



The Effect of Glutamine Supplementation to Fecal Calprotectin and Tumor Necrosis Factor- α Levels in the Fecal of Rats with Acute-Induced Diarrhea

Deddy Satriya Putra¹, Fadil Oenzil², Eryati Darwin³, Hafni Bachtiar⁴

¹Doctoral Student of Postgraduate Biomedical Science, Faculty of Medicine, Andalas University, Padang, Indonesia; ²Department of Nutritional Science, Faculty of Medicine, Andalas University, Padang, Indonesia; ³Department of Histological Science, Faculty of Medicine, Andalas University, Padang, Indonesia; ⁴Department of Public Health, Faculty of Medicine, Andalas University, Padang, Indonesia

Abstract

BACKGROUND: Glutamine is a non-essential amino acid as the main fuel in the gastrointestinal mucosa. By its various gastrointestinal functions, glutamine is thought to increase the protection of the intestinal mucosa against local or systemic injury from diarrhea.

AIM: This study aimed to determine the relationship between glutamine supplementation and fecal calprotectin and tumor necrosis factor- α (TNF- α) in the fecal of rats with acute and chronic diarrhea induced by EPEC.

MATERIALS AND METHODS: The research was conducted in an experimental laboratory with a randomized post-test only control group design. A total of 30 *Rattus norvegicus* strain Wistar were divided into five groups. The treatment group was induced to have diarrhea using EPEC at a dose of 108 CFU/ml, after each group diagnosed by acute and chronic diarrhea, followed by glutamine supplementation at a dose of 810 mg/200 g for 14 days. The inflammatory cytokines which is fecal calprotectin and TNF- α , of each group was assessed and the p value was measured by one-way ANOVA and *post hoc* Bonferroni test to find out the relationship between these variables.

RESULT: Supplementation of glutamine for 14 days can reduce fecal calprotectin levels in chronic diarrhea and reduce TNF- α level in rats with acute and chronic diarrhea.

CONCLUSION: Glutamine supplementation has an effect on the fecal calprotectin and TNF- α of rats with acute and chronic diarrhea.

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***Correspondence:** Deddy Satriya Putra, Doctoral Student of Postgraduate Biomedical Science, Faculty of Medicine, Andalas University, Padang, Indonesia. E-mail: dsatriyap@yahoo.com
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Introduction

Diarrhea is a condition in which stools are passed with a watery consistency that usually occurs with a frequency of more than 3 times/day [1]. Diarrhea is a major cause of morbidity and mortality in children worldwide, where 98% of deaths occur in developing countries, including Indonesia [2], [3]. Results of the 2013 Indonesian Basic Health Research stated in Riau Province, diarrhea was the second most common cause of toddlers' mortality, accounting for 17.2% of all toddler deaths [4].

Glutamine is a non-essential amino acid with an abundant amount in the human body. In the gastrointestinal tract, glutamine is known as the main fuel of the intestinal mucosa that triggers enterocyte growth, increases intestinal barrier function, and increases blood circulation to the intestine, plays an important role in the synthesis of nucleic and amino acids in the intestinal barrier, maintains integrity of the tight junction, and acts as an immunomodulator. Glutamine

is produced by the body in sufficient quantities, but its use tends to increase during illness including diarrhea, which causes glutamine depletion. This condition will interfere with the functions of glutamine and aggravate an illness. Diarrhea will be followed by damage to the intestine, and glutamine deficiency will inhibit its repair. Therefore, the supplementation of exogenous glutamine is needed to help the regeneration of damaged intestinal mucosa [5].

The benefits of glutamine as an adjuvant in managing diarrhea have been obtained based on several studies. A study by Radha and Samak and Vicario *et al.* and Carneiro-Filho *et al.* showed that glutamine has protective properties for mucosal barriers because of its effects on maintaining the integrity of intestinal epithelial tight junction cells [6], [7], [8]. In addition, glutamine could also facilitate enteral absorption of nutrients and electrolytes in diarrhea and reduce the severity of diarrhea by increasing water and salt uptake [9].

Based on a previous study, glutamine can improve intestinal growth, promote enterocyte proliferation and survival, and regulate the intestinal

barrier function during injury, infection, stress, and catabolic conditions. This effect occurs because glutamine can maintain intracellular redox status and regulate gene expression associated with various signaling pathways [10]. Research from Roedi Irawan also supported that glutamine could repair ileal microvilli in malnourished rats. In addition, an increase in intestinal enzyme activity, namely, sucrase, maltase, lactase, spectrin, as well as clathrin was found. This growing evidence supports the idea that glutamine is beneficial for intestinal repair [11]. Furthermore, a study by Huang showed the beneficial of glutamine on improvement of *Rattus norvegicus* strain Wistar's intestinal that is experiencing chronic diarrhea [12]. However, different results are shown from study by Kamucakhi about the benefits of glutamine supplementation in children who experience chronic diarrhea, in which glutamine supplementation showed no useful results [13]. This study aimed to determine the relationship between glutamine supplementation and fecal calprotectin, sIgA, TNF- α in the fecal of rats with acute and chronic diarrhea induced by EPEC.

Materials and Methods

This study was conducted in an experimental laboratory study using a randomized post-test only control group design.

Experimental samples

The sample of this study consisted of male rats (Wistar strain of *Rattus norvegicus*) aged 10–16 weeks, with a body weight of 160–250 g. The samples were divided into five groups with six rats/group: Negative control group (K), acute diarrhea group (A), acute diarrhea group supplemented with glutamine (Kyowa Hakko Bio Co., Made in Japan) at a dose of 810 mg/200 g rat (B), chronic diarrhea group (C), and chronic diarrhea group supplemented with glutamine at a dose of 810 mg/200g rat (D). The dose of glutamine given to the rats was based on the human-to-animal dose conversion factors. The dose in humans is 45 mg/day at a body weight of 70 kg, converted into 0.018 mg/day for rat.

Experimental design

Rats are placed in plastic cages with a lid made of ram wire and are given a filter paper mat to observe rats faces. The pad is replaced 3 times a week. The cage is placed in a well-ventilated room with good air circulation. The environment of cage is made so that it is not humid, the temperature of the cage is maintained according to room temperature

(20–26°C), and the exchange of dark and light every 12 h. Each rat is placed in its own cage. All rats are giving food and drink is given *ad libitum* and monitored its health every day. Acute diarrhea was induced in Groups A and B using EPEC (2 ml with a population of 108 CFU/ml). Chronic diarrhea was induced in Groups C and D using EPEC (2 ml with a population of 108 CFU/ml, given doses of repetition every 3 days). Diarrheic rats in Groups A and C were given the standard feeding, while those in Groups B and D were given glutamine for 14 days at a dose of 810 mg/200 g/day as soon after diagnosed by acute and chronic diarrhea. On the 15th day after standard feeding and glutamine supplementation, glutamine is given at a dose of 810 mg/200 g which is dissolved with aqua as much as 4 ml orally using sonde every day for 14 days. The rats' feces were taken from the ileum, weighing 15 mg for 100 μ l PBS.

Fecal calprotectin analysis

Fecal calprotectin levels were analyzed according to the instruction manual on the rat calprotectin ELISA Kit. Feces/stool taken from the caecum is centrifuged for 20 min at a speed of 5000 rpm. Supernatants are carefully taken, the sample can be stored at -20°C or -80°C. Before the examination is carried out, the sample is placed at room temperature (18°C–25°C) for 30 min. Next prepare the standard well, sample well, and blank well. Standard solutions, samples, and blanks of 100 μ l were put into each well on the plate, and then the plate was closed and incubated at room temperature for 1 h. The solution is well aspirated, and then the plate is washed up to three times with wash solution. Then, 50 μ l of chromogen Reagent A solution is added to each well and re-incubated for 15 min at room temperature, keeping the plate closed in the dark. Next 50 ml of chromogen Reagent B solution was added and incubated for 15 min at room temperature. The plates are washed 5 times, then 90 μ l substrate solution (Tetramethylbenzidine) is added to each well and incubated for 10 min at room temperature in a dark room (the liquid will turn blue). After 10 min, 50 μ l of stop solution (sulfuric acid) is added to each well (the liquid will turn yellow), then read on a microplate reader with a wavelength of 450 nm.

Fecal TNF- α level analysis

TNF- α levels were analyzed according to the instruction manual on the TNF- α ELISA Kit Rat. Stool taken from the caecum is centrifuged for 20 min at a speed of 5000 rpm. Supernatants are carefully taken, the sample can be stored at -20°C or -80°C. Before the examination is carried out, the sample is placed at room temperature (18°C–25°C) for 30 min. Next prepare the standard well, sample well, and blank well. Standard solutions, samples, and blanks of 100 μ l were

put into each well on the plate, and then the plate was closed and incubated at room temperature for 1 h. The solution is well aspirated, and then the plate is washed up to 3 times with wash solution. Then, 50 μ l of the detection Reagent A solution is added to each well and re-incubated for 15 min at room temperature, keeping the plate closed in the dark. Next 50 μ l of detection Reagent B solution was added and incubated for 15 min at room temperature. The plates are washed 5 times, then 90 μ l substrate solution (Tetramethylbenzidine) is added to each well and incubated for 10 min at room temperature in a dark room (the liquid will turn blue). After 10 min, 50 μ l of stop solution (sulfuric acid) is added to each well (the liquid will turn yellow), then read on a microplate reader with a wavelength of 450 nm.

Statistical analysis

Statistical analysis was performed with SPSS version 19. The sample was tested to determine different effect of glutamine on fecal calprotectin and TNF- α in acute and chronic diarrhea rats, one-way ANOVA test was used, if it was obtained $p < 0.05$ followed by *post hoc* Bonferroni test.

Research ethics requirements

This study has obtained ethical approval from the Faculty of Medicine, University of Riau (15.3/UN19.5.1.1.8/UEPKK/2018).

Results

Fecal calprotectin and TNF- α were assessed on day 15 after glutamine supplementation, in the K group (negative control), A group (rats with acute diarrhea not given glutamine), B group (rats with acute diarrhea given glutamine supplementation), C group (rats with chronic diarrhea not given glutamine), and D group (rats with chronic diarrhea given glutamine supplementation). Furthermore, an analytical test using the one-way ANOVA method is presented in Table 1.

Table 1: Effect of glutamine administration on the mean levels of fecal calprotectin in rats on acute diarrhea group

Group	n	Mean (SD) μ g/ml	p
K	6	62.379 (8.45)	0.107
A	6	74.723 (11.76)	
B	6	72.130 (9.11)	

K: Control, A: Acute diarrhea, B: Acute diarrhea + glutamine

Table 1 shows that there were differences in the mean levels of fecal calprotectin between rats in Groups K, A, and B. The rates of fecal calprotectin increased in rats in Group A, namely, acute diarrhea rats (74.723 μ g/ml) compared with rats in Group B, namely, acute diarrhea rats with glutamine administration

(72.130 μ g/ml), and are higher than the average level of fecal calprotectin in Group K rats, namely, the control group rats (62.379 μ g/ml). However, statistically obtained $p > 0.05$, so it can be concluded that there is no significant difference from the average fecal calprotectin levels in acute diarrhea group rats.

Table 2 shows that there were no significant differences in the fecal calprotectin levels of Groups K and A rats with $p > 0.05$. There were no significant differences in the fecal calprotectin levels in Groups K and B rats as well as in rats Group A and B with $p > 0.05$. It can be concluded that the difference in fecal calprotectin levels in each acute diarrhea group was not statistically significant.

Table 2: Effect of glutamine administration on fecal calprotectin levels in each rats on acute diarrhea group

Group	K	A	B
K	-	0.141	0.324
A	0.141	-	1.000
B	0.324	1.000	-

K: Control, A: Acute diarrhea, B: Acute diarrhea + glutamine

Table 3 shows that there were differences in the mean TNF- α levels between Group K, A, and B rats. There was an increase in TNF- α mean in Group A (223.878 μ g/ml) and Group B (229.382 μ g/ml) compared to Group K rats (161.358 μ g/ml). Group A is lower than Group B. Statistically obtained $p < 0.05$ so that it can be concluded that there is a significant difference in the mean TNF- α levels in rats in the acute diarrhea group. Furthermore, to find out the significant differences in each group, an analysis using *post hoc* Bonferroni is presented in Table 4.

Table 3: Effect of glutamine administration on the mean levels of TNF- α in the rats on acute diarrhea group

Group	n	Mean (SD) μ g/ml	p
K	6	161.358 (26.68)	0.016
A	6	223.878 (52.29)	
B	6	229.382 (34.69)	

K: Control, A: Acute diarrhea, B: Acute diarrhea + glutamine

Table 4 shows that there were significant differences in the TNF- α levels of rats in Groups K and A and in rats in Groups K and B with $p < 0.05$. There was no significant difference in TNF- α levels in rats Group A and B with $p > 0.05$. It can be concluded that the differences in the TNF- α levels of rats in Groups K and A and Groups K and B are statistically significant.

Table 4: Effect of glutamine administration on TNF- α levels in each rats on acute diarrhea group

Group	K	A	B
K	-	0.045*	0.027*
A	0.045*	-	1.000
B	0.027*	1.000	-

K: Control, A: Acute diarrhea, B: Acute diarrhea + glutamine

Discussion

In this study was shown an increase in levels of fecal calprotectin in rats in the acute diarrhea group and acute diarrhea with glutamine. This shows the

occurrence of inflammation in the intestinal cells of experimental rats induced by EPEC. Fecal calprotectin can be used as a biomarker for inflammation of intestinal cells and increases significantly in diarrhea due to bacterial infection. High fecal calprotectin levels also correlate with the clinical severity of infectious diarrhea in children, the more severe the clinical, and the higher the fecal calprotectin; therefore, fecal calprotectin can be used as a marker for inflammation in the intestine and as an evaluation of therapeutic management [14].

Although in this study the levels of fecal calprotectin in acute diarrhea rats given glutamine were lower than those in diarrhea rats that were not given glutamine, which suggests that glutamine might have the effect of reducing inflammation, statistically this was not significant. The previous studies conducted by Vidal-Casariogot (2013), also found no significant difference in calprotectin levels after glutamine administration to prevent enteritis in patients on radiotherapy [15], although the pathophysiology of the study was different.

Calprotectin is a protein complex found in mammals. In the presence of calcium, calprotectin can inhibit the transition from manganese and zinc through the mechanism of chelation, this mechanism (inhibiting the transition from manganese and zinc) is an attempt to kill bacteria in innate immunity; therefore, calprotectin is known to have an antimicrobial function. Calprotectin is mostly (up to 60%) present in dissolved proteins in cytosols and neutrophils, although it can be found in monocytes, macrophages, and squamous epithelial cells [16]. Secretion from calprotectin will increase in the inflammatory response. Calprotectin can be found in the intestinal lumen through several mechanisms including increased effects of leukocytes, active secretion, cell disruption, and cell death. As a result, there is an increase in fecal calprotectin levels, which can be detected in feces. An increase in fecal calprotectin indicates there is migration of neutrophils into the intestinal mucosa, due to intestinal inflammation [17].

The results of this study indicate that statistically there are significant differences in the TNF- α group of acute diarrhea. These results differ from the previous studies conducted by Ameho in 1997, which showed glutamine can suppress TNF- α in colitis-induced rats, where the average TNF- α concentration in the group given glutamine was 4% and 2% lower than in the control group, the concentration this low level indicates the role of glutamine in inhibiting the synthesis, release, and action of pro-inflammatory cytokines which have an impact on the clinical improvement of the disease [18].

This average difference might occur because there are different glutamine doses, in which in this study a dose of 810 mg/200 g of rats (4.05 g/kg) was used, whereas the study from Ameho used a higher dose of 40 g/kg/day and a dose of 20 g/kg/day. Research from Ameho also shows better results from groups of rats with higher doses than lower ones [18]. Research from

Yalcin in 2004, with low doses (0.3 g/kg) also found no benefit of glutamine immunologically [19].

In this study also found levels of TNF- α in Groups A and B there were no statistically significant differences. This was also found in Ameho, where in the control group and the treatment group, although there were clinical differences, the difference in TNF- α was not statistically significant. Research from Ameho has not yet found an explanation for this, however, it is well known that the serum concentration of TNF- α is not always consistent with the severity of the disease being low [18].

The results of this study are consistent with the previous studies conducted by Ameho in 1997, which showed glutamine can suppress TNF- α in colitis-induced rats, where the average TNF- α concentration in the group given glutamine was 4% and 2% lower than the control group, this low concentration indicates the role of glutamine in inhibiting the synthesis, release, and action of pro-inflammatory cytokines, which have an impact on the clinical improvement of the disease [18].

In the early stages of infection, activation of the inflammatory reaction occurs. NF- κ B, which is a multiprotein complex, composed of p50, p52, p65, RelB, and c-Rel. In good health, NF- κ B is stored in the cytoplasm and remains inactive with the help of an inhibitor of κ B (I- κ B). In response to extracellular stimulus, I- κ B kinase performs phosphorylation of the I- κ B protein, resulting in I- κ B degradation which causes the release and activation of NF- κ B. This active NF- κ B complex then moves into the nucleus and induces expression of the NF- κ B-binding element gene, such as IL-6 and TNF- α . In this inflammatory state, production of IL-6 and TNF- α increases rapidly, which stimulates immune responses, such as antigen-presenting cells and T-cells [10].

Glutamine predicted to inhibit these pathways with multimodal. There are several mechanisms by which glutamine can suppress TNF- α , including inhibition of I κ B α -ubiquitination, inhibition of activation of NF- κ B, which has an impact on suppressing the production of IL-6, IL-8, and TNF- α , and reducing regulation of signal transducers and activators of transcription (transcription). The previous studies have shown that glutamine supplementation can be a promising candidate for treating this inflammatory disease [10]. Supplementation of glutamine for 14 days can reduce fecal calprotectin levels in chronic diarrhea and reduce TNF- α level in rats with acute diarrhea.

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