USA: Academic Press; 2019. p. 125-34.
152. Kazantzis KT, Koutsonikoli K, Mavroidi B, Zachariadis M, Alexiou P, Pelecanou M., Politopoulos K, *et al*. Curcumin derivatives as photosensitizers in photodynamic therapy: Photophysical properties and *in vitro* studies with prostate cancer cells. *Photochem Photobiol Sci*. 2010;19:193-206.
153. Dahl TA, McGowan WM, Shand MA, Srinivasan VS. Photokilling of bacteria by the natural dye curcumin. *Arch Microbiol*. 1989;151(2):183-5. <https://doi.org/10.1007/BF00414437>  
PMid:2655550
154. Haukvik T, Bruzell E, Kristensen S, Tønnesen HH. Photokilling of bacteria by curcumin in selected polyethylene glycol 400 (PEG 400) preparations. *Studies on curcumin and curcuminoids, XLI. Pharmazie*. 2010;65(8):600-6.
155. Dougherty TJ, Gomer CJ, Henderson BW, Jori G, Kessel D, Korbelik M, *et al*. Photodynamic therapy. *J Natl Cancer Inst*. 1998;90:889-905.
156. Gupta S, Dwarakanath BS, Muralidhar K, Koru-Sengul T, Jain V. Non-monotonic changes in clonogenic cell survival induced by disulphonated aluminum phthalocyanine photodynamic treatment in a human glioma cell line. *J Transl Med*. 2010;8:43. <https://doi.org/10.1186/1479-5876-8-43>  
PMid:20433757
157. Oleinick NL, Morris RL, Belichenko I. The role of apoptosis in response to photodynamic therapy: What, where, why, and how. *Photochem Photobiol Sci*. 2002;1(1):1-21. <https://doi.org/10.1039/b108586g>  
PMid:12659143
158. Zhu TC, Finlay JC. The role of photodynamic therapy (PDT) physics. *Med Phys*. 2008;35(7):3127-36. <https://doi.org/10.1118/1.2937440>  
PMid:18697538
159. Postiglione I, Chiaviello A, Palumbo G. Enhancing photodynamic therapy efficacy by combination therapy: Dated, current and oncoming strategies. *Cancers (Basel)*. 2011;3(2):2597-629. <https://doi.org/10.3390/cancers3022597>  
PMid:24212824
160. Misiewicz K, Skupińska K, Graczyk A, Kasprzycka-Guttman T. Influence of protoporphyrin IX amino acid substituents on affinity to human breast adenocarcinoma MCF-7 cells. *Biotechnic Histochem*. 2009;84(1):17-23.
161. Morgan J, Oseroff AR. Mitochondria-based photodynamic anti-cancer therapy. *Adv Drug Deliv Rev*. 2001;49(1-2):71-86. [https://doi.org/10.1016/s0169-409x\(01\)00126-0](https://doi.org/10.1016/s0169-409x(01)00126-0)  
PMid:11377804
162. Merlin JL, Gautier H, Barberi-Heyob M, Teiten MH, Guillemain F. The multidrug resistance modulator SDZ-PSC 833 potentiates the photodynamic activity of chlorin e6 independently of P-glycoprotein in multidrug resistant human breast adenocarcinoma cells. *Int J Oncol*. 2003;22(4):733-9.
163. Li Y, Yu Y, Kang L, Lu Y. Effects of chlorin e6-mediated photodynamic therapy on human colon cancer SW480 cells. *Int J Clin Exp Med*. 2014;7(12):4867-76.
164. Kessel D, Poretz RD. Sites of photodamage induced by photodynamic therapy with a chlorin e6 triacetoxymethyl ester (CAME). *Photochem Photobiol*. 2000;71(1):94-6. [https://doi.org/10.1562/0031-8655\(2000\)071<0094:sopibp>2.0.co;2](https://doi.org/10.1562/0031-8655(2000)071<0094:sopibp>2.0.co;2)  
PMid:10649895
165. Kessel D, Woodburn K, Gomer CJ, Jagerovic N, Smith KM. Photosensitization with derivatives of chlorin p6. *J Photochem Photobiol B*. 1995;28(1):13-8. [https://doi.org/10.1016/1011-1344\(94\)07085-3](https://doi.org/10.1016/1011-1344(94)07085-3)  
PMid:7791001
166. Broekgaarden M, Weijer R, van Gulik TM, Hamblin MR, Heger M. Tumor cell survival pathways activated by photodynamic therapy: A molecular basis for pharmacological inhibition strategies. *Cancer Metastasis Rev*. 2015;34(4):643-90. <https://doi.org/10.1007/s10555-015-9588-7>  
PMid:26516076
167. Dang J, He H, Chen D, Yin L. Manipulating tumor hypoxia toward enhanced photodynamic therapy (PDT). *Biomater Sci*. 2017;5(8):1500-11. <https://doi.org/10.1039/c7bm00392g>  
PMid:28681887
168. van Straten D, Mashayekhi V, de Bruijn HS, Oliveira S, Robinson DJ. Oncologic photodynamic therapy: Basic principles, current clinical status and future directions. *Cancers (Basel)*. 2017;9(2):19. <https://doi.org/10.3390/cancers9020019>  
PMid:28218708
169. Elmore S. Apoptosis: A review of programmed cell death. *Toxicol Pathol*. 2007;35(4):495-516. <https://doi.org/10.1080/01926230701320337>  
PMid:17562483
170. Moon YH, Kwon SM, Kim HJ, Jung KY, Park JH, Kim SA, *et al*. Efficient preparation of highly pure chlorin e6 and its photodynamic anti-cancer activity in a rat tumor model. *Oncol Rep*. 2009;22(5):1085-91.