









Urinary Phthalate Levels in Relation to Obesity among a Sample of Egyptian Children

Manal A. Mohsen¹ , Mai M. Youssef¹ , Ebtissam M. Salah El-Din¹ , Samar M. E. Salem¹ , Hala Mohamed Salah El Din Megahed¹ , Mones M. AbuShady¹ , Dalia Medhat² , Amr S. Gouda³ , Walaa Nazim³ , Manal A. Shehata^{1*} 

¹Department of Child Health, Medical Research and Clinical Studies Institute, National Research Centre, Giza, Egypt; ²Department of Medical Biochemistry, Medical Research and Clinical Studies Institute, National Research Centre, Giza, Egypt; ³Department of Biochemical Genetics, Human Genetics and Genome Research Institute, National Research Centre, Giza, Egypt

Abstract

Edited by: Ksenija Bogoeva-Kostovska
Citation: Mohsen MA, Youssef MM, Salah El-Din EM, Salem SME, Megahed HMSED, AbuShady MM, Medhat D, Gouda AS, Nazim W, Shehata MA. Urinary Phthalate Levels in Relation to Obesity among a Sample of Egyptian Children. Open Access Maced J Med Sci. 2022 Feb 06; 10(B):674-680. https://doi.org/10.3889/oamjms.2022.8259
Keywords: Phthalates; Children; Obesity; Cholesterol; Triglycerides; Anthropometry
***Correspondence:** Manal A. Shehata, Department of Child Health, Medical Research and Clinical Studies Institute, National Research Centre (ID: 60014618), Giza. E-mail: dr_manalabdalkader@hotmail.com
Received: 14-Dec-2021
Revised: 22-Jan-2022
Accepted: 27-Jan-2022
Copyright: © 2022 Manal A. Mohsen, Mai M. Youssef, Ebtissam M. Salah El-Din, Samar M. E. Salem, Hala Mohamed Salah El Din Megahed, Mones M. AbuShady, Dalia Medhat, Amr S. Gouda, Walaa Nazim, Manal A. Shehata
Funding: This work was supported by a grant from The National Research Center (NRC) of Egypt. This study was one of the accepted and granted researches among the projects of the eleventh plan of NRC, dated from 2016 to 2019. The project's title was: Evaluation of the potential effects of phthalate on children health in Egypt. Its number was: 11010147. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.
Competing Interests: The authors have declared that no competing interests exist
Open Access: This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (CC BY-NC 4.0)

BACKGROUND: Childhood obesity is considered a risk factor for chronic diseases later in life. Phthalates (phthalate acid esters), predominant constituents of plasticizers, are well-thought-out global environmental contaminants.

AIM: This study aims to investigate the relationship between obesity and urinary phthalates in Egyptian children.

MATERIALS AND METHODS: This cross-sectional study included 210 children; 71 children were obese. Age ranged between 8.8 and 16 years with a mean of 12.93 ± 1.29 years. Sociodemographic data were collected. Clinical examination included measuring body weight, height, waist and hip circumferences (WC and HC), and calculation of body mass index (BMI). The lipid profile was analyzed. Urine samples were tested for phthalates levels using high-performance liquid chromatography.

RESULTS: Urinary phthalates metabolites mono benzyl (MBzP), monobutyl (MBP), monoethyl (MEP), and mono (2ethylhexyl) phthalate (MEHP) were detected in all urinary samples with varying levels. The median concentrations of MBzP, MEHP, MBP, and MEP were 1.4, 54.5, 29.9, and 490 (ng/ml), respectively. In obese children, urinary MBP, MEP, and MEHP demonstrated significantly higher mean levels than in non-obese children. Physical indicators of obesity as body weight, BMI, WC, and HC were significantly positively correlated with urinary levels of MEHP and MEP, while urinary MBzP demonstrated a significant positive association with serum triglycerides levels.

CONCLUSION: The present study suggests an association between phthalates exposure and childhood and adolescent adiposity.

Introduction

Childhood and puberty are critical periods for adipose tissue mass building [1]. The worldwide prevalence of overweight and obesity in children has amplified over the past three decades to 124 million [2] and WHO predicts by 2025 an additional 70 million [3]. Obesity in children and adolescents does not only disturb their health and social life but may have resounding adverse effects during forthcoming adulthood [4]. People with obesity may suffer one or more chronic diseases such as Type 2 diabetes mellitus, hyperlipidemia, cardiovascular disease, liver and gall bladder diseases, stroke, hypertension, and specified types of malignancies [5].

Besides the modern rise in popularity of fast-food and physical inactivity, exposure to environmental pollutants has been linked to childhood obesity [6].

Exposure to endocrine-disrupting chemicals as phthalate acid esters throughout this period may have harmful consequences on adipose function and metabolism [7], [8], [9]. Phthalates are commonly used as plasticizers in household products. They are utilized in the production of widely-distributed personal tools such as children's toys, paints, cosmetics, and food packages [10]. Phthalates easily leach from their compounds and enter the human body through ingestion, inhalation, and dermal contact [11], [12]. Children have been suggested to be more susceptible to exposure, mostly due to diverse behavioral attitudes and higher consumption in relation to body size [13].

Phthalates association with several human diseases are currently an eminent subject in the literature [14]. The evidence on the association of phthalates with obesity and lipid metabolism is limited and inconsistent [15].

The present study aimed to investigate the relationship between obesity in Egyptian children and phthalates exposure. This study also assessed the associations of urinary phthalate with physical (body mass index [BMI], waist and hip circumferences [WC and HC]), and biochemical indices of obesity (total serum cholesterol [TC], low-density lipoprotein [LDL], high-density lipoprotein [HDL], and triglycerides [TG]).

Materials and Methods

This study was conducted at the National Research Centre (NRC) in Egypt, as a part of a research project funded by NRC [16]. It was a cross-sectional study, comprising 210 children of both sexes. Age ranged between 8 and 16 years. They were randomly selected from Giza governorate schools from October 2017 to April 2018. All were more or less of similar social class. Exclusion criteria included liver diseases, renal diseases, thyroid disorders, and endocrinal or genetic causes of obesity.

Ethical issues

Permission to implement the study was approved by the Ministry of Education and the director of the school of children included in the research. Ethical approval of The Research and Ethical Committee of the NRC was obtained (Number: 16368). Parents were informed about the purpose of the study and their written consents were obtained.

Sociodemographic history and clinical examination

The participants were asked to complete a structured questionnaire. Questions included sociodemographic variables and medical history. Children were subjected to thorough clinical examination that included chest, heart, abdominal, and neurological examination. Anthropometric measures after the recommendations of the International Biological Program were assessed.

Anthropometric measurements

Bodyweight (kg), height (cm), WC, and HC (cm) were measured. The height was measured to the nearest 0.5 cm using a Holtain portable anthropometer. The weight was measured to the nearest 0.1 kg using scale balance while the child was dressed in minimal clothes and without shoes. BMI was calculated as weight in kilograms divided by height in meters squared. WC was measured at the level of the umbilicus while the participant

standing and breathing normally. HC was measured at the level of the iliac crest using non-stretchable plastic tape to the nearest 0.1 cm. Each measurement was taken as a mean of three consecutive measurements using standardized equipment and following the recommendation of the International Biological Program. Children were considered obese when their BMI Z score was higher than +2 for age and gender according to the WHO Child Growth Standards [17].

Laboratory investigations

Determination of phthalates using high-performance liquid chromatography.

Phthalates are metabolized to their monoesters within a short period, urinary phthalate monoesters are considered valid biomarkers for evaluating human phthalates exposure [18], [19]. One spot urine sample is sufficient to represent exposure over 6 months to permit its usage as an exposure estimate in epidemiological research [8], [20], [21] To adjust for dilution, we included urinary creatinine as a covariate in all analyzes. All metabolite concentrations were adjusted for creatinine levels and expressed as $\mu\text{g/g Cr}$.

Phthalates metabolites were measured instead of their parent compounds to lower the potential for exposure misclassification. A grouping of solid-phase extraction, high-pressure liquid chromatography, and tandem mass spectrometry was used to estimate phthalates metabolite concentrations using methods defined by Koch *et al.* [18] Urine samples were thawed and sonicated for 10–15 min, then (100 μl) was loaded into a glass vial (2 ml) contained ammonium acetate (AA, 20 μl , >98%, Sigma Aldrich Laboratory, Inc., St. Louis, MO, USA), β -glucuronidase (10 μl , E.coli K12, Roche Biomedical, Mannheim, Germany), and a combination of ten isotopic $^{13}\text{C}_4$ phthalate metabolite standards (100 μl . Cambridge Isotope Laboratory, Inc., Andover, MA, USA). Afterward, the samples were incubated (37°C, 90 min), a 270 μl solution (5% ACN), Merck, Darmstadt, Germany) with 0.1% formic acid (FA, Merck, Darmstadt, Germany) was added and sealed with the PTEF cap for analysis.

Monoethyl phthalate, monomethyl phthalate, monobenzyl phthalate, monobutyl phthalate, and bis-2ethylhexylphthalate were purchased from Sigma-Aldrich. β -glucuronidase was purchased from Roche. Stock solutions of standards were prepared in acetonitrile (3000ng/ml). Eleven-point calibration curve was constructed for each standard (0.5, 1, 2, 4, 8, 16, 32, 64, 128, 256, and 1000 ng/ml).

Sample preparation was done by solid-phase extraction according to the method described by Feng *et al.* [22] After evaporation of eluting, the residue was reconstituted in 100 μl 20% acetonitrile in water and injected.

Analysis was done on Waters Xevo TQD – ESI

triple quadruple mass spectrometer. Separation was done on Phenomenex Kinetex (150*2.1 mm 2.6) μ m phenyl-hexyl using 0.1% acetic acid in water (mobile Phase A) and 0.1 % acetic acid in acetonitrile (mobile phase B) with a linear gradient described in Feng *et al.* [22] 10 μ l was injected. Negative ionization and MRM mode were used. Source temperature was 150°C, desolvation temperature was 200°C, and desolvation gas flow was 650 L/Hr.

Blood samples

Fasting venous blood samples were collected following an overnight fast (12–14 h). Sera were separated by centrifugation and stored at –80 until assays. TC and TG were measured by quantitative enzymatic colorimetric technique [23]. Serum HDL-cholesterol was measured by the phosphotungstate precipitation method [24]. Serum LDL was calculated using the Friedewald formula [25].

Statistical analysis

Statistical analysis was carried out using the statistical package for the social sciences, version 21 for Windows (IBM Corp., Armonk, NY, USA). Continuous data were expressed as minimum, maximum, and mean \pm SD and were compared using Student's t-test. A general linear model was used to control the confounding factors. Categorical data were expressed as frequencies and percentages. Pearson's correlation analysis was carried out to evaluate the association between continuous exposure and continuous covariates.

Results

This study included 210 school children, 118 boys, and 92 girls. Their age ranged between 8.8 and 16 years with a mean of 12.93 ± 1.29 years. Their anthropometric and the clinical data included the body weight (kg), height (cm), WC and HC (cm) and serum triglycerides (mg/dl), cholesterol (mg/dl), HDL (mg/dl), and LDL (mg/dl) are shown in Table 1.

Table 1: Age, anthropometric, and biochemical data of the studied children

	Minimum	Maximum	Mean	Std. Deviation
Age	8.80	16.00	12.9307	1.28654
Weight (kg)	20.00	109.00	54.5980	16.31442
Height (cm)	100.00	184.00	152.6422	12.31931
HAZ	-3.19	3.55	-0.3483	1.35522
BAZ	-3.66	4.58	1.0820	1.67432
Waist circumference (cm)	47.00	111.00	75.8241	13.08472
Hip circumference (cm)	54.00	131.00	89.8054	14.03400
Triglycerides (mg/dl)	67.80	311.90	97.2231	63.07140
Cholesterol (mg/dl)	114.40	243.70	187.4731	40.00857
HDL (mg/dl)	35.00	65.00	49.88	8.18451
LDL (mg/dl)	55.00	177.00	116.65	35.627

Table 2: Classification of school children according to the BMI z-score category

Classes of obesity	Frequency	Percentage
Children with obesity	71	34.8
No obesity	134	65.2
Total	205	100

One-third of children had a BMI $> +2$ z score (equivalent to >95 percentiles) of the WHO Standards and hence considered having obesity (Table 2). Phthalate metabolites monobenzyl (MBzP), monobutyl (MBP), monoethyl (MEP), and mono-(2-ethylhexyl) phthalate (MEHP) (ng/ml) were detected in all urine specimens. The mean concentrations measured 1.73, 30.37, 523.16, and 58.29 ng/ml, respectively. The geometric mean concentrations were 1.58, 29.71, 488.73, and 55.46 ng/ml, respectively and the median concentrations were 1.44, 29.9, 490, and 54.5 ng/ml, respectively (Table 3).

Table 4 shows the correlations between the phthalate metabolites, the anthropometric measurements, and the biochemical indices. The table demonstrates a negative non-significant correlation between both MEHP and MEP on one side and triglycerides and cholesterol on the other. Monobutyl showed a positive non-significant correlation with triglycerides and with cholesterol. Monobenzyl assessment gave a positive significant correlation with triglycerides. Positive significant correlations between both MEHP and MEP on one hand and body weight, BMI, WC, and HC, on the other hand, were valid. Negative non-significant relations between children's height and the mean concentrations of the urinary MBzP, MBP, and MEP (ng/ml) were separately noted.

Table 3: Phthalate metabolite values expressed as median, mean \pm SD, and geometric mean

Phthalate metabolite	Minimum	Maximum	Median	Mean	SD	Geometric Mean
Monobenzyl (ng/ml)	0.64	4.11	1.44	1.73	0.77	1.5763
Monobutyl (ng/ml)	19.7	47.80	29.9	30.37	4.94574	29.7145
Monoethyl (ng/ml)	145	1321	490	523.16	205.97	488.73
Mono-(2-ethylhexyl) phthalate (ng/ml)	28.4	120.8	54.5	58.28	19.26	55.46

The urine of children with obesity had higher significant concentrations of MBP, MEP, and MEHP than the urine of normal-weight children (Table 5).

Discussion

Association between phthalates exposures and obesity in adults and children has been pointed out in the previous literature with conflicting results [26], [27].

The extensive phthalates exposure of humans was demonstrated through quantifying phthalates metabolites in urine samples, where the levels of urinary phthalates metabolites indicate recent exposure [28]. In this study, low molecular weight metabolites MBzP, MBP, MEP, and high molecular weight metabolites (MEHP)

Table 4: Correlation between phthalate metabolites and anthropometric measurements and biochemical indices

	Triglycerides (mg/dl)	Cholesterol (mg/dl)	HDL (mg/dl)	LDL (mg/dl)	Weight (kg)	Height (cm)	BAZ	Waist circumference (cm)	Hip circumference (cm)
MEHP (ng/ml)									
R	-0.148	-0.020	0.419*	-0.105	0.247**	0.060	0.263**	0.202*	0.252**
p	0.499	0.927	0.047	0.635	0.002	0.465	0.001	0.013	0.002
Monoethyl (ng/ml)									
r	-0.119	-0.134	0.107	-0.150	0.215**	-0.025	0.274**	0.134	0.175*
p	0.589	0.543	0.628	0.494	0.008	0.764	0.001	0.099	0.031
Monobenzyl (ng/ml)									
r	0.616**	0.172	-0.068	0.040	0.069	-0.114	0.144	0.082	0.101
p	0.002	0.432	0.758	0.857	0.400	0.163	0.077	0.317	0.215
Monobutyl (ng/ml)									
r	0.180	0.014	0.104	-0.100	0.101	-0.051	0.133	0.097	0.129
p	0.411	0.951	0.638	0.651	0.219	0.532	0.103	0.234	0.115

*Correlation is significant at the 0.05 level (2-tailed). **Correlation is significant at the 0.01 level (2-tailed).

were detected in urine samples of all subjects. This reflects increased exposure to the parent materials of these metabolites. The median concentration of MBzP, MEHP, MBP, and MEP was recorded as 1.4, 54.5, 29.9, and 490 ng/ml, respectively. The lowest urinary concentrations were observed with MBzP. These findings are consistent with that of Jeddi *et al.* [29] who found that MBzP had the lowest levels in both unadjusted and creatinine-adjusted models with a geometric mean value equal to 2.0 $\mu\text{g/L}$. On the other hand, Bamai *et al.* [30] found that the median concentrations of MBzP and MEHP in children aged 3–12 years were 17.05 and 16.9 $\mu\text{g/L}$, respectively, while Amin *et al.* [31] reported that the median concentrations of MBzP and MEHP were 240.77 and 101.97 $\mu\text{g/L}$, respectively. The European Food Safety Authority attributed in 2011 the differences in the reported urinary metabolite patterns to differences of the used patterns of phthalates in related products, variances in food consumption habits, and/or variances in socio-economic levels in different countries [32].

Table 5: Comparison of phthalate metabolites according to the BMI z-score category

BMI z-score category	Mean	Std. Deviation	t-test	p
Mono-(2ethylhexyl) phthalate (ng/ml)				
Obese	65.9241	23.28435	3.979	0.000*
Non-obese	53.6750	14.44615		
Monoethyl (ng/ml)				
Obese	606.2759	272.06351	3.823	0.000*
Non-obese	478.0870	136.72614		
Monobenzyl (ng/ml)				
Obese	1.8642	0.87290	1.735	0.085
Non-obese	1.6358	0.71872		
Monobutyl (ng/ml)				
Obese	31.7084	6.27011	2.062	0.041*
Non-obese	29.5349	6.24461		

*p < 0.05 is significant.

Metabolic processes showed greater vulnerability to environmental agitations throughout periods of development, characterized by hormonal variations and quick maturation of organs [33]. It has been assumed that phthalate disrupts lipid metabolism by affecting various human receptors: Peroxisome proliferator-activated receptors (PPAR α and PPAR γ), human estrogen receptors, androgen receptors, thyroid hormone receptors, and pregnane X receptor [34].

The present study showed a positive significant correlation between urinary monobenzyl and serum triglycerides, and insignificant negative associations between MEHP, MEP, and total cholesterol and triglycerides. Triglyceridemia correlated with phthalates exposure was evidenced in the previous studies [31], [34], [35]. In contrast, Jia *et al.* [36] reported

that high serum MEHP was associated with lower triglycerides, as well as lower levels of several fatty acid components in 318 pregnant Japanese women. In addition, reduction of TG concentrations following MEP exposure was documented in non-obese [37]. Reduction in cholesterol levels was also recently reported among Mexican youth at 8–14 years of age [38]. Nevertheless, the effect of phthalate exposure on lipid profile is evidently more complicated. It was postulated that phthalate activation of PPAR α may lead to hypolipidemic and antiadipogenic effects, while their activation of PPAR γ may lead to pro-adipogenic effects [39].

It was hypothesized that phthalates exposures prenatally, in childhood, and through adolescence were related to increased weight status and adiposity [40], [41], [42], [43], [44]. In the present study, it was found that the mean urinary levels of MBP, MEP, and MEHP in children with obesity were higher with a highly significant difference if compared with that of non-obese children. In addition, positive significant correlations between both MEHP and MEP on one side and body weight, BMI-Z score, WC, and HC on the other side were noted. This agrees with other studies, where MBzP, MEHP, and MEP were correlated with WC and BMI [26], [40]. Khalil *et al.* [45] showed a positive association between urinary phthalates and BMI in overweight children. In Iranian children, the high levels of phthalates were detected in children and adolescents with obesity in associations with increased BMI and WC [31]. WC as a tool to evaluate visceral obesity was more informative than BMI. These indicators have an association with increased cardiometabolic risks [46]. Moreover, a positive linear correlation between WC and urinary MEHP concentrations was also found in healthy normal-weight individuals [37]. Furthermore, recent epidemiological research has supported phthalates' role in triggering adverse cardiometabolic states in children and adolescents [31], [47].

The potential mechanisms assumed in the association between phthalates and obesity may include endocrine-disrupting activities and activation of PPARs. Activation of these receptors can increase lipid accumulation [48] and release adiposity hormones, causing an increased risk for developing obesity [49].

On the contrary, increased levels of MEP and DEHP were reported in the serum and urine of people undergoing weight loss [50]. Another study had reported a

negative correlation between levels of urinary phthalates and weight gain in children [51]. Other studies have not found associations between phthalate exposure and adiposity [52], [53]. Inconsistencies among reports can be explained by differences in methodological approaches, as variances in periods of exposure and outcome assessments (e.g., pre-puberty vs. post-puberty), data analysis methods (for instance, sex-stratified vs. all subjects), and populations risk factors such as genetics, epigenetics, and other confounders.

The principal limitation of this study is that it is cross-sectional, and thus a causal relationship cannot be proven.

Conclusion and Recommendation

The present study supports that phthalates exposure can lead to an increase in children's and adolescents' body weight, adiposity, and triglycerides. Community efforts to restrain phthalates use should be encouraged. Upcoming studies have to shed light on the risk of phthalates exposure in children and its association with later obesity and cardiovascular wellbeing, besides exploring the probable underlying mechanisms and causation.

References

- Holtrup B, Church CD, Berry R, Colman L, Jeffery E, Bober J, *et al.* Puberty is an important developmental period for the establishment of adipose tissue mass and metabolic homeostasis. *Adipocytes*. 2017;6:224-33. <https://doi.org/10.1080/21623945.2017.1349042>
PMid:28792785
- NCD Risk Factor Collaboration (NCD-RisC). Worldwide trends in body mass index, underweight, overweight, and obesity from 1975 to 2016: A pooled analysis of 2416 population-based measurement studies in 128.9million children, adolescents, and adults. *Lancet*. 2017;390:2627-42. [https://doi.org/10.1016/S0140-6736\(17\)32129-3](https://doi.org/10.1016/S0140-6736(17)32129-3)
PMid:29029897
- Commission on Ending Childhood Obesity. Report of the Commission on Ending Childhood Obesity; 2016. Available from: <http://www.who.int/end-childhood-obesity/en> [Last accessed on 2017 Feb 21].
- Reinehr T. Long-term effects of adolescent obesity: Time to act. *Nat Rev Endocrinol*. 2018;14(3):183-8. <https://doi.org/10.1038/nrendo.2017.147>
PMid:29170543
- Williams EP, Mesidor M, Winters K, Dubbert PM, Wyatt SB. Overweight and obesity: Prevalence, consequences, and causes of a growing public health problem. *Curr Obesity Rep*. 2015;4(3):363-70. <https://doi.org/10.1007/s13679-015-0169-4>
PMid:26627494
- Heindel JJ, Blumberg B, Cave M, Machtinger R, Mantovani A, Mendez MA, *et al.* Metabolism disrupting chemicals and metabolic disorders. *Reprod Toxicol*. 2017;68:3-33. <https://doi.org/10.1016/j.reprotox.2016.10.001>
- Apau J, Sefah W, Adua E. Human contact with phthalates during early life stages leads to weight gain and obesity. *Cogent Chem*. 2020;6:1815273. <https://doi.org/10.1080/23312009.2020.1815273>
- Choi J, Eom J, Kim J, Lee S, Kim Y. Association between some endocrine disrupting chemicals and childhood obesity in biological samples of young girls: Across-sectional study. *Environ Toxicol Pharmacol*. 2014;38:51-7. <https://doi.org/10.1016/j.etap.2014.04.004>
PMid:24908636
- Romano ME, Savitz DA, Braun JM. Challenges and future directions to evaluating the association between prenatal exposure to endocrine disrupting chemicals and childhood obesity. *Curr Epidemiol Rep*. 2014;1:57-66. <https://doi.org/10.1007/s40471-014-0007-3>
PMid:25328860
- Guo Y, Alomirah H, Cho HS, Minh TB, Mohd MA, Nakata H, *et al.* Occurrence of phthalate metabolites in human urine from several Asian countries. *Environ Sci Technol*. 2011;45(7):3138-44. <https://doi.org/10.1021/es103879m>
PMid:21395215
- Meeker JD, Sathyanarayana S, Swan SH. Phthalates and other additives in plastics: Human exposure and associated health outcomes. *Philos Trans R Soc Lond B Biol Sci*. 2009;364:2097-113. <https://doi.org/10.1098/rstb.2008.0268>
PMid:19528058
- Kim SH, Park MJ. Phthalate exposure and childhood obesity. *Ann Pediatr Endocrinol Metab*. 2014;19(2):69-75. <https://doi.org/10.6065/apem.2014.19.2.69>
PMid:25077088
- Wang Y, Zhu H, Kannan K. A review of biomonitoring of phthalate exposures. *Toxics*. 2019;7(2):21. <https://doi.org/10.3390/toxics7020021>
PMid:30959800
- Giuliani A, Zuccarini M, Cichelli A, Khan H, Reale M. Review critical review on the presence of phthalates in food and evidence of their biological impact. *Int J Environ Res Public Health*. 2020;17(16):5655. <https://doi.org/10.3390/ijerph17165655>
PMid:32764471
- Grün F, Blumberg B. Perturbed nuclear receptor signaling by environmental obesogens as emerging factors in the obesity crisis. *Rev Endocr Metab Disord*. 2007;8:161-71. <https://doi.org/10.1007/s11154-007-9049-x>
PMid:17657605
- Shehata MA, Salah EM, Youssef MM, Abushady MM, El-Alameey IR, Ashaat EA, *et al.* Comparing levels of urinary phthalate metabolites in Egyptian children with autism spectrum disorders and healthy control children: Referring to Sources of phthalate exposure. *Open Access Maced J Med Sci*. 2021;9:1640-6.
- WHO Multicentre Growth Reference Study Group. WHO child growth standards based on length/height, weight and age. *Acta Paediatr Suppl*. 2006;450:76-85. <https://doi.org/10.1111/j.1651-2227.2006.tb02378.x>
PMid:16817681
- Koch HM, Drexler H, Angerer J. Internal exposure of nursery-school children and their parents and teachers to di (2-ethylhexyl) phthalate (DEHP). *Int J Hyg Environ Health*. 2004;207(1):15-22. <https://doi.org/10.1078/1438-4639-00270>
PMid:14762970
- Hauser R, Calafat AM. Phthalates and human health. *J Occup*

- Environ Med. 2005;62(11):806-18.
20. Teitelbaum SL, Mervish N, Moshier EL, Vangeepuram N, Galvez MP, Calafat AM, et al. Associations between phthalate metabolite urinary concentrations and body size measures in New York City children. *Environ Res.* 2012;112:186-93. <https://doi.org/10.1016/j.envres.2011.12.006>
PMid:22222007
 21. Kim Y, Ha EH, Kim EJ, Park H, Ha M, Kim JH, et al. Prenatal exposure to phthalates and infant development at 6 months: Prospective mothers and children's environmental health (moceh) study. *Environ Health Perspect.* 2011;119(10):1495-500. <https://doi.org/10.1289/ehp.1003178>
PMid:21737372
 22. Feng YL, Liao X, Grenier G, Nguyen N, Chan P. Determination of 18 phthalate metabolites in human urine using a liquid chromatography-tandem mass spectrometer equipped with a core-shell column for rapid separation. *Anal Methods.* 2015;7:8048-59.
 23. Tietz NW. *Clinical Guide to Laboratory Tests.* 3rd ed. Philadelphia, PA: WB Saunders Company; 1995. p. 130-1.
 24. Lopez-Virella MF, Stone P, Ellis S, Colwell JA. Cholesterol determination by three different methods. *Clin Chem.* 1977;23:882-4.
PMid:192488
 25. Hata Y, Nakajima K. Application of Friedewald's LDL-cholesterol estimation formula to serum lipids in the Japanese population. *Jpn Circ J.* 1986;50(12):1191-200. <https://doi.org/10.1253/cj.50.1191>
PMid:3469420
 26. Stahlhut RW, van Wijngaarden E, Dye TD, Cook S, Swan SH. Concentrations of urinary phthalate metabolites are associated with increased waist circumference and insulin resistance in adult U.S. Males. *Environ Health Perspect.* 2007;115:876-82. <https://doi.org/10.1289/ehp.9882>
PMid:17589594
 27. Trasande L, Attina TM, Sathyanarayana S, Spanier AJ, Blustein J. Race/ethnicity-specific associations of urinary phthalates with childhood body mass in a nationally representative sample. *Environ Health Perspect.* 2013;121:501-6. <https://doi.org/10.1289/ehp.1205526>
PMid:23428635
 28. Frederiksen H, Skakkebaek NE, Andersson AM. Metabolism of phthalates in humans. *Mol Nutr Food Res.* 2007;51(7):899-911. <https://doi.org/10.1002/mnfr.200600243>
PMid:17604388
 29. Jeddi MZ, Gorji ME, Rietjens IM, Louise J, de Bruin YB, Liska R. Biomonitoring and subsequent risk assessment of combined exposure to phthalates in Iranian children and adolescents. *Int J Environ Res Public Health.* 2018;15(11):2336-58. <https://doi.org/10.3390/ijerph15112336>
PMid:30360526
 30. Bamai YA, Araki A, Kawai T, Tsuboi T, Yoshioka E, Kanazawa A, et al. Comparisons of urinary phthalate metabolites and daily phthalate intakes among Japanese families. *Int J Hyg Environ Health.* 2015;218(5):461-470. <https://doi.org/10.1016/j.ijheh.2015.03.013>
PMid:25888094
 31. Amin MM, Ebrahimpour K, Parastar S, Shoshtari-Yeganeh B, Hashemi M, Mansourian M, et al. Association of urinary concentrations of phthalate metabolites with cardiometabolic risk factors and obesity in children and adolescents. *Chemosphere.* 2018;211:547-56. <https://doi.org/10.1016/j.chemosphere.2018.07.172>
PMid:30092535
 32. European Food Safety Authority. Scientific report of EFSA. Overview of the procedures currently used at EFSA for the assessment of dietary exposure to different chemical substances. *EFSA J.* 2011;9:2490.
 33. Haney EM, Huffman LH, Cea B. *Screening for Lipid Disorders in Children and Adolescents.* Rockville, MD: Agency for Healthcare Research and Quality (US); 2007.
 34. James-Todd TM, Huang T, Seely EW, Saxena AR. The association between phthalates and metabolic syndrome: The National Health and Nutrition Examination Survey 2001-2010. *Environ Health.* 2016;15:52. <https://doi.org/10.1186/s12940-016-0136-x>
PMid:27079661
 35. Kim JH, Park H, Lee J, Cho G, Choi S, Choi G, et al. Association of diethylhexyl phthalate with obesity related markers and body mass change from birth to 3 months of age. *J Epidemiol Community Health.* 2016;70:466-72. <https://doi.org/10.1136/jech-2015-206315>
PMid:26834143
 36. Jia X, Harada Y, Tagawa M, Naito H, Hayashi Y, Yetti H, et al. Prenatal maternal blood triglyceride and fatty acid levels in relation to exposure to di(2-ethylhexyl) phthalate: A cross-sectional study. *Environ Health Prev Med.* 2014;20(3):168-78.
 37. Milošević N, Milanović M, Sudji J, Živanović DB, Stojanoski S, Vuković B, et al. Could phthalates exposure contribute to the development of metabolic syndrome and liver disease in humans? *Environ Sci Pollut Res.* 2020;27(1):772-84. <https://doi.org/10.1007/s11356-019-06831-2>
PMid:31808097
 38. Perng W, Watkins DJ, Cantoral A, Mercado-García A, Meeker JD, Téllez-Rojo MM, et al. Exposure to phthalates is associated with lipid profile in peripubertal Mexican youth. *Environ Res.* 2017;154:311-7. <https://doi.org/10.1016/j.envres.2017.01.033>
PMid:28152472
 39. Hurst CH, Waxman DJ. Activation of PPAR α and PPAR γ by environmental phthalate monoesters. *Toxicol Sci.* 2003;74:297-308. <https://doi.org/10.1093/toxsci/kfg145>
PMid:12805656
 40. Wang H, Zhou Y, Tang C, He Y, Wu J, Chen Y, et al. Urinary phthalate metabolites are associated with body mass index and waist circumference in Chinese school children. *PLoS One.* 2013;8(2):56800. <https://doi.org/10.1371/journal.pone.0056800>
PMid:23437242
 41. Harley KG, Berger K, Rauch S, Kogut K, Henn BC, Calafat AM, et al. Association of prenatal urinary phthalate metabolite concentrations and childhood BMI and obesity. *Pediatr Res.* 2017;82(3):405-15. <https://doi.org/10.1038/pr.2017.112>
PMid:28426647
 42. Shoaff J, Papandonatos GD, Calafat AM, Ye X, Chen A, Lanphear BP, et al. Early-life phthalate exposure and adiposity at 8 years of age. *Environ Health Perspect.* 2017;125(9):097008. <https://doi.org/10.1289/EHP1022>
PMid:28935615
 43. Vafeiadi M, Myridakis A, Roumeliotaki T, Margetaki K, Chalkiadaki G, Dermizaki E, et al. Association of early life exposure to phthalates with obesity and cardiometabolic traits in childhood: Sex specific associations. *Front Public Health.* 2018;6:327. <https://doi.org/10.3389/fpubh.2018.00327>
PMid:30538977
 44. Yang TC, Peterson KE, Meeker JD, Sánchez BN, Zhang Z, Cantoral A, et al. Exposure to Bisphenol A and phthalates metabolites in the third trimester of pregnancy and BMI trajectories. *Pediatr Obes.* 2018;13:550-7. <https://doi.org/10.1111/ijpo.12279>
PMid:29700996
 45. Khalil N, Chen A, Lee M. Endocrine disruptive compounds and

- cardio-metabolic risk factors in children. *Curr Opin Pharmacol*. 2014;19:120-4. <https://doi.org/10.1016/j.coph.2014.09.015>
PMid:25306432
46. Aguilar-Morales I, Colin-Ramirez E, Rivera-Mancía S, Vallejo M, Vázquez-Antona C. Performance of waist-to-height ratio, waist circumference, and body mass index in discriminating cardio-metabolic risk factors in a sample of school-aged Mexican Children. *Nutrients*. 2018;10(12):1850. <https://doi.org/10.3390/nu10121850>
PMid:30513720
47. Gaston SA, Tulve NS. Urinary phthalate metabolites and metabolic syndrome in U.S. adolescents: Cross-sectional results from the national health and nutrition examination survey (2003-2014) data. *Int J Hyg Environ Health*. 2019;222(2):195-204. <https://doi.org/10.1016/j.ijheh.2018.09.005>
PMid:30297147
48. Stojanoska MM, Milankov A, Vukovic B, Vukcevic D, Sudji J, Bajkin I, *et al*. Do diethyl phthalate (DEP) and di-2-ethylhexyl phthalate (DEHP) influence the metabolic syndrome parameters? Pilot study. *Environ Monit Assess*. 2015;187(8):526. <https://doi.org/10.1007/s10661-015-4754-5>
PMid:26210228
49. Taxvig C, Dreisig K, Boberg J, Nellemann C, Schelde AB, Pedersen D, *et al*. Differential effects of environmental chemicals and food contaminants on adipogenesis, biomarker release and PPAR gamma activation. *Mol Cell Endocrinol*. 2012;361(1-2):106-15. <https://doi.org/10.1016/j.mce.2012.03.021>
PMid:22526026
50. Dirtu AC, Geens T, Dirinck E, Malarvannan G, Neels H, Van Gaal L, *et al*. Phthalate metabolites in obese individuals undergoing weight loss: Urinary levels and estimation of the phthalates daily intake. *Environ Int*. 2013;59:344-53. <https://doi.org/10.1016/j.envint.2013.06.023>
51. Boas M, Frederiksen H, Feldt-Rasmussen U, Skakkebaek NE, Hegedus L, Hilsted L, *et al*. Childhood exposure to phthalates: Associations with thyroid function, insulin-like growth factor I, and growth. *Environ Health Perspect*. 2010;118:1458-64. <https://doi.org/10.1289/ehp.0901331>
PMid:20621847
52. Buckley JP, Engel SM, Mendez MA., Richardson DB, Daniels JL, Calafat AM, *et al*. Prenatal phthalate exposures and childhood fat mass in a New York City cohort. *Environ Health Perspect*. 2016;124:507-13. <https://doi.org/10.1289/ehp.1509788>
PMid:26308089
53. Carlsson A, Sørensen K, Andersson AM, Frederiksen H, Juul A. Bisphenol A. Phthalate metabolites and glucose homeostasis in healthy normal-weight children. *Endocr Connect*. 2018;7(1):232-8. <https://doi.org/10.1530/EC-17-0344>
PMid:29237763