Beet *(Beta vulgaris)* Improve Blood Glucose and AKT2 Gene Expression in High Fat and Fructose-induced Rats

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**Abstract**

**BACKGROUND:** Diet components significant effects on glucose homeostasis. A diet contains high saturated fat and fructose induces insulin resistance and enhanced blood glucose. In contrast, food containing flavonoids such as beet can improve glucose homeostasis via modulation of gene expression, for example, AKT2, involving glucose metabolism.

**AIM:** This study was to evaluate the benefit of beet on AKT2 gene expression and fasting glucose (FG).

**METHODS:** Twenty Wistar male was divided into five groups: Normal were fed a normal diet, group HFFD was given a diet containing high fat and fructose, and three groups (HFB1, HFB2, HFB3) were given a diet containing high fat and fructose for 8 weeks and continuous fed beet-contained normal diet for 6 weeks. The percentage of beet in the diet for each 6%, 9%, and 12%, respectively.

**RESULTS:** The FG was measured before and after the intervention, whereas the gene expression of AKT2 at the trancript level for high fat and fructose diet, and the expression of the AKT2 gene may have a role in the process.

**CONCLUSIONS:** The beet 9% substituted diet can improve glucose homeostasis from the effects of a high fat and fructose diet, and the expression of the AKT2 gene may have a role in the process.

**Introduction**

The composition of the diet has significant and clinically relevant effects on circulating glucose, and it is influenced by food components, such as form, kind, and amount. High saturated fat and high fructose have been known can induce insulin resistance in tissues and enhanced blood glucose. The rats fed a diet containing high fat or high fat and also a high fructose diet had fasting blood glucose (FBG) concentration higher than those in the rats control diet [1]. On the other hand, a high polyphenol diet can influence gene expression involved in insulin signaling, insulin secretion, and hepatic glucoseogenesis pathways. Many polyphenols were reported to influencing the expression of insulin receptors substrate 1 (IRS1), serine/threonine-protein kinase 1 (AKT1), and phosphoenolpyruvate carboxykinase in human hepatic cells (HEPG2) [2]. In addition, various polyphenols have been shown antioxidant properties that have biological effects through the starting responses of cell signaling and interplay with both extracellular and intracellular receptors. The interaction between polyphenol and cell membrane can induce changing of membrane function, for example., stimulation of signal transduction [3].
that the AKT2 has an important role in a pathway that directly integrates glucose, GLUT1 expression, and glucose availability to maintain the viability of AKT2-dependent cells [10].

**Methods**

**Animals and experimental studies**

Approval of the present research was obtained from the Ethical Committee of Integrated Research and Testing Laboratory, Universitas Gadjah Mada (Approval number: 00011/04/LPPT/V/2019). Wistar male rats at the age of 1 month (n = 20), weighing ±150 g, were obtained from Pharmacy Faculty, Universitas Gadjah Mada. Each rat was kept in cages in a room with a temperature of 25°C and the cycle of light/dark:12:12 h. The rats were acclimated for seven days using AIN93M formulation and water ad libitum and then were randomly assigned to five groups consisted of four rats for each group. Group N was fed a normal diet, group HFFD was fed a diet containing high fat and high fructose, and three groups (HFB1, HFB2, and HFB3) were fed a diet containing high fat and high fructose for eight weeks and continuously fed a beet-contained normal diet for 6 weeks. The substitution percentage of beet in the diet for each 6%, 9%, and 12%, respectively. All rats were given distilled water ad libitum during the study. The formulation of the diet showed in Table 1.

**Table 1: Diet composition**

<table>
<thead>
<tr>
<th>Substance (g/kg diet)</th>
<th>Standard diet (N)</th>
<th>High fat and fructose diet (HFFD)</th>
<th>Intervention diet with 6% beet</th>
<th>Intervention diet with 9% beet</th>
<th>Intervention diet with 12% beet</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cornstarch</td>
<td>121</td>
<td>300</td>
<td>561</td>
<td>531</td>
<td>501</td>
</tr>
<tr>
<td>Casein</td>
<td>140</td>
<td>140</td>
<td>140</td>
<td>140</td>
<td>140</td>
</tr>
<tr>
<td>Sucrose</td>
<td>100</td>
<td>-</td>
<td>100</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Trans fat</td>
<td>-</td>
<td>214</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Fructose</td>
<td>-</td>
<td>250</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Beta actin</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Alfalfa (Cell)</td>
<td>50</td>
<td>50</td>
<td>50</td>
<td>50</td>
<td>50</td>
</tr>
<tr>
<td>Mineral mix (AIN'93M-MX)</td>
<td>35</td>
<td>35</td>
<td>35</td>
<td>35</td>
<td>35</td>
</tr>
<tr>
<td>Vitamin mix (AIN'93-VX)</td>
<td>10</td>
<td>10</td>
<td>10</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>Methionine</td>
<td>1.8</td>
<td>1.8</td>
<td>1.8</td>
<td>1.8</td>
<td>1.8</td>
</tr>
<tr>
<td>Choline chloride</td>
<td>2.5</td>
<td>2.5</td>
<td>2.5</td>
<td>2.5</td>
<td>2.5</td>
</tr>
<tr>
<td>Terti-butylhydroquinone</td>
<td>0.008</td>
<td>0.008</td>
<td>0.008</td>
<td>0.008</td>
<td>0.008</td>
</tr>
</tbody>
</table>

**Laboratory analyses**

The levels of fasting glucose (FG) were measured before and after the intervention, whereas the gene expression of AKT2 at skeletal muscle tissue was determined after the intervention. The serum obtained from fasting (8–10 h without diet) whole blood that taken from sinus orbitalis, enter the tube and stay for 2 h at room temperature, and then centrifuged at 3000 rpm for 15 min, the top layer solution is serum. The rats were anesthetized to get skeletal muscle tissue using intraperitoneal injection of ketamine and sacrificed. The levels of serum glucose were measured by the colorimetric method according to the protocol in glucose Diasys Kit. The steps of analysis of AKT2 gene expression using quantitative polymerase chain reaction (qPCR) are as follows: The total RNA of rats skeletal muscle was isolated with a TriRNA (Favorgen) based on instructions manufacturer. The RNA concentration was determined by Nanodrop, and the synthesis of cDNA using kit from Revertaid First Strand cDNA Synthesis Kit (Thermo scientific). The gene expression of AKT2 was analyzed using SSoAdvanced Universal SYBR Green Supermix (Biorad). Primer pairs are listed in Table 2.

**Table 2: Primers of reverse transcription PCR analysis**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequence</th>
<th>Product (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td>AKT2</td>
<td>F : 5’GGAGGTCATGGAGCATCGGTTC3’</td>
<td>80</td>
</tr>
<tr>
<td>Beta actin</td>
<td>R : 5’GTTTGAAGGGTGCCAGCGAC3’</td>
<td>149</td>
</tr>
</tbody>
</table>

**Statistical analysis**

The results are presented as mean±standard deviation. The FBG and AKT2 gene expression were determined by one-way ANOVA and next to a post hoc multiple comparison test. Comparative analysis of pretest-posttest data using paired sample t-test.

**Results**

Changes in the levels of blood glucose of fasting rats after received a diet high in fat and fructose (before rats get a beet-contained diet) and after given a beet-contained diet were presented in Table 3. The HFFD, HFB1, HFB2, and HFB3 groups had FBG levels significantly higher than those in the N groups (p < 0.05). The beet 9%-contained diet can significantly reduce the FBG levels.

**Table 3: The mean of the levels of blood glucose of fasting rats after induction and after intervention**

<table>
<thead>
<tr>
<th>Groups</th>
<th>N</th>
<th>Average of FBG levels (mg/dL)</th>
<th>ΔFBG</th>
<th>P**</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>After induction HFFD</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HFFD</td>
<td>4</td>
<td>109.33 ± 8.22</td>
<td>-1.13</td>
<td>0.824</td>
</tr>
<tr>
<td>HFB1</td>
<td>4</td>
<td>132.25 ± 3.60</td>
<td>131 ± 10.03</td>
<td>-1.25</td>
</tr>
<tr>
<td>HFB2</td>
<td>4</td>
<td>129.5 ± 6.24</td>
<td>117 ± 8.29</td>
<td>-12.50</td>
</tr>
<tr>
<td>HFB3</td>
<td>4</td>
<td>133 ± 8.29</td>
<td>104.33 ± 5.44</td>
<td>-28.68</td>
</tr>
<tr>
<td></td>
<td></td>
<td>After beet intervention</td>
<td></td>
<td></td>
</tr>
<tr>
<td>HFFD</td>
<td>4</td>
<td>131.25 ± 4.03</td>
<td>127 ± 3.74</td>
<td>4.25</td>
</tr>
<tr>
<td>HFB1</td>
<td>4</td>
<td>130 ± 6.24</td>
<td>117 ± 8.29</td>
<td>-12.50</td>
</tr>
<tr>
<td>HFB2</td>
<td>4</td>
<td>133 ± 8.29</td>
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<td>-28.68</td>
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<tr>
<td>HFB3</td>
<td>4</td>
<td>131.25 ± 4.03</td>
<td>127 ± 3.74</td>
<td>4.25</td>
</tr>
</tbody>
</table>

*Normal control group; HFFD: Rats received high fat and fructose; HFB1: HFFD fed by beet 6%-contained diet; HFB2: HFFD fed by beet 9%-contained diet; HFB3: HFFD get beet 12%-contained diet. Data are presented in mean±standard deviation. Significance at a: P < 0.05 according to One Way ANOVA and Tukey test. There is no difference in either a or b: P**: Difference of FBG levels between groups; P**: Difference of FBG levels before and after beet intervention at same groups; Sign # shows that significant: FBG: Fasting blood glucose.

There was no significant correction between the expression of the AKT2 gene and the FG because...
glucose metabolism involved many pathways, such as
the previous study (Figure 1). This study showed that
the effect of beet on AKT2 gene expression was not
significant, although the AKT2 gene expression in the
HFFD group was lowest, but the AKT2 gene expression
in the HFB2 group closely to those in the N group
(Figure 2).

Discussion

This present study showed that the rats in HFFD, HFB1, HFB2, and HFB3 groups had FG between 130,135 g/dL, and it more than the normal group (N). According to the previous studies that the levels of blood glucose of fasting rats fed a diet containing high fat and fructose for six weeks were higher than rats fed the standard diet [11]. Haroun et al. [12] also reported that rats fed a diet containing high fat only or fructose alone for 5 weeks had higher blood glucose levels than normal control rats. Another study showed that the combination of a high-fat diet and high fructose beverages induced fasting hyperglycemia after 6 months [13]. In some studies, using animals showed that high-fat diet impaired glucose tolerance that was associated with decreased basal and insulin-stimulated glucose metabolism, whereas, in the human, a high-fat diet reduced insulin sensitivity. In this study, we used saturated fat, and saturated fat is more deleterious with respect to fat-induced insulin insensitivity than monounsaturated and polyunsaturated fat [14]. Insulin insensitivity or insulin resistance is also reported to be associated with high fructose consumption. Fructose-induced insulin resistance can be mediated by an increase of reactive oxygen species that mediates a proinflammatory cascade to lead to an increase of adipogenesis, release of inflammatory cytokines, and decrease in adiponectin cause insulin resistance [15], [16].

After getting a beet-contained diet, the FBG in HFB1, HFB2, and HFB groups decreased, especially in the HFB2 group that gets a beet 9% contained diet, seen significantly. It showed that beet 9% as a physiological dose having the best effects to maintain glucose homeostasis. The result of this present study, in accordance with the previous research by Lorizola et al. [17] reported the rats treated with the stem and leaf parts of beets had a decrease in blood glucose levels compared to the untreated rat group. The result of this study according with the study Gezginci-Oktayoglu that reported that extract of B. vulgaris L. var. cicla can increase GLUT2 via AKT2 and defence of antioxidant in the liver lead to improve hyperglycemia [18]. AKT is involved in the metabolism regulation of glucose and the pathway of intracellular insulin transduction and metabolism of energy in the liver [19].

The AKT2 was reported to modulate the availability of glucose through expression regulation of GLUT1 at the level of transcript and abrogating expression of AKT2 impaired glucose uptake by the cell [10]. In the present research, the effect of beet-substituted diet on the gene expression of AKT between groups was not statistically significantly different, but the rats get beet substituted diet indicated AKT gene expression higher than those in the HFFD rat group, and its expression closely to normal rat group was beet 9% substituted diet (Figure 2). It suggested that substitution of beet 9% in the diet can meet the physiological need to improve the effects of a diet containing high fat and fructose related to glucose homeostasis. In low doses (beet 6%) may allow less effect on ligands, whereas a higher dose (beet 12%) over a long period of time may trigger prooxidant activity. Flavonoids given in high amounts can act as prooxidants. Fang et al. [20] reported that flavonoids become prooxidant activity occurs by triggering OH in the presence of Cu + and H2O2. In addition, the expression of the AKT2 gene has a negative correlation with FG, but it was not statically significant. It is because glucose metabolism involved many pathways. Although the skeletal muscle AKT2 has
been believed to play an important role in homeostasis glucose, however, the mice’s skeletal muscle only lacking AKT2 showed cannot stimulate insulin resistance or inhibit the uptake of glucose. It showed that besides skeletal muscle AKT2, another signaling molecule might be involved in perturbation tolerance of glucose and sensitivity of insulin in vivo [10].

Conclusion

The beet 9% substituted diet can improve glucose homeostasis from the effects of a diet containing high fat and fructose, and the expression of the AKT2 gene may have a role in the process.

Acknowledgments

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References

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