

Prominent Secondary Metabolites from Selected Genus of Avicennia Leaves

Mohammad Basyuni^{1, 2*}, Didi Nurhadi Illian³, Meighina Atika Istiqomah³, Dini Permata Sari³, Arif Nuryawan^{1, 2}, Poppy Anjelisa Zaitun Hasibuan³, Sumaiyah Sumaiyah³, Etti Sartina Siregar⁴

¹Center of Excellence for Mangroves, Universitas Sumatera Utara, Medan 20155, Indonesia; ²Department of Forestry, Faculty of Forestry, Universitas Sumatera Utara, Medan 20155, Indonesia; ³Department of Pharmacy, Faculty of Mathematics and Natural Science, University of Syiah Kuala, Banda Aceh,23111, Indonesia; ⁴ Faculty of Mathematics and Natural Sciences, Universitas Sumatera Utara, Medan 20155, Indonesia

Abstract

Citation: Basyuni M, Illian DN, Istiqomah MA, Sari DP, Nuryawan A, Hasibuan PAZ, Sumaiyah S, Siregar ES. Prominent Secondary Metabolites from Selected Genus of Avicennia Leaves. Open Access Maced J Med Sci. https://doi.org/10.3889/oamjms.2019.499

Keywords: Physicochemical; Microscopic; Phytochemical; Avicennia spp

*Correspondence: Mohammad Basyuni. Center of Excellence for Mangroves, Universitas Sumatera Utara, Medan 20155, Indonesia; Department of Forestry, Faculty of Forestry, Universitas Sumatera Utara, Medan 20155, Indonesia. E-mail: m.basyuni@usu.ac.id

Received: 25-Sep-2019; Revised: 17-Oct-2019; Accepted: 18-Oct-2019; Online first: 14-Nov-2019

Copyright: © 2019 Mohammad Basyuni, Didi Nurhadi Illian, Meighina Atika Istigomah, Dini Permata Sari, Arif Nuryawan, Poppy Anjelisa Zaitun Hasibuan, Sumaiyah Sumaiyah, Etti Sartina Siregar. This is an open-access article distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 International License (CC BY-NC 4.0)

Funding: This research did not receive any financial support

Competing Interests: The authors have declared that no competing interests exist

BACKGROUND: Mangrove plants distributed in the intertidal of the tropical and subtropical region including in North Sumatra, Indonesia. The production of secondary metabolite compounds is well known to mangroves. Characterisation of prominent compounds from mangrove plants such as genus of *Avicennia* is required to explore for their biological and pharmacological properties of these compounds.

AIM: The purpose of this research was to analyse the prominent secondary metabolites through the characterisation of phytochemical, physicochemical, and microscopic of the mangrove genus Avicennia leaves, particularly *Avicennia alba*, *A. lanata*, *A. marina*, and *A. officinalis*.

METHODS: Phytochemical screening was carried out on *Avicennia* spp leaves to the established process. Physicochemical characters of mangrove leaves were investigated by simplicial powder consisting of moisture content, water-soluble, ethanol-soluble, ash content and ash soluble acid according to the WHO formula. Microscopic analysis on the simplicial powder was carried out based on the WHO procedure.

RESULTS: The result showed that physicochemical feature displays diversity among the species and important findings on the water concentration was less than 10% as a prerequisite for the drug. The phytochemical search of simplified grain also depicted divergence among the species, only alkaloid, saponin, and triterpenoid or phytosterol were found entirely in *Avicennia* spp leaves. Microscopic search found a similar type of stoma in Avicennia spp leaves, namely diacytic.

CONCLUSION: The prominent secondary metabolites in *Avicennia* spp leaves consisting of alkaloid and saponin in simplicial and triterpenoid/sterol was either in simplicial or hexane extract. The present study may provide significant pharmacological properties from mangrove *Avicennia* genus green foliages, which could accelerate another prospect for non-wood mangrove utilisation.

Introduction

Mangrove plants widespread in the intertidal of the tropical and subtropical region have numerous activities as medicinal plants, and only a few have been exposed [1]. Mangroves are famous for yielding secondary metabolite compounds mainly from isoprenoid compound groups includina polyisoprenoids. The polyisoprenoids comprises of two families, i.e., polyprenol and dolichol. Polyprenol is recognised to have some pharmacological property such as anticancer [2], antidyslipidemic [3], antiinfluenza, and antiviral activity [4].

The composition and diversity of polyisoprenoids compounds in mangrove forests of Iriomote Island, Japan, and North Sumatra, Indonesia have been reported [5], [6], [7]. Mangrove plants, including mangrove associates or coastal plants, are an excellent source of secondary metabolites largely derived from isoprenoid alcohol [8]. In virtually all living organisms, polyisoprenoid alcohols undeviating five-unit polymers. Secondary metabolites are valued and exploited for their phytochemical and other

activities in their interaction with the environment [9].

Isoprenoids are regarded to b necessary as an active biological compound for medicinal properties their widespread biological due to and pharmacological properties [10], and mangrove plants have long been having been in traditional medicine for a long time [11]. Therefore, this study aims to analyse prominent secondary metabolites through the phytochemical, physicochemical, and microscopic of a particular genus of Avicennia leaves, namely Avicennia alba, A. lanata, A. marina, and A. officinalis.

Material and Methods

Avicennia alba, A. lanata, A. marina, and A. officinalis (Acanthaceae) was compiled from a mangrove community at Lubuk Kertang, Langkat, North Sumatra, Indonesia. The species of plants are determined at the Bogor Research Center for Biology, the Indonesian Institute of Science (LIPI). There the specimen vouchers were deposited. Avicennia leaves were compiled, wet sorting was done to separate dirt material, washed under tap water, drained and weighed. The mangrove leaves are then dried in a drying cabinet; dry sorting was performed to separate unwanted parts of plants and other impurities that remain in dry Simplicia, then weighed and stored in tightly closed plastic containers [12], [13].

Each 200 g of Simplicia powder from Avicennia leaves was dissolved with 2000 mL of a mixture of chloroform: methanol (2: 1, v / v) for 48 hours. Lipid extract from leaves was saponified at a temperature of 65°C for 24 hours in 86% ethanol comprising KOH 2 M. The non-saponified portion was then dissolved with n-hexane, then the solvent was evaporated. Then re-dissolve in n-hexane, pour or filter. Concentration was carried out using a rotary evaporator at 40°C. Then dried as a concentrated extract was obtained [5], [6], [7]. Avicennia leaves' physicochemical parameters were analysed by simplified powder involving moisture content, watersoluble, ethanol-soluble, ash content, and the essay extract was implemented by the WHO standard [12] and earlier works [13], [14].

On Avicennia green foliages to the standard process as described above were performed, a phytochemical method is simplified and hexane extract [12]. As previously reported, phytochemical analysis consisting of secondary metabolites, i.e. alkaloids, flavonoids, glycosides, saponins, tannins, triterpenoids/sterols [13], [14].

Microscopic studies on the simplification of true Avicennia spp leaves mangroves were conducted based on the WHO technique [12]. Specifically, a microscopic examination of the simplified powder was performed by robust chloral hydrate solution on the slide, scattered with simplified powder, then protected by a covering glass and detected under a microscope.

Results

Table 1 recapitulated the physicochemical of *Avicennia* spp photosynthetic tissues. The characterisation in this observation led to simplicial of *A. alba, A. lanata, A. marina,* and *A. officinalis* moisture content was 7.95%, 7.52%, 7.74%, and 7.23% respectively. The lower water content of usually incorporated to the necessity for drug progress was less than 10%. The water concentration enclosed \geq 10% can occur the process of material degradation and damage, could not be stored for a long time.

Table 1: Physicochemical characteristic of Avicennia spp leaves

No	Species	Water	Water-	Ethanol	Total ash (%)	Ash insoluble
		content (%)	soluble (%)	soluble (%)		acid (%)
1	A. alba	7.95	28.16	17.59	10.69	1.93
2	A. lanata	7.52	28.57	18.84	9.23	1.54
3	A. marina	7.74	25.78	18.45	7.12	1.91
4	A. officinalis	7.23	26.35	18.98	8.65	1.78

Phytochemical analysis of simplified powder and n-hexane extracts of preferred *Avicennia* genus mangrove leaves, *A. alba*, *A. lanata*, *A. marina*, and *A. officinalis* performed to collect secondary metabolite group data. Table 2 depicted the results of simplicial grain on the phytochemical screening and *Avicennia* spp n-hexane extract. The results of the grouping are then applied as an advance prominence to classify which types of compounds operate in contradiction of phytopharmacological activities.

Table 2: Phytochemical screening in simplicial and hexane extract Avicennia spp

١o.	Compound	Solvent	Mangrove species	Simplicial	Hexane
	Alkaloids	Dragendorff	A. alba	+	-
		Bouchardat	A. lanata	+	-
		Mayer	A. marina	+	-
			A. officinalis	+	-
2.	Flavonoids	Zn + HCl	A. alba	-	-
			A. lanata	+	-
			A. marina	+	-
			A. officinalis	-	-
3.	Glycosides	Molisch	A. alba	-	-
			A. lanata	+	-
			A. marina	+	-
			A. officinalis	-	-
4.	Saponin	Hot water + HCL	A. alba	+	-
			A. lanata	+	-
			A. marina	+	-
			A. officinalis	+	-
5.	Tannins	FeCl ₃ 1%	A. alba	-	-
			A. lanata	+	-
			A. marina	+	-
			A. officinalis	-	-
6.	Triterpenoid/	Liebermann-	A. alba	+	+
	Steroid	Burchard	A. lanata	+	+
			A. marina	+	+
			A. officinalis	+	+

Where (+): contain the compound; (-): not contain.

Simplicial powder phytochemical investigation reflects the diversity among *Avicennia* species; for

example, alkaloid and saponin were detected in *Avicennia* spp. The phytochemical investigation in hexane extract, on the other hand, did not show any alkaloids, flavonoids, glycosides, saponin, and tannin, but only triterpenoid or phytosterol found in *Avicennia* spp. Phytochemical tests showed that flavonoids, glycosides, and tannins were part of the simplified powder, as summarised in Table 2. Adding Molisch reagent and absorbed sulfuric acid created a purple ring that showed glycoside compounds (Table 2).

Table 3: Isoprenoid and polyisoprenoid pattern in Avicennia spp

Species	Tissue	Isoprenoid (%)		Polyisoprenoid (%)		Ref	
Species		Triterpenoid	Phytosterol	Polyprenol	Dolichol		
A. alba	Leaves	70.1	29.9	nd	100	[5], [15]	
A. lanata	Leaves	ni	ni	nd	100	[6]	
A. marina	Leaves	91.0	9.0	4.2	95.8	[5], [16]	
A. officinalis	Leaves	61.5	38.5	nd	100	[6], [15]	
A. alba	Roots	47.4	52.6	2.2	97.8	[5], [15]	
A. lanata	Roots	ni	ni	nd	100	[6]	
A. marina	Roots	57.8	42.2	nd	100	[5], [16]	
A. officinalis	Roots	100	nd	7.7	92.3	[6], [14]	
Where, nd = not detected; ni = no information.							

While Dragendorf, Bouchardat, and Mayer reagent solutions add definite findings as *Avicennia* spp (Table 2), the highest percentage of isoprenoid and polyisoprenoid secondary metabolites for *Avicennia* leaves and roots was shown in Table 3.

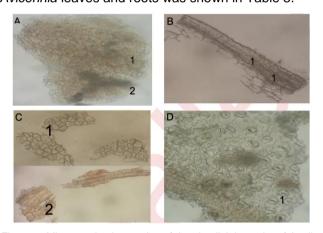


Figure 1: Microscopic observation of the simplicial powder of A. alba leaves; A) which 1. Diacytic type stomata, 2. Prismatic oxalate crystal; B) Simplicial crumb of A. lanata leaves, where 1. Prismatic oxalate crystal; C) Simplicial grain of A. marina leaves, where 1. Diacytic type stomata, 2. Parenchyma tissue; D) Simplicial powder of A. officinalis leaves, where 1. Diacytic type stomata

In the leaves, triterpenoid, and dolichol as well as dominated over phytosterol and polyprenol, respectively (Table 3). By contrast, in the roots, the isoprenoid composition showed variation, only in *A. officinalis* triterpenoid predominated over phytosterol. It is noteworthy that the occurrence of both triterpenoid and phytosterol were detected in *A. alba* and *A. marina* roots (Table 3). Microscopic search detected a similar type of stoma in *Avicennia* spp leaves, namely diacytic type stomata as depicted in Figure 1. In this perspective, a microscopic method is one of the simplest and cheapest ways to study for clarification of the source materials of medicinal plants such as in mangrove plants.

Discussion

Characterisation of Avicennia genera's phytochemical investigation enabled us to analyse the content of ethanol-soluble specifies the constituents existing in ethanol solvents that are considered to be the polar and non-polar signal. Analysis of the water and ethanol soluble content was done to show the number of water and ethanol soluble polar compounds. This result suggested the occurrence of more dissolved constituents in water solvents. To destroy organic compounds and their derivatives, ash content is determined [13], [14]. To define levels of inorganic compounds that are not soluble in acids such as silica gel, the ash content is not acid-soluble. The high total ash concentration describes the presence of inorganic metal elements, for instance, lead, calcium, iron, magnesium, which may be partly caused by contamination [17].

The improvement of solid foam with shaky water and the addition of 2N HCl did not lose an indication that saponin compounds exist [13]. Adding Mg powder and concentrated hydrochloric acid produces a red solution, and with amyl alcohol quantity, the red colour is involved in the amyl alcohol layer indicating the occurrence of flavonoids [13]. FeCl3's addition gave a blackish-blue colour that depicts tannin compounds [18]. Adding LB reagents created blue-green, violet and red-violet colors that specified triterpenoid/steroid compounds [15], [16], [19].

In line with this finding, the polyprenols in the root tissues were also dominated by dolichols [5], [7]. Polyisoprenoids have been tested *in vitro* to be more suitable as an antimicrobial agent and an oxidant in associated mangrove leaves and roots [20], consisting mostly of dolichols. Therefore, these results supported the previous view that polyisoprenoid composition in mangroves was not polyprenols but dolichols.

Microscopic search detected a similar type of stoma in *Avicennia* spp leaves, namely diacytic type stomata. The microscopic study is a vegetation type indicator [21]. In this perspective, one of the simplest and cheapest ways to study a microscopic method is by establishing to clarify the source materials of medicinal plants such as in mangrove plants. Thus, the phytochemical pattern, including microscopic information, will help in standardisation of quality, purity and sample identification.

References

1. Prasad N, Yang B, Kong KW, Khoo HE, Sun J, Azlan A, et al. Phytochemicals and antioxidant capacity from Nypa fruticans Wurmb fruit. Evidence-Based Complementary and Alternative Medicine. 2013; 2013:1-9. <u>https://doi.org/10.1155/2013/154606</u> PMid:23710209 PMCid:PMC3654328

2. Kuznecovs S, Jegina K, Kuznecovs I. Inhibition of P-glycoprotein by polyprenol in human breast cancer cells. The Breast. 2007; 16:515-21. https://doi.org/10.1016/S0960-9776(07)70074-7

3. Singh G, Gupta P, Rawat P, Puri A, Bhatia G, Maurya R. Antidyslipidemic activity of polyprenol from Coccinia grandis in high-fat diet fed hamster model. Phytomedicine. 2007; 14(12):792-8. https://doi.org/10.1016/j.phymed.2007.06.008 PMid:17689941

4. Safatov AS, Boldyrev AN, Bulychev LE, Buryak GA, Kukina TP, Poryvaev VD, et al. Aprototype prophylactic anti-influenza preparation in aerosol form on the basis of Abies sibirica polyprenols. Journal of Aerosol Medicine. 2005; 18(1):55-62. https://doi.org/10.1089/jam.2005.18.55 PMid:15741774

5. Basyuni M, Sagami H, Baba S, Iwasaki H, Oku H. Diversity of polyisoprenoid in ten Okinawan mangroves. Dendrobiology. 2016; 75:167-75. https://doi.org/10.12657/denbio.075.016

6. Basyuni M, Sagami H, Baba S, Oku H. Distribution and occurrence of new polyprenyl acetone and other polyisoprenoids in Indonesian mangroves. Dendrobiology. 2017; 78:18-31. https://doi.org/10.12657/denbio.078.003

7. Basyuni M, Wati R, Sagami H, Sumardi, Baba S, Oku H. Diversity and abundance of polyisoprenoid composition in coastal plant species from North Sumatra, Indonesia. Biodiversitas. 2018; 19:1-11. <u>https://doi.org/10.13057/biodiv/d190101</u>

8. Basyuni M, Sagami H, Baba S, Putri LA, Wati R, Oku H. Salinity alters the polyisoprenoid alcohol content and composition of both salt-secreting and non-salt-secreting mangrove seedlings. HAYATI J Biosci. 2017; 24:206-14. https://doi.org/10.1016/j.hjb.2017.11.006

9. Atanasov AG, Waltenberger B, Pferschy-Wenzig EM, Linder T, Wawrosch C, Uhrin P, Temml V, Wang L, Schwaiger S, Heiss EH, Rollinger JM. Discovery and resupply of pharmacologically active plant-derived natural products: A review. Biotechnol Advances. 2015; 33:1582-1614.

https://doi.org/10.1016/j.biotechadv.2015.08.001 PMCid:PMC4748402

10. Ksouri R, Ksouri WM, Jallali I, Debez A, Magné C, Hiroko I, Abdelly C. Medicinal halophytes: potent source of health-promoting biomolecules with medical, nutraceutical and food applications. Critical Review in Biotechnology. 2012; 32:289-326. https://doi.org/10.3109/07388551.2011.630647 PMid:22129270

11. Bandaranayake WM. Traditional and medicinal uses of

mangroves. Mangroves and Salt Marshes. 1998; 2:133-48. https://doi.org/10.1023/A:1009988607044

12. WHO (World Health Organization). Quality Control Methods for Medicinal Plant Materials: Geneva, 1998:28-31.

13. Basyuni M, Ginting PY, Lesmana I. Phytochemical analysis of binahong (Anredera cordifolia) leaves extract to inhibit in vitro growth of Aeromonas hydrophila. AIP Conference Proceedings. 2017; 1904:020072. <u>https://doi.org/10.1063/1.5011929</u>

14. Basyuni M, Amri N, Putri LA, Syahputra I, Arifiyanto D. Characteristics of fresh fruit bunch yield and the physicochemical qualities of palm oil during storage in North Sumatra. Indonesian Journal of Chemistry. 2017; 17:182-90. https://doi.org/10.22146/ijc.24910

15. Basyuni M, Putri LA, Nurainun H, Oku H. Non-saponifiable lipid composition of four salt-secretor and non-secretor mangrove species from North Sumatra, Indonesia. Makara. Journal of Science. 2012; 16:89-94. <u>https://doi.org/10.7454/mss.v16i2.1402</u>

16. Basyuni M, Oku H, Baba S, Takara K, Iwasaki H. Isoprenoids of Okinawan mangroves as lipid input into estuarine ecosystem. Journal of Oceanography. 2007; 63(4):601-8. https://doi.org/10.1007/s10872-007-0053-2

17. Wuana RA, Okieimen FE. Heavy metals in contaminated soils: a review of sources, chemistry, risks and best available strategies for remediation. ISRN Ecology. 2011; 24:402647. https://doi.org/10.5402/2011/402647

18. Naz R, Bano A. Antimicrobial potential of Ricinus communis leaf extracts in different solvents against pathogenic bacterial and fungal strains. Asian Pacific Journal of Tropical Biomedicine. 2012; 2:944-7. <u>https://doi.org/10.1016/S2221-1691(13)60004-0</u>

19. Basyuni M, Wati R. Bioinformatics analysis of the oxidosqualene gene and the amino acid sequence in mangrove plants. Journal of Physics: Conference Series. 2017; 801:012011. https://doi.org/10.1088/1742-6596/801/1/012011

20. Sumardi S, Basyuni M, Wati R. Antimicrobial activity of polyisoprenoids of sixteen mangrove species from North Sumatra, Indonesia. Biodiversitas Journal of Biological Diversity. 2018; 19(4):1243-8. https://doi.org/10.13057/biodiv/d190409

21. Umbanhowar Jr CE, Mcgrath MJ. Experimental production and analysis of microscopic charcoal from wood, leaves, and grasses. The Holocene. 1998; 8:341-6. https://doi.org/10.1191/095968398666496051