




Medicinal Properties of Ant Nest Plant (*Myrmecodia* Genus): A Comprehensive Review

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Abstract

BACKGROUND: The *Myrmecodia* genus belongs to the *Rubiaceae* family, which has the characteristic of an epiphytic plant. This genus spread from Malaysia, through the Philippines, Sumatra, Borneo, Java, New Guinea (Papua), Cape York to the Solomon Islands and Pacific region. It has been empirically proven to treat gout, inflammation, muscle pain relief, strengthen the body's immunity, and cancer treatment.

AIM: This article reviewed the scientific work about the *Myrmecodia* genus concerning their traditional uses, phytochemical compounds, and pharmacological activities of *Myrmecodia*.

METHODS: The sources were from the scientific literature online database, including Science Direct, PubMed, and Google Scholar between 1978 and 2021. Furthermore, 112 articles were collected, while 61 full-text were assessed for eligibility.

RESULTS: The *Myrmecodia* extract has many pharmacological activities, namely antioxidant, antibacterial, cytotoxic, and anticancer.

CONCLUSION: Moreover, this review may be used as scientific literature for the next investigations on the genus *Myrmecodia*, especially regarding pharmacological properties such as *in vitro* and *in vivo* studies and isolation of the active compounds.

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Introduction

Natural therapy has been utilized since long ago, where the knowledge is obtained through generations so that it is known as traditional medicine in the community for local wisdom [1]. Nowadays, most of the world's population still relies on traditional medicines for healthcare purposes. Plants, in particular, are always used as herbal medicine, as they contain have a wide range of therapeutics for the treatment of diseases. Indonesia, a country in Southeast Asia, has a diverse flora that potential as medicinal plants from nature. One of the selected plants is the ant nest plant (*sarang semut* locally name). The ant nest plant *Myrmecodia* is an epiphytic plant of the *Rubiaceae* family [2]. Five known genera are belonging this family with hypocotyl and epiphytic plants such as *Myrmecodia*, *Hydrophytum*, *Anthorrhiza*, *Myrmephytum*, and *Squamellaria*. Only two of them are associated with ants, *Myrmecodia* and *Hydrophytum*. The *Myrmecodia* genus is the second most diverse species in the world, after the genera *Hydrophytum*. The number of species of *Myrmecodia*

genera is known 26 species. *Myrmecodia* is distributed from Malaysia through the Philippines, Sumatera, Borneo, Java, New Guinea (Papua), Cape York to the Solomon Islands, the Pacific region. The greatest diversity of *Myrmecodia* species is found on the island of Papua-Indonesia and Papua New Guinea [3]. *Sarang semut* (*Ant nest*) were used as folk medicine by local people in Indonesia, especially in the Papua region, to treat various diseases empirically [4]. This study is a comprehensive review, using and collecting the scientific literature online database on Science Direct, PubMed, and Google Scholar of *Myrmecodia* genus which is basically based on some criteria. The search keywords included "Myrmecodia," "Myrmecodia for anticancer," "Myrmecodia and phytochemical," and "Myrmecodia and pharmacological activity." Based on ethnomedicinal properties, only several species of the *Myrmecodia* genus are evaluated from scientific evidence-based medicine. In this articles, 112 articles were collected and identified from the scientific literature online database, between 1978 and 2021, while 61 full-text were included. This comprehensive review aims to determine the scientific evidenced base medicine of *Myrmecodia*

species in ethnopharmacological use and analyze phytochemical constituents and biological activities for the drug discovery from Indonesian medicinal plants.

Traditional Uses

The *Myrmecodia* genus has been traditionally used as folk medicine throughout Southeast Asia to treat ulcer, swelling, headache, hemorrhoid, nosebleed, backache, skin rashes, allergy, uric acid disorder, renal problems, tuberculosis, tumor, coronary artery disease, hepatitis, rheumatism, and diarrhea [5], [6], [7], [8]. Based on an ethnopharmacology study from some selected regions in Indonesia, the genus of *Myrmecodia* has been used as traditional medicine by the community in East Indonesia, especially in Papua, to treat inflammation, muscle pain relief, and strengthen the body's immunity [9]. Rural people in the Papua community use the tuber (hypocotyl) powder of the *Myrmecodia* plant as a brewed drink like tea with decoction method [10], [11]. In the Papua region, *Myrmecodia pendens* Merr. and Perry can treat severe diseases, such as nausea, breast cancer, and immunomodulator [12], [13]. Meanwhile, in Belu Regency, East Nusa Tenggara, people used this plant as a traditional medicine to treat blood circulation and gout [14]. People in Borneo Island used *M. pendens* as an antibacterial [15]. *Myrmecodia tuberosa* Jack. is generally used in the West Papua region-Indonesia to natural healing remedies and treat cancer [16]. In Wamena-Papua, this plant was applied to enhance body immunity for a long time [17]. Meanwhile, people in Mindanao-Philippines used fresh tubers of this species to treat several diseases such as goiter, stomachache, and fever [18]. In Vietnam, *M. tuberosa* was used as a folk medicine to treat hepatitis [19]. Another species of *Myrmecodia* genus *Myrmecodia platytyrea* Becc. is believed to have medicinal value, commonly used for cancer

Table 1: Traditional uses of the tuber of *Myrmecodia* species

Species	Region	Traditional uses
<i>M. pendens</i> Merr. and Perry	Wamena-Papua	to treat ulcer, hemorrhoid, nosebleed, backache, allergy, uric acid disorder, stroke, rheumatism and diarrhea [4]
	Bintuni-West Papua	to treat nausea and complementary medicine for breast cancer [5]
	Belu-NTT	for treatment of blood circulation and gout [14]
<i>M. tuberosa</i> Jack.	West Papua	natural healing remedies for tumor [16]
	Lamdong-Vietnam	to treat hepatitis and stomachache [19]
<i>M. platytyrea</i> Becc.	Indonesia and Malaysia	treatment for cancer and tumor and to reduce the glucose level in the blood [25]
	Throughout Southeast Asia and Papua	complementary medicine for the treatment of cancer [20] for cancer treatments, and prevent and cure diarrhea [21]
<i>M. beccarii</i> Hook.f.	Merauke-Papua	strengthening immunity and treatment for gout [23]
<i>M. rumphii</i> Becc.		treatment for rheumatic and tumor [24]

treatment, keeping blood glucose and cholesterol at normal levels [20], [21], [22]. *Myrmecodia beccarii* Hook.f. and *Myrmecodia rumphii* Becc. A resident in Merauke Regency, Papua, used as herbal medicine for gout and immunostimulant [23], [24]. The usages of *Myrmecodia* species as traditional medicine can be found in Table 1.

Phytochemistry

The widespread use of the *Myrmecodia* genus has resulted in the phytochemical constituent analysis of different species. Based on a phytochemical screening test, generally, the hypocotyl of the *Myrmecodia* genus contained flavonoid, tannin, saponin, glycoside, carbohydrates, and quinone [4], alkaloids [18], iridoid [19], [25], steroid/triterpenoid [26], diterpenoid [27], phenolics [28], and polyphenolics [29], [30]. The phytochemical screening showed the major compounds of *Myrmecodia* species are phenolic and flavonoid (Figure 1). Several species of them have been successfully identified, *M. pendens* contained kaempferol, luteolin, rutin, quercetin and apigenin, rosmarinic acid, procyanidin B1, the polymer of procyanidin B1, gallic acid, (+)-catechin, caffeic acid, p-coumaric acid, and ferulic acid [7], [31], [32]. Some researchers reported that phenolic glycoside with aliphatic moiety [13], dibenzo-p-dioxin-2,8-dicarboxylic acid [33], guaicol, 1,4-di-tert-butoxybenzene, 4-methylcatechol and 2,6-dimethoxy-phenol [34], anthocyanin [35] and biflavonoid [36] have been isolated from *M. pendens*. Other isolates of flavonoid and phenolic compounds were found in *M. platytyrea* such as, acylated flavanone, flavanone (liquiritigenin or isoliquiritigenin), O-methylated isoflavone (calycosin) and 2-(2-methylbutyryl) phloroglucinol glucoside [37]. Some iridoid compounds were found in *M. tuberosa*, namely, myrmecodoide A, myrmecodoide B, asperulosidic acid, deacetylasperulosidic acid, premnosidic acid, asperuloside [19], and morindolide in *M. platytyrea* [37]. Stigmasterol was identified from *M. platytyrea* [21], [29], [30], [37] and the other terpenoid compounds have been isolated from *M. pendens* such as, diterpenoid type labdane [27], stigmast-4-ene-3-one, pomolic acid, 6'-O-tridecanoyl-3-O- β -D glucosyl-sitosterol, phloroglucinol sesquiterpene, and betulin [33].

Pharmacological Activities

Myrmecodia species have been screened for various pharmacological activities *in vitro* and *in vivo* using animal models.

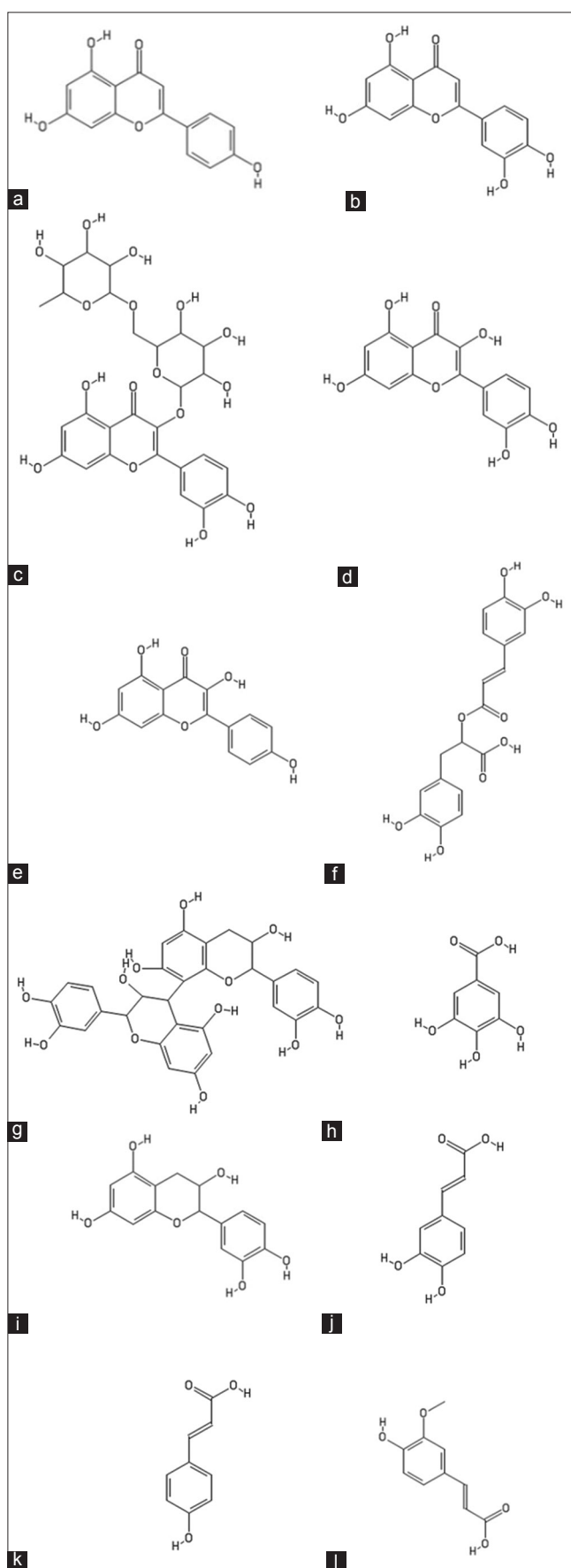


Figure 1: Some structures of phenolic compounds that have been identified from *Myrmecodia* Species. (a) kaempferol; (b) luteolin; (c) rutin; (d) quercetin; (e) apigenin; (f) rosmarinic acid; (g) procyanidin B1; (h) gallic acid; (i) (+)-catechin; (j) caffeic acid; (k) p-coumaric acid; (l) ferulic acid.

Antioxidant activity

Many *Myrmecodia* species showed potential antioxidant activity with 2,2-diphenyl-1-picrylhydrazyl (DPPH) method. The free radical scavenging activity (antioxidant activity) of the ethanol extract of hypocotyl *M. pendens* was evaluated using DPPH radical and the IC_{50} value occurred at 96.21 $\mu\text{g/mL}$ of extract and contained total phenol and flavonoid contents 330.61 mg GAE/g and 63.28 mg QE/g of dry extract, respectively. The extract was analyzed by High Performance Liquid Chromatography and five flavonoid compounds were identified and quantified, kaempferol (13.767 mg/g), luteolin (0.005 mg/g), rutin (0.003 mg/g), quercetin (0.030 mg/g), and apigenin (4.700 mg/g) of dry extract [31]. The ethyl acetate fraction of this extract contained procyanidin B1 dimer (3.236 mg/g dry sample) and rosmarinic acid (20.688 mg/g dry sample) showed significant free radical scavenging capacities with IC_{50} values of 27.59 $\mu\text{g/mL}$ and 35.80 $\mu\text{g/mL}$, respectively [32]. Meanwhile, the antioxidant activities of the aqueous fractions of *M. pendens* were higher than chloroform fractions [15]. The new extraction method was successfully investigated, Supercritical Carbon dioxide ($SC\text{-CO}_2$) with DPPH-scavenging test showed antioxidant capacity of *M. pendens* with an IC_{50} level of 3.62 mg/mL [7] and The Microwave-Assisted Extraction (MAE) showed scavenging radical activity with IC_{50} 0.98 $\mu\text{g/mL}$ [35]. Ethanol extract of stem bark, leaves and tuber of *M. tuberosa* showed antioxidant activity with 95.17%, 94.55%, and 93.42% DPPH scavenging activities, respectively [25]. The previous study investigated ethyl acetate fractions of the hypocotyl of *M. platytyrea* showed antioxidant activity with IC_{50} 21.57 $\mu\text{g/mL}$ [37]. Polyphenols and flavonoids of ethanol and ethyl acetate extracts expressed the highest antioxidant activity compared to dichloromethane and methanol extracts of *M. platytyrea* with High Performance Thin Layer Chromatography (HPTLC)-DPPH bioautographic methods [21], [29], [30]. The methanol extract of *M. beccarii* and *M. rumphii* showed antioxidant activity with IC_{50} 8.18 $\mu\text{g/mL}$ [23] and 90.98% DPPH radical scavenging activity, respectively [24].

Antibacterial activity

Some of *Myrmecodia* species demonstrated significant antibacterial agents against various types of bacterial spectrum. *M. pendens* gave had antibacterial activity against *Escherichia coli*, *Salmonella* sp. and *Bacillus* sp. [15], *Candida albicans*, *Staphylococcus aureus* [38], *Streptococcus mutans* [39], *Shigella dysenteriae* [40], *Streptococcus sanguinis* [41], [42], [43], *Enterococcus faecalis* [34], [44], *Porphyromonas gingivalis* [45] and *Treponema denticola* [42].

Alibasyah et al. reported that biflavonoid and diterpenoid compounds were isolated from ethyl acetate extract of *M. pendens* exposed antibacterial

activity against *P. gingivalis* pathogen using Kirby-Bauer method with minimum inhibitory concentration (MIC) of 19.57 µg/mL and 39.06 µg/mL, respectively [36]. Six compounds phenolic and terpenoid group have been found in *M. pendens* and some of them gave antibacterial activity by Kirby-Bauer method with the inhibition zone of isolates at concentration of 5000 µg/mL, such as dibenzo-p-dioxin-2,8-dicarboxylic acid against *E. faecalis* (8.55 mm), stigmast-4-ene-3-one and pomolic acid against *S. mutans* were 9.00 mm and 10.24 mm respectively and phloroglucinol sesquiterpene against *P. gingivalis* (12.31 mm). Gartika *et al.* evaluated labdane diterpene isolate from ethyl acetate extract of *M. pendens* had antibacterial activity against *S. mutans* biofilm as 50 µg/mL, and the minimum biofilm eradication concentration value for one min induction time was 40% [27].

The extract of *M. tuberosa* was evaluated for antimicrobial activity test against *C. albicans*, *E. coli* and *S. aureus* by disc diffusion method with MIC value 0.8%, 0.8%, and 1.6% w/v, respectively, and minimum bactericidal concentration (MBC) value by microdilution assay were >6.4% w/v against *C. albicans*, 6.4% w/v against *E. coli* and 1.6% w/v against *S. aureus* [38]. Six iridoids have been isolated from methanol extract *M. tuberosa* had antibacterial activity against *S. aureus* with MIC value of 100.0 µg/mL [19], and phenolic compound of ethyl acetate fraction showed the potential antibacterial activity [28].

Kuswandani *et al.* figured that the combination of n-hexane and ethyl acetate fractions of *M. pendens* gave the best effect against *E. faecalis* using serial microdilution method with MIC 0.049 mg/mL and Mueller-Hinton method with MBC 12.50 mg/mL [46].

Cytotoxic activity

Many assays have been used to evaluate the toxicity activities of *Myrmecodia* species that associated with the potential of anticancer activity. Some crude extracts and compounds showed the inhibitory activity against various *in vitro* cancer cell lines (Table 2) [48], [49], [50], [51], [52], [53], [54], [55], [56], [57], [58]. Besides, Suharyanto and Purwono reported that *M. pendens* had effective anticancer activity *in vivo* model series. The pre-clinical trials revealed the optimum dose 750 mg/kg body weight of *M. pendens* extract could use to treat lung cancer cell from necrosis [59].

Immunomodulatory activity

The hypocotyl of *M. tuberosa* ethanol extract (50 µg/mL) showed the highest macrophage phagocytic index and for the species *M. pendens* ethyl acetate fraction (50 µg/mL) presented the highest activity in lymphocytes proliferation assay. Both were potential candidate as immunomodulatory agents from herbal medicine [5]. Sumardi *et al.* reported that the active

Table 2: Cytotoxic activity of crude extract, fraction and isolate of Myrmecodia species

Species	Extraction method	Extract/fraction	Extract/ fraction/ compound	Cancer cell line (IC ₅₀ µg/mL)
<i>M. pendens</i> Merr. and Perry	Reflux	Gradient: EtOAc, BuOH, water	Water extract	Hela (29.36), MCM-B2 (74.20) [4]
			EtOAc	Hela (48.13), MCM-B2 (111.06) [4]
			BuOH	Hela (42.33), MCM-B2 (87.13) [4]
			Water fraction	Hela (27.61), MCM-B2 (64.57) [4]
			Doxorubicin	Hela (5), MCM-B2 (5.0) [4]
	Soxhlet	EtOH	EtOH	Hela (33.28) [47]
			Doxorubicin	Hela (5.56) [47]
	Maceration	Gradient: EtOH, n-hexane, EtOAc, water	EtOH	SP-C1 (937.562) [48], [49], [50]
			n-Hexane	SP-C1 (2691.535) [48], [49], [50]
			EtOAc	SP-C1 (452.059) [48], [49], [50]
			Water	SP-C1 (12302.69) [48], [49], [50]
			EtOH	B88 (471.79) [51]
	Maceration Soxhlet	Gradient: EtOAc, Methanol	Terpenoid	SKOV-3 (481.0) 48 hrs [52]
			Isolate	SKOV-3 (463.0) 72 hrs [52]
	Reflux	Water	EtOH	Fibroblast (<1000) [12]
EtOH			Fibroblast (<1000) [12]	
Maceration	EtOH	Water	Fibroblast (<1000) [12]	
		EtOH	Fibroblast (<1000) [12]	
Maceration	MeOH: partition n-Hexane, EtOAc	MeOH	Caco-2 (<100) [53]	
		n-Hexane	Caco-2 (24), HCT-116 (33.0) [53]	
Maceration	EtOAc	EtOAc	Caco-2 (<100) [53]	
		EtOAc	Burkitt's Lymphoma (<500) [54]	
Maceration	EtOAc: partition n-Hexane, EtOH, Water	EtOAc	Burkitt's Lymphoma (<1000) [55]	
		n-Hexane	Burkitt's Lymphoma (<1000) [55]	
		EtOH	Burkitt's Lymphoma (<1000) [55]	
Decoction	Water	Water	Burkitt's Lymphoma (<1000) [55]	
		Water	HSC-3 (<5000) 24 h; (<3000) 48 h [56]	
		Doxorubicin	HSC-3 (0.005 mg/ml) [56]	
Decoction	Water	Water	HSC-3 (<5000) 24 h; (<2500) 48 h [57]	
		Doxorubicin	HSC-3 (5000) [57]	
<i>M. platytyrea</i> Becc.	Soxhlet	MeOH	MeOH ^b	HCC (<100) [20]
<i>M. tuberosa</i> Jack.	Maceration	EtOH	EtOH ^a	HT-29 (16.0) [25]
			EtOH ^c	Hela (14.0) [25]
	Maceration	EtOH	EtOH	MCF-7 (6.0) [25]
			Docetaxel	KB (215.0) [17]
	Decoction	Water	Water ^a	KB (12.5) [17]
			Water ^b	Brine Shrimp (132.6)* [18]
	Maceration	EtOH	EtOH	Brine Shrimp (441.6)* [18]
			EtOH	Brine Shrimp (38.68)* [18]
	Maceration	EtOH: partition n-Hexane	Non hexane fraction (NHF)	Brine Shrimp (126.62)* [18]
			EtOH	Hela (0.54) [58]
				MCF-7 (0.22) [58]
				T47D (0.23) [58]

^ahypocotyl (tuber), ^bleaves, ^cbark; ^dLC: Lethal Concentration.

n-hexane fraction of *M. tuberosa* was administrated orally on doxorubicin-induced rats for 28 days, could maintain the number of TCD4+ cells, with ED₅₀ value 24.24 mg/kg body weight [16].

Antidiabetic activity

The dichloromethane extract of *M. platytyrea* showed significant α-amylase inhibitory activity and correlated with stigmasterol compound present in this extract [29], [30]. The ethyl acetate extract of *M. tuberosa* tuber figured moderate level activity for inhibition of α-glucosidase enzyme [25].

Other pharmacological activities

Simanjuntak *et al.* expressed that a chemical compound was successfully isolated as xanthine oxidase inhibitor with IC₅₀ 79.77% from n-butanol fraction of *M. pendens* extract [60] and the ethanol extract affected treating inflammation of dental pulp [61]. *M. platytyrea* water extract was able to lower LDL cholesterol concentration in blood and potential to remedy hypercholesterolemia related diseases, especially atherosclerosis [22].

Conclusion

The present comprehensive review discussed the traditional uses, pharmacological activities, and phytochemical properties of the *Myrmecodia* genus. A literature survey had been conducted that most of the species are used as folk medicine in Indonesia especially in the Papua region. According to evidence-based pharmacological activities, most of the *Myrmecodia* species were limited to the *in vitro* screening of biological activities and still limited *in vivo* model. Indeed, phytochemical research with guidance bioactivities of *Myrmecodia* species may be useful for searching active compounds from natural medicine.

References

- Thalib A, Masadah R, Prihartono P, Hamid F, Hasan H, Keliwawa S, *et al.* *Laportea decumana* (Robx) Wedd. Herbal endemic potential from Indonesia: A literature review. Open Access Maced J Med Sci. 2021;9(F):639-43. <https://doi.org/10.3889/oamjms.2021.7759>
- Huxley CR. The ant-plants *Myrmecodia* and *Hydnophytum* (Rubiaceae) and the relationship between them, ant occupants, physiology and ecology. New Phytol. 1978;80(1):231-68. <https://doi.org/10.1111/j.14698137.1978.tb02285.x>
- Huxley CR, Jebb MH. The tuberous of epiphytes of the *Rubiaceae* 5: A revision of *Myrmecodia*. Blumea. 1993;37(2):271-34.
- Soeksmanto A, Subroto MA, Wijaya H, Simanjuntak P. Anticancer activity test of sarang semut plant (*Myrmecodia pendens*) to Hela and MCM-B2 cells. Pak J Biol Sci. 2010;13(3):148-51. <https://doi.org/10.3923/pjbs.2010.148.151> PMID:2043775
- Hertiani T, Sasmito E, Sumardi, Ulfah M. Preliminary study on immunomodulatory effect of sarang-semut tubers *Myrmecodia tuberosa* and *Myrmecodia pendens*. Online J Biol Sci. 2010;10(3):136-41. <https://doi.org/10.3844/ojbsci.2010.136.141>
- Hamsar MN, Mizaton HH. Potential of ant-nest plants as an alternative cancer treatment. J Pharm Res. 2012;5(6):3063-6.
- Sanjaya RE, Tedjo YY, Kurniawan A, Ju YH, Ayucitra A, Ismadji S. Investigation on supercritical CO₂ extraction of phenolic-phytochemicals from an epiphytic plant tuber (*Myrmecodia pendans*). J CO₂ Util. 2014;6:26-33. <https://doi.org/10.1016/j.jcou.2014.03.001>
- Nugraha AS, Triatmok B, Wangchuk P, Keller PA. Vascular epiphytic medicinal plants as sources of therapeutic agents: Their ethnopharmacological uses, chemical composition, and biological activities. Biomolecules. 2020;10(2):181-247. <https://doi.org/10.3390/biom10020181>
- Dirgantara S, Dewi K, Natalia J, Simanjuntak TL. Botanical and phytochemical study of three specieses sarang semut from sub-Province Merauk, Papua. J Farm Sai Ter. 2015;2(2):20-2. <https://doi.org/10.33508/jfst.v2i2.718>
- Mardany MP, Chrystomo LY, Karim AK. Phytochemical screening and cytotoxic activity test of ant nest plant *Myrmecodia beccarii* Hook.f. from Merauke region. J Biol Papua. 2016;8(1): 13-22. <https://doi.org/10.31957/jbp.41>
- Pasaribu YP, Buyang Y, Rettob AL, Latuheru R, Marlissa I. Phytochemical Screening of ant Plant *Myrmecodia rumphii* Becc. In: Proceedings of the International Conference on Science and Technology, Bali, Indonesia; 2018. <https://doi.org/10.2991/icst-18.2018.60>
- Sudiono J, Oka CT, Trisfilha P. The scientific base of *Myrmecodia pendans* as herbal remedies. Br J Med Med Res. 2015;8(3):230-7. <https://doi.org/10.9734/BJMMR/2015/17465>
- Ulfah M, Sasmito E, Hertiani T. Immunomodulatory compound from *Myrmecodia pendens* Merr. and Perry. Ind J Pharm Sci. 2013;11(2):167-74.
- Buang Y, Noya E, Ola PD, Da Cunha T. Antioxidant activities of chloroform and aqueous fractions of *Myrmecodia pendens* extract: A preliminary study. J Appl Chem Sci. 2013;2(1):187-95.
- Crisnaningtyas F, Rachmadi AT. The use of ant nest (*Myrmecodia pendens*) from South Kalimantan as antibacterial agent. J Riset Ind Hasil Hutan. 2010;2(2):31-5. <https://doi.org/10.24111/jrihh.v2i2.1144>
- Sumardi, Hertiani T, Sasmito E. Ant plant (*Myrmecodia tuberosa*) hypocotyl extract modulates TCD4+ and TCD8+ cell profile of doxorubicin-induced immune-suppressed Sprague Dawley rats *in vivo*. Sci Pharm. 2013;81(4):1057-69. <https://doi.org/10.3797/scipharm.1302-03> PMID:24482773
- Yuletnawati SE, Meiyanto E, Agustina D. High antitumor activity of ethanolic extracts of Papua's ant nest plant (*Myrmecodia tuberosa*) on an oral carcinoma (KB) cell line. Int J Sci. 2016;5(1):1619-23. <https://doi.org/10.21275/v5i1.nov1531391>
- Demetillo MT, Baguio ML, Uy MM, Nuñez OM. Phytochemical analysis and cytotoxic activity of ant plant (*Myrmecodia tuberosa* Jack.). J Bio Environ Sci. 2017;11(6):85-91.
- Hanh NP, Phan NH, Thuan NT, Han TT, Vien LT, Thao NP, *et al.* Two new simple iridoids from the ant-plant *Myrmecodia tuberosa* and their antimicrobial effects. Nat Prod Res. 2016;30(18):2071-6. <https://doi.org/10.1080/14786419.2015.1113412> PMID:26595668
- Ju A, Cho YC, Kim BR, Lee S, Le HT, Vuong HL, *et al.* Anticancer effects of methanol extract of *Myrmecodia platytyrea* Becc. Leaves against human hepatocellular carcinoma cells via inhibition of ERK and STAT3 signaling pathway. Int J Oncol. 2018;52(1):201-10. <https://doi.org/10.3892/ijo.2017.4178> PMID:29075791
- Agatonovic-Kustrin S, Morton DW, Adam A, Mizaton HH, Zakaria H. High-performance thin-layer chromatographic methods in the evaluation of the antioxidant and anti-hyperglycemic activity of *Myrmecodia platytyrea* as a promising opportunity in diabetes treatment. J Chromatogr A. 2017;1530:192-6. <https://doi.org/10.1016/j.chroma.2017.11.012> PMID:29132827
- Nik Hasan MK, Wahab IA, Mizaton HH, Rasadah MA. Efficacy of *Myrmecodia platytyrae* (MyP) water extract in reducing

- cholesterol level in hypercholesterolemia induced Sprague Dawley rat. *Malays J Med Biol Res.* 2020;7(1):27-38. <https://doi.org/10.18034/mjmb.v7i1.491>
23. Dirgantara S, Nawawi A, Insanu M. Antioxidant activity test of three species of sarang semut plant from Merauke region, Papua. *J Biol Papua.* 2013;5(1):10-4. <https://doi.org/10.31957/jbp.517>
 24. Pasaribu YP, Buyang Y, Pallitin ID, Ersam T, Nimah YL. Preparation and antioxidant activity of methanol extract of *Myrmecodia rumphii* Becc. *Indian J Public Health Res Dev.* 2018;9(1):391-6. <https://doi.org/10.5958/0976-5506.2018.00072.4>
 25. Rasemi S, Yen KH, Ahmad R, Hasan MH. Total phenolic contents, antioxidant, anticancer and antidiabetic properties of *Myrmecodia tuberosa* (Rubiaceae). *J Adv Chem.* 2014;9(3):2000-4. <https://doi.org/10.24297/jac.v9i3.1007>
 26. Sari YP, Kustiawan W, Sukartiningih, Ruchaemi A. The potential of secondary metabolites of *Myrmecodia tuberosa* from different host trees. *Nus Biosci.* 2017;9(2):170-4. <https://doi.org/10.13057/nusbiosci/n090211>
 27. Gartika M, Pramesti HT, Kurnia D, Satari MH. A terpenoid isolated from sarang semut (*Myrmecodia pendans*) bulb and its potential for the inhibition and eradication of *Streptococcus mutans* biofilm. *BMC Complement Altern Med.* 2018;18(1):151-8. <https://doi.org/10.1186/s12906-018-2213-x> PMID:29739390
 28. Setyani W, Hertiani T, Murti YB. Isolation and identification of antimicrobial compound from sarang-semut tubers (*Myrmecodia tuberosa* (non Jack.) Bl). *Med Farm Indones.* 2012;7(1):275-82.
 29. Agatonovic-Kustrin S, Morton DW, Mizaton HH, Zakaria H. The relationship between major polyphenolic acids and stigmasterol to antioxidant activity in different extracts of *Myrmecodia platytyrea*. *S Afr J Bot.* 2018;115:94-9. <https://doi.org/10.1016/j.sajb.2017.12.011>
 30. Agatonovic-Kustrin S, Morton DW. HPTLC-Bioautographic methods for selective detection of the antioxidant and α -amylase inhibitory activity in plant extracts. *MethodsX.* 2018;5:797-802. <https://doi.org/10.1016/j.mex.2018.07.013> PMID:30101083
 31. Engida AM, Kasim NS, Suryadi YA, Tsigie I, Huynh LH, Ju YH. Extraction, identification and quantitative HPLC analysis of flavonoids from sarang semut (*Myrmecodia pendans*). *Ind Crops Prod.* 2013;41:392-6. <https://doi.org/10.1016/j.indcrop.2012.04.043>
 32. Engida AM, Faika S, Nguyen-Thi BT, Ju YH. Analysis of major antioxidants from extracts of *Myrmecodia pendans* by UV/visible spectrophotometer, liquid chromatography/tandem mass spectrometry, and high-performance liquid chromatography/UV techniques. *J Food Drug Anal.* 2015;23(2):303-9. <https://doi.org/10.1016/j.jfda.2014.07.005> PMID:28911386
 33. Kurnia D, Sumiarsa D, Dharsono D, Satari M. Bioactive compounds isolated from Indonesian epiphytic plant of sarang semut and their antibacterial activity against pathogenic oral bacteria. *Nat Prod Commun.* 2017;12(8):1201-4. <https://doi.org/10.1177/1934578X1701200814>
 34. Binartha CT, Suprastiwi E, Kurnia D, Margono A, Artiningsih DA. Antibacterial effects of sarang semut (*Myrmecodia pendans*) fractions using three different solvents toward *Enterococcus faecalis* CPS2. *Int J Appl Pharm.* 2020;12(1):271-5. <https://doi.org/10.22519/ijap.2020.v12s1.iwdr01>
 35. Maulida NF, Yanuaritamala B, Yanuar A, Saputri FC, Mun'im A. Optimization of microwave-assisted extraction to obtain optimum antioxidant activity and anthocyanin concentration from *Myrmecodia pendans* tubers using response surface methodology. *Pharmacog Res.* 2018;10(3):253-7. https://doi.org/10.4103/pr.pr_114_17
 36. Alibasyah ZM, Purba A, Setiabudiawan B, Hendra, Adhita D, Kurnia D, *et al.* The effectiveness of flavonoid and terpenoid isolate sarang semut (*Myrmecodia pendens* Merr. and Perry) against *Porphyromonas gingivalis* ATCC 33277. *Int J Dev.* 2017;7(7):13557-61.
 37. Haris NF, Hasan MK, Wahab IA, Hasan MH, Ponto T, Adam A. Compounds from the antioxidant active fraction of *M. platytyrea*. *J Health Sci.* 2016;14(1):23-9. <https://doi.org/10.17576/jskm-2016-1401-05>
 38. Efendi YN, Hertiani T. Antimicrobial potency of ant-plant extract (*Myrmecodia tuberosa* Jack.) against *Candida albicans*, *Escherichia coli*, and *Staphylococcus aureus*. *J Tradit Med.* 2013;18(1):53-8. <https://doi.org/10.22146/tradmedj.7944>
 39. Apriyanti EA, Satari MH, Laksono B. The antibacterial potential differences between the methanolic extract of ant-plant (*Myrmecodia pendens* Merr. and Perry) tubers and NaOCl towards *Streptococcus mutans* (ATCC 25175). *J Kedokteran Gigi.* 2016;28(2):106-12. <https://doi.org/10.24198/jkg.v29i1.18598>
 40. Kusuma AS, Kusuma SA, Sulistyaniingsih. Antibacterial activity of Papuan ant-nest (*Myrmecodia pendans* L.M. and Perry) ethanol extract against *Shigella dysenteriae*. *Asian J Pharm Clin Res.* 2017;10:46-9. <https://doi.org/10.22159/ajpcr.2017.v10s2.19485>
 41. Attamimi FA, Ruslami R, Maskoen AM. Antibacterial activity test of ant nest Tuber (*Myrmecodia pendens*) crude extract against *Streptococcus sanguinis* compared to chlorhexidine. *Majalah Kedokteran Bandung.* 2017;49(2):94-101. <https://doi.org/10.15395/mkb.v49n2.1053>
 42. Fatriadi F, Kurnia K, Satari MH. Antibacterial activity of ethyl acetate fraction from methanolic extracts of ant-plant tubers towards *Streptococcus sanguis* ATCC 10566. *Padjadjaran J Dent.* 2018;30(3):189-92. <https://doi.org/10.24198/pjd.vol30no3.20002>
 43. Soviati N, Widyarman AS, Binatha CT. The effect ant-nest plant (*Myrmecodia pendans*) extract on *Streptococcus sanguinis* and *Treponema denticola* biofilms. *J Indones Dent Assoc.* 2020;3(1):11-5. <https://doi.org/10.32793/jida.v3i1.401>
 44. Soraya C, Dharsono HD, Aripin D, Satari MH, Kurnia D, Hilmanto D. Effects of sarang semut (*Myrmecodia pendens* Merr. and Perry) extracts on *Enterococcus faecalis* sensitivity. *Dent J.* 2016;49(4):175-80. <https://doi.org/10.20473/j.djmk.v49i4.p175-180>
 45. Alibasyah ZM, Purba A, Setiabudiawan B, Hendra, Adhita D, Kurnia D, *et al.* The efficacy of sarang semut extract (*Myrmecodia pendens* Merr. and Perry) in inhibiting *Porphyromonas gingivalis* biofilm formation. *Dent J.* 2017;50(2):55-60. <https://doi.org/10.20473/j.djmk.v50.i2.p55-60>
 46. Kuswandani F, Satari MH, Maskoen AM. Antimicrobial efficacy of *Myrmecodia pendens* extract and fraction combination against *Enterococcus faecalis* ATCC 29212. *J Dent Indones.* 2019;26(3):119-25. <https://doi.org/10.14693/jdi.v26i3.1085>
 47. Fatmawati D, Puspitasari PK, Yusuf I. Cytotoxic effect of ethanolic extract of sarang semut (*Myrmecodia pendens*) on HeLa cervix cancer cell cine *in vitro* experimental study. *Sains Med.* 2011;3(2):112-20. <https://doi.org/10.26532/sainsmed.v3i2.393>
 48. Achmad A, Chandha MH, Ramadhany S, Handayani H, Samad R. The role of sarang semut (*Myrmecodia pendans*) flavonoid's fraction in proliferation and angiogenesis inhibition of human tongue squamous cell carcinoma. *J Biol Agric Healthc.* 2014;4(21):65-9.
 49. Achmad A, Armyn SA, Supriatno, Singgih MF. Anti-cancer activity and anti-proliferation ant nests flavonoid fraction test (*Myrmecodya pendans*) human tongue cancer cells in SP-C1. *IOSR J Dent Med Sci.* 2014;13(6):1-5.
 50. Achmad A, Supriatno, Singgih MF, Hendrastuti H. Akt signal transduction pathways and Nuclear Factor-kappa B (NF- κ B) transcription as a molecular target of oral tongue squamous cell carcinoma (SP-C1) using Papua's anthill plant (*Myrmecodia*

- pendans*). Pak J Bio Sci. 2016;19(8-9):323-30. <https://doi.org/10.3923/pjbs.2016.323.330>
PMid:29023018
51. Supriatno. Antitumor activity of Papua's *Myrmecodia pendans* in human oral tongue squamous cell carcinoma cell line through induction of cyclin-dependent kinase inhibitor p27Kip1 and suppression of cyclin E. J Cancer Res Ther. 2014;2(3):48-53. <https://doi.org/10.14312/2052-4994.2014-7>
 52. Hasanuddin, Rifayani KS, Supriadi G, Kurnia D, Adhita D. Potential of terpenoid bioactive compound isolated from Papua ant nest as an alternative ovarian cancer treatment. Open J Obstet Gynecol. 2015;5:406-11. <https://doi.org/10.4236/ojog.2015.57058>
 53. Bashari MH, Hidayat S, Ruswandi YA, Putri T, Qomarilla N, Dwiwina RG, et al. The n-hexane fraction of *Myrmecodia pendans* inhibits cell survival and proliferation in colon cancer cell line. Int J Pharm Pharm Sci. 2018;10(1):108-12. <https://doi.org/10.22159/ijpps.2018v10i1.21882>
 54. Achmad A, Horax S, Ramadhany S, Rieuwpassa IE, Sari M, Handayani H, et al. Anti-proliferation activity of ethyl acetate extract from ant nest (*Myrmecodia pendans*) in Burkitt's Lymphoma cancer cells. Pesqui Bras Odontopediatr Clin Integr. 2019;19(1):e4325. <https://doi.org/10.4034/PBOCI.2019.191.29>
 55. Achmad H, Horax S, Ramadhany S, Handayani H, Pratiwi R, Oktawati S, et al. Resistivity of ant nest (*Myrmecodia pendans*) on ethanol fraction Burkitt's Lymphoma cancer cells (*in vitro*) through Interleukin 8 angiogenesis obstacles (II-8). J Int Dent Med Res. 2019;12(2):516-23.
 56. Irfanita N, Eddie S, Aziz NA, Ichwana SJ, Lestari W. Preliminary investigations of anti-cancer properties of *Myrmecodia pendans* on human oral cancer HSC-3 cell lines. Mater Today. 2019;16:2197-2203. <https://doi.org/10.1016/j.matpr.2019.06.110>
 57. Lestari W, Yusry WN, Iskandar SH, Ichwana SJ, Irfan NI, Suriyah WH. Gene expression of selected apoptotic markers in human oral squamous carcinoma HSC-3 cell line treated with *Myrmecodia pendans* plant extract. Makara J Health Res. 2019;23(2):121-6. <https://doi.org/10.7454/msk.v23i2.10632>
 58. Sasmito E, Mulyadi SM, Hertiani, Fathdhieny AQ, Witsqa AA, Laksono YS. Combinational effects of non n-hexane fractions of ant-plant (*Myrmecodia tuberosa* Jack.) hypocotyl with doxorubicin against lymphocyte and cancer cells. Pak J Pharm Sci. 2017;30(5):1987-90.
PMid:29105632
 59. Suharyanto, Purwono B. Extraction method of *Myrmecodia pendans* with maceration techniques top produce an alternative drug for lung cancer. J Pharm. 2015;4(1):19-24. <https://doi.org/10.37013/jf.v4i1.28>
 60. Simanjuntak P, Fanny, Subroto MA. Isolation of active compound from hypocotyl extract of ant nest (*Myrmecodia pendans*) as xanthine oxidase inhibitor. J Ilmu Kefarmasian Indones. 2010;8(1):49-54.
 61. Sudiono J, Hardina M. The effect of *Myrmecodia pendans* ethanol extract on inflamed pulp: study on Sprague Dawley rats. Mol Cell Biomed Sci. 2019;3(2):115-21. <https://doi.org/10.21705/mcbs.v3i2.70>